



- 2 Multifunctional nanocarriers for lung drug delivery
- **3** Jorge F. Pontes ^{1,2} and Ana Grenha ^{1,2,3,*}
- Centre for Marine Sciences (CCMAR), University of Algarve, Campus de Gambelas, 8005-139 Faro, Portugal;
 pontes.jorge21@gmail.com
- ⁶ ² Drug Delivery Laboratory, Centre for Biomedical Research (CBMR), University of Algarve, Campus de
 Gambelas, 8005-139, Faro, Portugal
- B ³ Department of Chemistry and Pharmacy, Faculty of Sciences and Technology, University of Algarve,
 Campus de Gambelas, 8005-139 Faro, Portugal
- 10 * Correspondence: amgrenha@ualg.pt; Tel.: +351-289-244-441; Fax: +351-289-800-066
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12 Abstract: Nanocarriers have been increasingly proposed for lung drug delivery applications. The 13 strategy of combining the intrinsic and more general advantages of the nanostructures with 14 specificities that improve the therapeutic outcomes of particular clinical situations is frequent. These 15 include surface engineering of the carriers, by means of alteration of the materials structure (i.e. 16 chemical modifications), the addition of specific ligands so that predefined targets are reached, or 17 even the tuning of the carrier properties to respond to specific stimuli. The devised strategies are 18 mainly directed to three distinct areas of lung drug delivery, encompassing the delivery of proteins 19 and protein-based materials, either for local or systemic application, the delivery of antibiotics and 20 the delivery of anticancer drugs, the latter two comprising local delivery approaches. This review 21 addresses the applications of nanocarriers aimed at lung drug delivery of active biological and 22 pharmaceutical ingredients, focusing with particular interest nanocarriers that exhibit 23 multifunctional properties. A final section addresses the expectations regarding the future use of 24 nanocarriers in the area.

Keywords: Antibiotics, cancer, drug delivery, lung delivery, nanocarriers, nanopharmaceuticals,
 proteins.

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Review

28 **1. Introduction**

29 The appearance of new therapies and alternative strategies for the delivery of drug molecules 30 has been changing the paradigm of therapeutic approaches [1-3]. Indeed, therapeutic solutions have 31 been implemented around one of two possible objectives: one is the development of better and more 32 effective therapies, usually involving new drugs; the other relies on exploring different ways to 33 deliver molecules, potentiating their action and, in many cases, simultaneously, eliminating adverse 34 effects or, at least, decreasing their impact. The latter approach has been often used for drug 35 repurposing, finding new applications for de-risked compounds, with potentially lower overall 36 development costs and shorter development timelines. The literature displays some recent and 37 valuable reviews on the topic of drug repurposing [4,5]. The adverse effects derived from 38 pharmaceuticals have always caused concern, as some can be devastating, leading to therapeutic non-39 compliance. Thus, exploring delivery strategies is as important as the discovery of new molecules 40 and targets, providing the molecules with specific orientation towards their targets, avoiding major 41 biological stresses and, overall, improving the therapeutic quality [6]. Actually, in many cases, the 42 two referred approaches are addressed at a time.

43 Considering that, in most cases, the delivery of unformulated drug molecules is not successful, 44 formulation plays a role of utmost importance in therapy. Conventional drug delivery systems 45 encompass numerous restrictions, which include limited targeting, low therapeutic index, poor 46 aqueous solubility, and the potentiation of drug resistance [7]. The design and production of systems 47 in which drug molecules are included in a carrier, being either embedded in the matrix or adsorbed 48 to the surface, is frequently the next step towards a more effective therapy. The reasons justifying the 49 need for drug formulation are in the annals of pharmaceutical technology, going from a simple 50 protection of drugs to the more complex targeting of cells or tissues. In between, the need to achieve 51 control over drug release has also been a hallmark of drug delivery research. A useful historical 52 perpective on the generations of controlled drug delivery systems is available in [8]. In fact, in an era 53 where drug molecules are expected to answer to increasingly complex environments, their 54 formulation takes on a role never seen. That role assumed, a great variety of advanced drug delivery 55 systems has been reported through the years, with important variations on their properties, including 56 size and composition. Size is one of the most relevant features in the field. In this context, both micro-57 and nanocarriers have been reported to be viable approaches, the final selection being objectively 58 dependent on the specific application that is envisaged. The potential of micron-sized carriers has 59 been highlighted for different applications [9-13], but lies out of the scope of this review. As for the 60 nanoscaled carriers, in drug delivery these fall into the designation of nanopharmaceuticals, defined 61 by Rivera Gil et al as "pharmaceuticals where the nanomaterial plays the pivotal therapeutic role or 62 adds additional functionality to the previous compound" [14]. The International Organization for 63 Standardization defines nanoparticles as those having at least one dimension less than 100 nm [15]. 64 In turn, the American Food and Drug Administration (FDA) indicates that products involve 65 nanotechnology, and should therefore be evaluated as such, when they are "engineered to exhibit 66 properties or phenomena attributable to dimensions up to 1000 nm" [16]. This broader definition is 67 the most typically seen in academic research in drug delivery and will be adopted in this review. 68 Therefore, all submicron systems will be considered nanocarriers.

69 As well indicated in the historical description of [8], after an initial period back in the 1980s and 70 1990s, where many micron-sized formulations became popular and reached the market, 71 nanotechnology has been leading the interest of drug delivery scientists since the turn of the new 72 century. In fact, the first mention to a system capable of encapsulating a molecule and providing its 73 transport through a membrane dates back to 1965, under the name of liposome [17], which is, indeed, 74 a nanosystem. From that point on, many other nanoformulations were described and explored, some 75 of them consisting in particulate-based systems, with significant structural differences comparing 76 with the vesicle-based systems comprised by liposomes. Particulate carriers at the nanoscale include 77 polymeric nanoparticles [18], solid lipid nanoparticles [19,20], nanostructured lipid carriers [21,22] 78 and magnetic and silica nanoparticles [23,24]. Probably, issues like increased stability [6], the closer 79 interaction with cell structures [25], the propensity to provide increased drug absorption [26] and the 80 great ability for surface functionalisation [27] have driven the higher popularity of these materials. 81 Regrettably, the clinical translation of nanoparticulate-based systems is very limited so far and only 82 one formulation is available in the market: Abraxane[®], marketed since 2005 [28]. This comprises 83 albumin-conjugated paclitaxel, being used in metastatic breast cancer and non-small-cell lung cancer 84 [29]. In this manner, despite the extensive research on particulate-based nanopharmaceuticals, their 85 market absence is notorious, mainly due to tightened regulations. Even so, there are many 86 nanoformulations currently undergoing clinical trials focusing varied routes of administration, and 87 it is expected that some of them make their way into the market in some years [30].

88 From the referred formulations undergoing a translational process and to the knowledge of the 89 authors of this review, none is directed to lung drug delivery. Nevertheless, this is a delivery route 90 that has been gaining popularity in recent years, essentially owing to its non-invasiveness and the 91 increased demonstration of its potential, not only for local therapies, but also to provide systemic 92 action. Actually, according to the World Health Organization (WHO), chronic obstructive pulmonary 93 disease (COPD), lower respiratory infections and lung cancer are, respectively, the third, fourth and 94 sixth causes of death worldwide [31], which illustrates the existing therapeutic limitations. In 95 addition, numerous other respiratory disorders are characterised by an urgent and unmet therapeutic 96 need. The myriad of routes of administration poses the question of which one is the most adequate

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to deliver a drug intended to treat a specific disease. In parallel, the search for routes of administration other than the oral has been increasing for some years. The lung is now being taken into high consideration for this purpose, as clearly demonstrated in Figure 1, where the number of publications per year that specifically refer to "lung drug delivery", as retrieved from ISI Web of Science, can be observed. It has been considered a viable alternative in the delivery of drugs and the popularity of this delivery route is reflected in the growing number of publications, especially from 2014 onwards, indicating a clear interest from the scientific community.



119Figure 1 – Number of scientific publications under the topics of "lung drug delivery" (blue) and "lung120drug delivery and nano" (orange) on ISI Web of Science, as function of the publication year (last121updated in January 2020).

122 The established popularity of the lung route relies on several advantages and specific features. 123 Apart from the already mentioned ability to provide either local or systemic effect, characteristics 124 such as high vascularisation and extensive area available for absorption are highly appealing for 125 systemic delivery, while the low metabolic activity compared with the oral route serves both 126 modalities [32]. Furthermore, the possibility to use lower doses and the low incidence of systemic 127 side effects are relevant pros for local delivery [33]. A very useful and up-to-date review on the 128 challenges and opportunities of lung delivery can be found in [33]. Despite the mentioned 129 advantages, some limitations are also to be referred, which mainly include the mucociliary clearance 130 as the main mechanism of defence, the patient variability on pathophysiological aspects of the organ 131 and the need to endow the drugs with suitable aerodynamic properties to reach a specific area of the 132 lung [34]. Regarding the latter aspect, the aerodynamic diameter of the drugs or carriers to be 133 delivered through inhalation assumes a crucial role. The aerodynamic diameter is the diameter of a 134 spherical particle with density of 1 g/cm³ and the same settling velocity as the particle of interest. In 135 this context, it is reported that the smaller airways can be reached by particles with aerodynamic 136 diameter lower than 5 μ m, while those with less than 2 μ m may arrive to the respiratory zone, which 137 includes the alveoli [32].

138 Drugs have been administered by inhalation for millennia, but inhaled therapeutics have been 139 used predominantly to manage common pulmonary diseases like asthma and COPD. In these areas, 140 inhalable drugs have been dominating the market. Systemic formulations, in turn, have been facing 141 many limitations, with significant technical hurdles requiring being addressed before success is 142 achieved. Nevertheless, it has become consensual that, given the offered advantages, the posed 143 challenges are worth addressing. The possiblities have long been debated, especially considering the 144 emergence of biological drugs that are degraded in the gastrointestinal tract and, so, rely uniquely 145 on injection to find efficacy. The scientific community has, thus, been recognising the potential of the 146 lung to be used as a systemic pathway, and many of the papers contributing to Figure 1 deal with 147 systemic lung delivery, although so far this interest is not mirrored by the market. In fact, inhalable

148 insulin is one of the exceptions to mention, appearing first as Exubera®, from Pfizer (2006), but being 149 discontinued one year after approval [35,36], the company justifying the withdrawal with 'comercial 150 reasons'. Another product of inhalable insulin became available in 2014, as Afrezza®, from Mannkind 151 Corporation, and incorporates the Technosphere® technology [36,37]. These inhalable insulin 152 products are not based on nanotechnologies, but the scientific community has been recognising the 153 potential of nanocarriers in lung delivery and nanoformulations have been increasingly proposed, as 154 can be also observed in Figure 1. An integrated analysis of this figure shows that publications 155 involving nanosystems usually comprise more than half of the total number of publications on the 156 topic of lung delivery, which demonstrates well their popularity. In fact, the superiority of 157 nanosystems has been demonstrated in certain applications of the respiratory field, as will be 158 described in the following sections of the review. The nanocarriers permit drug protection, provide 159 a greater ability to interact with the tissues and cells, owing to the high surface area, often allowing 160 specific targeting and/or controlled drug release [38]. However, the proposal of nanocarriers must 161 not be blind and it is important to note that some applications may take greater benefit from the use 162 of microcarriers, for example if the therapeutic target is phagocytic cells such as macrophages. 163 Moreover, despite the large amount of works describing nanocarriers for lung delivery applications, 164 it is worth saying that a closer reading of the searched documents reveals that many of the works 165 propose the nanocarriers as having potential from a conceptual point of view, but a much smaller 166 amount approaches the practical concept of preparing the carriers for inhalation, endowing them 167 with the required properties, namely aerodynamic, for the purpose.

168 While initial approaches on the development of drug nanocarriers essentially addressed issues 169 of drug stability and control over the release, these were rapidly replaced or completed with 170 advanced techniques of particle engineering. Thus, the proposal of more complex carriers naturally 171 came along, with particle engineering techniques endowing the nanocarriers with specific properties 172 well beyond their role of carrying a drug or molecule of interest. Such carriers were named as 173 multifunctional and their applications have been explored in all areas of delivery. The 174 multifunctional nanocarrier can be one composed of a material that provides, itself, a specific 175 function, or one that was modified to exhibit a determined feature. Lung delivery can strongly benefit 176 from the features of these carriers. In fact, while most asthma and COPD drugs are delivered to the 177 lung with relatively low efficiency and still ensure therapeutic efficacy, drugs aimed at a systemic 178 action or used to treat orphan diseases or cancer, require optimisation of delivery efficiency. This will 179 render the treatment cost-effective, while potentiating clinical effectiveness and minimising side 180 effects.

181 Considering the interest of nanocarriers within the context of lung drug delivery, this review 182 will focus on their applications, placing particular emphasis on the functionality that is provided by 183 the proper carriers. For the effects of the review, only works addressing directly the issue of 184 pulmonary administration of the carriers, either by adequate in vitro testing or by suitable in vivo 185 delivery, will be considered, thus going beyond the theoretical concept of suitability for lung delivery 186 purposes. The specific features of the carriers will be referred and the achieved outcomes described. 187 The envisaged applications of the nanocarriers in lung delivery are diverse, but particularly address 188 the delivery of proteins or protein-based materials, either for local or systemic effect [39,40], cancer 189 treatment [41,42] and local delivery of antibiotics [43,44]. Therefore, the review will specifically focus 190 on these topics. As an introductory element to the following sections, Table 1 depicts the major 191 respiratory diseases, along with their main limitations and the potential improvements imparted by 192 pulmonary delivered therapy.

194Table 1 – General overview of the major respiratory diseases, along with their main limitations and improvements imparted by pulmonary delivery of the195drugs. Indication of the application, in each disease, of the drug classes addressed in the review.

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Respiratory disease	Main limitations	Improvements from lung delivery	Proteins	Antibiotics	Anticancer drugs
Asthma	Low therapeutic efficacy of delivered drugs; inefficient control of the disease; airway inflammation	n.a.*	х		
COPD	Persistent inflammation; parenchymal lung tissue destruction; abnormalities of the small airways	n.a.*	x	x	
Pneumonia	Low amount of drug reaches infection site; antimicrobial resistance	Higher drug accumulation in infection site; co- localisation of drug and infectious agent	x	x	
Cystic fibrosis	Thick viscous mucus; recurrent lung infections; progressive impairment of lung airways	Mucus-penetrating carriers; increased lung drug retention; delivery of genetic material to restore CFTR function	х	x	
Tuberculosis	Reduced amount of drug reaches infection site; antimicrobial resistance; long therapeutic regimen	Co-localisation of drug and infectious agent; reduction of antibiotic resistance incidence; possibility of add-on therapy (along with oral); reduce treatment duration		x	
Lung cancer	Non-specificity of drugs; difficulties to reach the affected tissues; severity of systemic adverse effects	Vectorisation to cancer cells; reduction of systemic adverse effects			x
CFTR: Cystic fibrosis transmembrane conductance regulator; COPD: chronic obstructive pulmonary disease					

n.a.: not applicable; *conventional therapy already administered via inhalation

199 2. Lung drug delivery mediated by multifunctional nanocarriers

Multifunctional nanocarriers can be produced from a wide range of materials. In parallel, it is also wide the number of molecules that can be associated to the carriers to provide specific effects and improve their perfomance, either being adsorbed or chemically-bound. This variety arises from the necessity to meet different challenges and address a vast number of diseases with intrinsic different characteristics.

In the present section, the three main topics mentioned above will be approached. As a summary of the contents, Figure 2 provides a depiction of the type of carriers used in each topic, along with the materials selected for nanoparticle matrix and surface modification, when applicable, and also the

associated molecules of interest.



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210 Figure 2 – General overview of the types of nanocarriers used in the delivery of proteins, 211 antibiotics and anticancer drugs, along with the materials applied in the carrier matrix (inside the 212 circle), the ligands used for surface functionalisation and the associated molecules of interest (the 213 cargo). The circle indicates the carrier. BSA: bovine serum albumin, EpCAM: epithelial cell adhesion 214 molecule, HSA: human serum albumin, IgG: immunoglobulin G, NP: nanoparticles, PEG: 215 polyethylene glycol, PEI: polyamidoamine, PGA-co-PDL: poly(glycerol adipate-co-ω-216 pentadecalactone), PLA: polylactic acid, PLGA: polylactic-co-glycolic acid, RGD: tripeptide Arg-Gly-217 Asp, siRNA: small interfering RNA, SLN: solid lipid nanoparticles, SPIO: superparamagnetic iron 218 oxide, TRAIL: tumor necrosis factor-related apoptosis-inducing ligand, "copolymer based on 219 methoxy poly(ethylene glycol)-poly(ethylenimine)-poly(L-glutamate), *lipids: glyceryl 220 monostearate, cholesterol.

221 2.1. Delivery of proteins and protein-based materials

The marked biotechnological advances observed in recent decades resulted in the appearance of many protein-based drugs. Fundamentally, the oral delivery of these molecules is prevented by the degrading effect of abundant protease content and the possibilities of delivery are essentially

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225 reduced to injection-based strategies. However, this approach is more expensive and not appreciated 226 by the patients, mainly due to the discomfort associated with the administration, but also because of 227 some issues related with aesthetics, including bruising and skin marks that may compromise 228 therapeutic compliance [45]. The pulmonary route thus appears as a sound alternative when a 229 systemic effect is desired, but protein-based drugs also find applications in the treatment of local lung 230 diseases. In fact, the first inhaled protein reaching the market was recombinant human DNase 231 (rhDNase), indicated in the treatment of cystic fibrosis and available since the late 90s. Inhalable 232 insulin appeared approximately 10 years after and lessons learnt from its development resulted in 233 the current availability of many elegant inhalation devices and formulations. So far, no other inhaled 234 biological drug aimed at systemic delivery reached the market, despite those being the drugs 235 focusing most of the attention within the context of systemic delivery mediated by inhalation. None 236 of the referred marketed formulations encompasses the use of nanocarriers. Actually, members of 237 our group participated in the first work proposing the inhalation of insulin encapsulated in polymeric 238 nanoparticles, dated back to 2005 [39,46,47]. At that time, chitosan was proposed as matrix material, 239 resulting in non-toxic nanoparticles and endowing the system with mucoadhesivity [48]. In order to 240 provide the nanoparticles with suitable aerodynamic properties to reach the alveolar zone, a nano-241 in-micro system was developed, using spray-drying to microencapsulate the nanoparticles in 242 mannitol microspheres. These, expectedly released the nanoparticles after dissolving in the lung 243 lining fuid, providing the release of the protein that could, thus, be absorbed systemically [39]. An in 244 vivo study in rats evidenced that microencapsulated insulin-loaded chitosan nanoparticles 245 administered intratracheally (IT, 16.7 IU/kg) induced a more pronounced and prolonged 246 hypoglycemic effect compared with insulin solution, as observed in Figure 3 [47], thus demonstrating 247 the contribute of the carrier itself to the observed therapeutic effect. A similar approach was later on 248 proposed by other authors, using poly-L-lysine to modify the surface of self-assembled pure insulin 249 nanoaggregates, benefiting from the adhesive properties of poly-L-lysine. After IT administration to 250 diabetic rats, the modified nanostructures (5 IU/kg) induced hypoglycemic effect as stronger as 251 subcutaneous delivery (1 IU/kg), but increasing the drug half-life from 1.28 h to 2.75 h. Although not 252 statistically significant, the hypoglycemic effect obtained from nanoparticles was also more 253 prolonged, achieving 23.4% relative bioavailability [49]. 254



268 Figure 3 - Hypoglycemic profiles following intratracheal administration to rats of 269 microencapsulated insulin-loaded chitosan nanoparticles (INS-loaded CS NPs) prepared using 270 chitosans of different MW (CS 113 and CS 213), and control formulations (mean \pm SD, $n \geq$ 3): (\blacklozenge) 271 Microencapsulated INS-loaded CS NPs – CS 113; (**a**) Microencapsulated INS-loaded CS NPs – CS 272 213; (\circ) Microencapsulated blank (without insulin) CS NPs - CS 113; (\Box) Mannitol microspheres 273 containing INS; (Δ) Suspension of INS-loaded CS NPs – CS 113; (\bullet) INS solution in PBS pH 7.4; 274 *Statistically significant differences from microencapsulated blank CS NPs (p < 0.05); #Statistically 275 significant differences from INS solution (p < 0.05). Reprinted with permission from [47].

276 Solid lipid nanoparticles (SLN) were also proposed for this end and were reported to provide 277 homogeneous distribution through the lung upon delivery to diabetic rats by nebulisation, showing 278 relative bioavailability of insulin of 22.3% comparing with subcutaneous injection [50]. An approach 279 similar to that referred above of chitosan nanoparticles microencapsulated in mannitol microparticles 280 was later reported for the systemic delivery of calcitonin. The inhalable carriers had mass median 281 aerodynamic diameter (MMAD) of 2.7 µm and fine particle fraction (FPF) of 64%, the latter 282 representing the fraction of particles with aerodynamic diameter lower than 5 µm [51]. After IT 283 administration, around 85% relative bioavailability was determined, comparing with subcutaneous 284 delivery. The bioavailability was also superior to that obtained after inhalation of native calcitonin 285 [52]. Another approach in the same line, proposed the delivery of IgG mediated by poly(lactic-co-286 glycolide) acid (PLGA)-based nanoparticles produced by double emulsification and subsequently 287 spray-dried to acquire suitable aerodynamic properties. Leucine was further included to improve 288 aerosolisation. MMAD around 4 µm and FPF of approximately 50% indicated suitability to reach the 289 deep lung, while a prolonged release up to 35 days was observed in PBS pH 7.4, enabling applications 290 where prolonged release is envisaged [53].

291 More than simply avoiding injections, the driving force fostering investment on systemic drug 292 delivery through the lung relies on the improvement of pharmacokinetics, which could be an 293 advantage for drugs currently delivered through the oral, buccal or transdermal routes. The studies 294 reported above reinforce the potential of the lung to provide an access to the systemic compartment, 295 but above all, they show that the nanocarriers can play a role in improving the therapeutic 296 performance. Nevertheless, one of the limitations that is relatively tranversal to works on lung 297 delivery is the fact that, in most cases, the *in vivo* testing of inhalable formulations is performed by IT 298 administration, which does not mimic the reality when human delivery is concerned. This aspect still 299 requires some advancement in order to better predict in vivo outcomes.

300 The delivery of drugs that are specifically directed to the lung, is the other side of the picture of 301 lung delivery. Local treatment of lung diseases usually aims at low systemic bioavailability in order 302 to avoid the risk of unwanted side effects in other organs due to rapid drug translocation via the air-303 blood barrier. Some respiratory pathologies are ineffectively treated with existing small molecule-304 based therapies. RNAi effectors, such as small interfering RNA (siRNA), have shown to enable the 305 post-transcriptional silencing of key molecular disease factors that cannot be readily targeted with 306 conventional small molecule drugs [54]. Therefore, some therapeutic alternatives are currently being 307 proposed in this context. The type of cell that is targeted in this approach is variable and depends on 308 the specific airway disease. Epithelial cells are key players in cystic fibrosis, for instance, while 309 dendritic cells, macrophages and T lymphocytes are the targets in inflammatory diseases like asthma 310 or COPD [54]. The local therapeutic response to siRNA can be markedly enhanced through the use 311 of nanoparticles, essentially due to the possibility to provide specific cell targeting. The period of time 312 that siRNA is retained in the lung plays an important role on the success of the approaches. This 313 period is affected by rapid elimination due to mucociliary clearance, translocation to systemic 314 circulation and secondary organs, and phagocytosis by alveolar macrophages. The complexation of 315 siRNA with polyethylenimine (PEI), forming polyplexes, was demonstrated to reduce the 316 translocation and extend siRNA retention time in lung, while preventing substantial phagocytosis by 317 macrophages and avoiding extensive mucociliary clearance [55]. Additionally, it has been shown in 318 several occasions [56,57] that the contact of nanoparticles with the surfactant present in the alveolar 319 zone leads to the coating of nanocarriers by a biomolecular corona, composed of lipids and proteins. 320 This corona affects nanoparticle hydrophobicity and possibly enhances biorecognition, with 321 consequences on the subsequent interactions with cells and other biological entities. Most works 322 report a negative impact of this process on the therapeutic outcomes. Interestingly, with regards to 323 the delivery of siRNA, recent works have suggested that modifying the surface of siRNA-loaded 324 nanoparticles with lung surfactant (by a simple incubation) provides improved siRNA transfer 325 activity due to facilitated cellular uptake [54]. Improved transfection efficiency of pDNA was also 326 reported previously in presence of lung surfactant [58]. siRNA-dendrimer (polyamidoamine, 327 generation 4) complexes of ~100-130 nm were microencapsulated in trehalose-inulin microparticles,

328 which displayed aerodynamic diameters of 4.5 – 5.5 µm, adequate to reach the deep lung. These 329 microparticles dissolved in aqueous medium, releasing the nanocomplexes, which showed enhanced 330 cellular uptake and transfection in RAW264.7 macrophages, comparing with native siRNA [59]. 331 Protein-based molecules are, in many cases, regarded as sensitive and their manipulation in delivery 332 devices such as inhalers is often feared. A study demonstrated the stability of mRNA upon 333 nebulisation, showing no effect of nebulisation on protein duration of action or the cytotoxicity of the 334 formed PEI polyplexes [60].

335 Inhalable vaccines have also been the focus of several works and, although many pulmonary 336 vaccines have been proposed, only few involve nanocarriers. An interesting approach was reported 337 that uses a double emulsion formed by water/PLGA in organic/lactose-water, with IgG, the model 338 antibody, dissolved in the inner aqueous phase. The emulsion was spray-dried, resulting in PLGA 339 nanoparticles within lactose microparticles. Suitable properties for inhalation were observed, with 340 60% FPF. Submicron-particles were released after contact with aqueous medium, and approximately 341 70% IgG released after 6 days in pH 7.4 [53]. Another approach reported poly(glycerol adipate-co- ω -342 pentadecalactone) (PGA-co-PDL) nanoparticles that were modified to express on their surface the 343 pneumococcal surface protein A, which is an important antigen of S Pneumoniae (~20 mg antigen/mg 344 of nanoparticles). Nanoparticles of approximately 150 nm were then microencapsulated in leucine 345 microparticles to provide respirability. The latter registered MMAD of 1.7 µm and 74% FPF, which 346 grants the ability to reach the broncho-alveolar zone, potentiating the uptake by dendritic cells, as 347 demonstrated experimentally [61]. Silica nanoparticles were also reported for this end. Nanoparticles 348 were associated with plant-derived H1N1 influenza hemagglutinin antigen (HAC1) and proposed as 349 inhalable vaccine against influenza virus. A mucosal adjuvant (bis-(3',5')-cyclic dimeric guanosine 350 monophos-phate (c-di-GMP)) was further tested. After IT vaccination of mice, the double-adjuvanted 351 vaccine (nanoparticles plus mucosal adjuvant) were observed to induce high systemic antibody 352 responses, comparable to the systemic vaccination control. Moreover, local IgG and IgA responses 353 were observed in the bronchoalveolar lavage [62].

The described works clearly demonstrate that the lung provides a suitable route for the delivery of protein-based molecules, serving, in this context, the purpose of both systemic and local delivery.

356 2.2. Delivery of antibiotics

357 The delivery of antibiotics to the lung seems a very reasonable approach in the treatment of 358 infections that are based in that organ. In fact, the most common routes of delivery of antibiotics are 359 the oral and parenteral, even if the treatment of respiratory infections is intended. Addressing local 360 lung infections requires reaching effective concentrations of drug in the organ, which implies the 361 administration of significantly high doses and a general exposure of the organism to the drugs. The 362 direct administration to the infection site would, thus, permit using lower doses and avoid or 363 decrease systemic exposure, with the consequent reduction of systemic side-effects. Additionally, the 364 more targeted delivery is a premise to decrease the incidence of antimicrobial resistance, an important 365 current goal in antibiotic therapy [63,64]. Antibiotic resistance has been, for many years, one of the 366 greatest public health problems. The increasing misuse of these molecules, ever since their discovery, 367 has been making bacteria progressively resistant, by means of the development of specific cellular 368 mechanisms. This has been continuously and consistently posing a renewed challenge to the 369 treatment of infectious diseases [65].

370 The market makes available some formulations of inhaled antibiotics, including tobramycin, 371 colistin and aztreonan, which are mainly directed to the treatment of infections associatied with cystic 372 fibrosis conditions [66]. Other applications have been reported occasionally, such as the use of 373 aerosolised antibiotics in hospital-acquired pneumonia [67]. Research in the area has been increasing 374 consistently and a recent review on inhalable antibiotic formulations is available in [66]. Along with 375 the discovery of new antibiotics, the development of delivery systems to improve the therapeutic 376 performance of the molecules has been object of scientific efforts and both approaches are, in fact, 377 effective countermeasures against antibiotic resistance. The search for new drug molecules is known 378 to be slower than the development of drug delivery systems that lead the antibiotics to the intended

379 site of action. Of the marketed formulations referred above, none is based on nanocarriers, but the 380 literature provides many works reporting their use to improve the performance of lung delivered 381 antibiotics, addressing, among others, the improvement of kinetic profiles and issues related with 382 side effects. One of the most popular respiratory infections is tuberculosis, caused by Mycobacterium 383 tuberculosis, which primarily accumulates and replicates inside alveolar macrophages located in the 384 alveolar zone of the lung [68]. Despite the existence of effective therapy of tuberculosis for many 385 decades, the fact is that it still remains a global epidemic, being a major healthcare problem, as 386 portrayed by the last data published by WHO [69]. Not only the established therapy is prolonged 387 and associated with severe side effects, which decreases therapeutic compliance, but also the issues 388 of co-morbility with HIV and the existing bacterial resistance are relevant. A great number of works 389 propose the use of nanocarriers for tuberculosis treatment, in many cases envisaging lung delivery 390 applications. Very frequently, the developed carriers involve strategies of surface chemical 391 functionalisation, namely mannosylation. The rationale behind this approach is based on the fact that 392 bacterial hosts, the macrophages, have several surface receptors that are likely to be used as 393 therapeutic targets [70,71]. The mannose receptor is one of the main, which may provide a favourable 394 interaction with some units and chemical groups present on the carriers' surface, including the 395 mannose units, but also others like fucose and N-acetylglucosamine [72]. In principle, considering 396 that the bacteria are hosted by the macrophages located in the alveoli, this is the zone to be reached 397 in the design of any strategy aimed at treating tuberculosis by lung delivery.

398 SLN have been proposed as carriers for this end. Rifabutin-loaded SLN prepared with glyceryl 399 dibehenate (~100 nm) were further encapsulated in mannitol microparticles to acquire adequate 400 aerodynamic properties to reach the alveolar zone (~44% of particles with less than 6.4 μ m). An *in* 401 vivo test in a murine model of infection (Mycobacterium tuberculosis strain H37Rv) demonstrated that 402 the inhalation of the dry powder permitted effective delivery of the antibiotic to the lung, along with 403 drug distribution to liver and spleen. Moreover, an enhancement of antibacterial activity was 404 observed compared to nontreated animals [43]. Another formulation of SLN, this time composed of 405 palmitic acid and cholesteryl myristate and loaded with rifampicin (~ 400 nm), was further freeze-406 dried to obtain an inhalable powder. MMAD around 5 – 7 μ m and FPF within 30% and 50% were 407 determined. The SLN were mannosylated to improve their targeting ability, which was verified 408 experimentally, with increased macrophage uptake (~80%) compared to non-functionalised SLN 409 (~40%) [71,73]. Rifampicin was also the chosen antitubercular drug to encapsulate in polymer-410 glycerosomes, which showed to be more stable than conventional liposomes [74]. These are 411 phospholipid/glycerol vesicles combined with trimethyl chitosan or hyaluronic acid (80 – 110 nm). 412 Upon nebulisation, MMAD of approximately 4 µm was obtained along with FPF up to 77%. In any 413 case, the aerodynamic performance of the carriers was always better than that of the free drug and 414 drug incorporation in the vesicles was found to increase its efficacy against Staphylococcus aureus. 415 Following IT administration to rats, glycerosomes promoted the accumulation of rifampicin in the 416 lung, with lower systemic distribution, and low accumulation in other organs. The formulation 417 containing hyaluronic acid was found to perform more favourably [75]. Although it was not 418 discussed, the use of hyaluronic acid might be beneficial due to a favourable interaction of its N-419 acteylglucosamine units with CD44 [76,77] and mannose receptors [78]. Chitosan and chitosan-folate 420 were further used to functionalise oleic acid-based nanoemulsions loaded with rifampicin, which 421 were nebulised to render adequate respirability (MMAD of $3 - 4 \mu m$ and FPF of 62-73%). It was found 422 that chitosan-folate provided increased cell internalisation, proposed to result from a favourable 423 interaction with macrophages by both chitosan units and folate groups. Additionally, this 424 formulation provided in vivo higher lung drug content and reduced plasma drug concentration [79]. 425 Chitosan nanoparticles prepared by ionic gelation with tripolyphosphate were also proposed a 426 couple of times as carriers in antitubercular drug delivery. A first work described the association of 427 isoniazid and used spray-drying with lactose and leucine to reach an FPF of 45% [80]. More recently, 428 similar nanoparticles associated bedaquiline (size varying within 70 and 700 nm depending on 429 preparation conditions). A powder form of the nanoparticles was obtained by freeze-drying, 430 registering 28% FPF and 3.38 µm MMAD, which was better than the conventional DPI formulation 431 used as control (15% FPF and MMAD of 4 µm). The study determined absence of toxicity of the 432 nanoparticles in vivo in rats and further demonstrated higher drug concentration in lungs upon 433 inhalation of the microencapsulated nanocarriers [81]. Frequently, the choice of chitosan as 434 nanoparticle matrix material is not explicitly justified, leaving the readers with the sensation that the 435 polymer is only used because of its high popularity, a natural consequence of its favourable 436 properties regarding mucoadhesion and absence of toxicity. In this latter work, the authors justified 437 the positive results with a possible favoured uptake of nanoparticles by alveolar macrophages 438 mediated by an interaction of chitosan positive charges (from amino groups) with the negatively 439 charged surface of macrophages. However, most of the works fail to point out that the strong affinity 440 of macrophages by chitosan is possibly a result of the recognition of N-acetylglucosamine units of the 441 polymer by macrophage surface receptors, as was proposed in a work from our group reporting 442 chitosan microparticles as antitubercular drug carriers [78,82]. The use of chitosan as matrix material 443 was also proposed in genipin-crosslinked carboxymetylchitosan nanoparticles loaded with isoniazid 444 and rifampicin, which were freeze-dried to obtain a powder. After inhalation by rats, a greater 445 accumulation of drugs was observed in the lung upon delivery of the carriers compared with the free 446 drugs. Additionally, extended residence time of drugs in the lung was achieved and lower levels in 447 other organs (liver, kidney) were registered [83].

As a whole, several nanoparticle-based formulations are proposed in the frame of tuberculosis therapy, in most cases showing improved results attributed to specific functionalisation of their surface or benefits from their proper composition (e.g. chitosan). In order to provide adequate respirability, the nanocarries are either nebulised or transformed in inhalable powders using sprayor freeze-drying. In the works showing *in vivo* results, the delivery by inhalation tipically provided increased lung concentrations of the drug and lower systemic exposure.

- 454 Other lung diseases work as a door for opportunistic infections, cystic fibrosis being a major 455 example. This is a genetic disorder caused by mutations in the cystic fibrosis transmembrane 456 conductance regulator (CFTR) gene. This gene is of the utmost importance, as it encodes a protein 457 that forms an ion channel in epithelial cell membranes. The genetic disfunction may translate into 458 different defects of the protein, in any case ending up in bronchial obstruction that occurs due to the 459 secretion and accumulation of a thick and sticky mucus in the airways. The accumulation of mucus 460 creates the adequate conditions for bacterial colonisation, which tipically involves Pseudomonas 461 aeruginosa and Staphilococcus aureus [84-86]. This justifies that cystic fibrosis therapy requires regular 462 administration of antibiotics, apart from bronchodilators and mucolytics.
- 463 A solution of tobramycin for inhalation was the first approved aerosolised antibiotic to be used 464 against *P. aeruginosa* and, recently, a dry powder form of tobramycin has become available. However, 465 this drug shows poor mucus penetration, rapid clearance and suboptimal concentrations at the site 466 of infection, which are frequently not enough to stop the complications derived from the bacterial 467 infection [87]. The need for better therapies is one of the emmergent objectives in the field of cystic 468 fibrosis. Nanotechnology can bring forth some solutions in this context. Mucus penetration is, indeed, 469 a major issue. If it is possible to overcome this barrier, enabling a more effective delivery of drugs, 470 infections can be eliminated with higher efficiency. In a very interesting work, Schneider et al. (2017) 471 demonstrated that mucus penetrating nanoparticles (polystyrene nanoparticles coated with 472 polyethylene glycol - PEG) of size up to 300 nm have higher retention in the lung and more uniform 473 distribution compared with similar sized nanoparticles devoid of PEG and, thus, mucoadhesive [88]. 474 Regretably, no biological assays were reported so far, either in vitro or in vivo. Colistin was 475 encapsulated in PLGA nanoparticles which were further surface-modified with chitosan (270 nm) or 476 polyvinyl alcool (PVA, 330 nm) and then spray-dried to reach adequate aerodynamic properties. 477 MMAD less than $< 5 \mu m$ was obtained when lactose was used as carrier, while the use of mannitol 478 resulted in MMAD < 8 µm. In vitro assays revealed increased ability of chitosan-modified particles to 479 penetrate artifical mucus and also suggested a role of the nanoparticles in potentiating the anti-480 biofilm activity of colistin, possibly due to the ability of nanoparticles to penetrate the biofilm and to 481 sustain drug release [89]. A previous work from the same group, where tobramycin was encapsulated 482 in similar nanoparticles, demonstrated in vivo that PVA-modified nanoparticles reached the alveoli,

while particles modified with chitosan tend to appear in the upper airways, possibly as a consequence of their specific aerodynamic characteristics [90]. Ciprofloxacin was self-assembled with PEG-*g*phthaloyl chitosan nanoparticles (218 nm) and further microencapsulated by spray-drying in swellable alginate microparticles (volume mean diameter of 3.9 μm). Upon IT delivery to rats, the encapsulated molecule was found in higher concentration in lung tissue and lung lavage comparing with the administration of the control consisting of a physical mixture of lactose and micronised drug [91].

490 Importantly, many bacteria regulate pathogenicity via a cell-to-cell communication system that 491 is known as quorum sensing. This is dependent on cell density and involves the production of 492 virulence factors to coordinate group behaviours [92]. Antibacterial strategies based on the inhibition 493 of quorum sensing are currently growing and this represents, indeed, a novel form of therapy. A very 494 interesting approach in the delivery of antibiotics for the treatment of *Pseudomonas aeruginosa* 495 infection involved SLN (< 100 nm) loaded with a quorum sensing inhibitor. Nebulisation has resulted 496 in MMAD of 2.2 µm and FPF around 85% was determined, enabling the deposition of a certain 497 fraction in the bronchial region. The SLN demonstrated to penetrate into artifical sputum, but the 498 most important finding was that the proper SLN have anti-virulent effect, acting in addition to the 499 quorum sensing inhibitor to decrease the virulence factor pyocyanin [93].

500 As can be verified, under the scope of antibiotic delivery, a great deal of attention is given to 501 tuberculosis. Anyway, the number of works addressing antibiotic delivery mediated by nanocarriers 502 that present a certain degree of multifunctionality while providing real demonstration of potential 503 for lung delivery, represents only a fraction. Apart from tuberculosis, Pseudomonas aeruginosa and 504 Staphilococcus aureus are the two main targets, being frequently associated with cystic fibrosis and 505 pneumonia, although they can be also involved in hospital-acquired lung infections, for instance. It 506 was demonstrated in the several works described that the nanocarriers can provide extra strength to 507 antibiotic-mediated therapies.

508 2.3. Applications in cancer therapy

509 The WHO refers to lung cancer as one of the most lethal [94]. In 2018, 18.4% of cancer-related 510 deaths were of lung cancer, and the number of new cases (11.8%) was one of the highest, on par with 511 breast cancer [95]. WHO has a set of goals to fight cancer aggresively, and the development of new 512 strategies in cancer treatment is a priority worldwide. Lung cancer can be categorised into non-small 513 cell lung cancer (NSCLC) and small cell lung cancer (SCLC). NSCLC is considered aggressive and 514 comprises approximately 85% of all occurrences, in which various subtypes are included such as 515 adenocarninoma and squamous cell lung cancer. SCLC is even more aggressive, comprising the 516 remaining 15% of cases [96]. The high probability of metastasis derived from SCLC, and the 517 frequently late diagnosis, contribute to the high mortatility [97,98]. At earlier stages, the treatment for 518 both types of lung cancer is surgery, enabling the removal of the affected area. However, at later 519 stages, chemotherapy and radiation are the valid options, often to reduce the tumor mass before any 520 surgical procedure [97,99]. Regretably, these options have great impact on patient's physiology, as 521 both cancerous and healthy cells are attacked, resulting in symptoms that are difficult to manage. As 522 a consequence, patient susceptibility to other diseases is increased.

523 The scientific community has been working to develop more targeted therapies, which is 524 facilitated by the increasing information on molecular pathways, specific receptors and cancer 525 microenvironment, enabling different treatment approaches. Although the intravenous route is the 526 most used to deliver anticancer drugs, inclusive in lung cancer, the use of the lung route is an 527 alternative yet to be fully explored in lung cancer therapy. This approach would allow a more 528 targeted delivery, reaching directly the affected area, possibly with higher effectiveness than that 529 provided by systemic delivery. Importantly, the lung can be considered the main route for the 530 delivery of anticancer drugs in cases of lung cancer, but can also be used as add-on therapy for the 531 treatment of lung metastasis secondary to other cancers. Overall, it is considered that this approach 532 would potentially enable the use of lower doses of anticancer drugs, with reduced systemic exposure

and consequent residual metabolisation of the molecules [100]. This strategy further helps on the reduction of adverse effects, contributing for the increased quality of life of the patients.

The number of nanocarriers proposed for an application in lung cancer mediated by lung delivery is high. In most cases, a therapeutic effect is envisaged, but some of the works address diagnostic purposes. Although this is of great importance in cancer, especially at early stages of development, these strategies will not be detailed further, as they are out of the scope of the review. For further reading on this matter, Silva et al. (2019) and Mottaghitalab et al. (2019) comprise two comprehensive reviews on potential diagnostic stategies [101,102]. Therefore, only works on nanocarriers envisaging therapeutic approaches will be considered.

542 The general observation of the literature indicates that, in most cases, the proposal of 543 nanocarriers for an application in cancer therapy implies functionalisation, that is, carriers with some 544 sort of surface modification that benefits their interaction with the tumour environment. One of the 545 strategies often reported in this context relies on the use of a matrix that is added of molecules 546 potentially recognised by cell receptors prevailing in cancer cells comparing with healthy cells. Such 547 an approach was already discussed briefly in the previous section, referring to carriers endowed with 548 cell targeting ability mediated by mannose moieties. In the context of lung cancer, lactoferrin-549 chondroitin sulfate nanocomplexes (~190 nm) were reported to co-deliver doxorrubicin (Dox) and 550 elagic acid. The latter was first converted into water soluble nanocrystals due to its hydrophobicity. 551 The nanocomplexes were prepared by electrostatic interaction between lactoferrin and chondroitin 552 sulfate and the two drugs incorporated during this process. Due to the overexpression of CD44 and 553 lactoferrin receptors on the surface of lung cancer cells, these nanocomplexes were shown to have 554 favoured cell recognition, mediated by chondroitin sulfate and lactoferrin content, respectively. The 555 authors further hypothesised that clathrin-mediated endocytosis could have contributed favourably 556 to the internalisation of nanocomplexes, as their size is within the range of the pore size of the clathrin 557 receptor (up to 200 nm) [103]. Therefore, the functionality of these carriers is provided not only by 558 their size but also by their composition, which ensures specific targeting ability. To provide adequate 559 aerodynamics for lung delivery, the nanocomplexes were then microencapsulated into a mannitol 560 matrix, reaching FPF close to 90% and MMAD of 2.56 um. After IT insufflation of the 561 microencapsulated nanocomplexes in tumour-bearing mice, tumour growth biomarkers were 562 quantified and revealed lower levels when the inhalable formulation was used, in comparison with 563 the inhalation of free drugs or intravenous administration [42].

564 These cell recognition strategies were also addressed in works with gold nanoparticles. Such 565 carriers have strong interest in cancer therapy, finding applications in photothermal therapy, 566 radiotherapy and also as drug carriers. Their inhalation has been demonstrated to provide lung 567 accumulation, which can be useful in lung cancer therapy [104]. A very recent review on the topic is 568 available in [105]. Gold nanoparticles (2 nm) that were coated with functional derivatives of thiolated 569 PEG have shown invisibility towards the immune system provided by PEG [106], but also enabled 570 attaching other moieties to provide specific targeting. The surface of the nanocarriers was thus 571 modified with the ligand RGD, a peptide with relatively high and specific affinity for integrins 572 overexpressed in tumour neovasculature [107,108]. A mice model of single-nodule lung 573 adenocarcinoma [109] was used to establish which route of administration, either inhalation or 574 intravenous delivery, would be more effective on adenocarcinoma targeting using the nanocarriers. 575 The biodistribution data demonstrated higher concentration of the carriers upon inhalation [110]. In 576 another approach, gold nanoparticles were loaded with temozolomide (~40 nm), an alkylating agent 577 already in use in other cancer types. The IT administration to healthy mice indicated the safety of 578 gold nanoparticles upon quantification of lactate dehydrogenase and the tumour markers 579 carcinoembryonic antigen and alpha-fetoprotein. The proper carriers were reported to induce 580 oxidative damage and ability to inhibit cell proliferation and cell cycle in G1-phase, while the delivery 581 of drug-loaded carriers to mice bearing lung cancer demonstrated a synergic effect between the 582 carriers and the loaded drug [111].

583 The optimisation of the interaction of nanocarriers with cancer cells has also been reported using 584 SLN. A complex nanodelivery system based on SLN was proposed, being composed of multi585 compartmental lipid nanocomposites (190-225 nm). Berberin and rapamycin, with demonstrated 586 synergic anticancer effect, were initially encapsulated in SLN. To optimise the rate of delivery of both 587 drugs, multicompartment systems were developed. Berberin was incorporated as hydrophobic ion 588 pair with sodium dodecyl sulfate in SLN's core, sustaining its release, while rapamycin was pre-589 formulated as phospholipid complex, thus helping to improve its solubility and relatively enhance 590 its release. The tumour targeting ability was improved by layer-by-layer assembly of the cationic 591 lactoferrin and the anionic hyaluronic acid, which target the CD44 and lactoferrin receptors 592 overexpressed by lung cancer cells. Adequate aerodynamics were achieved after spray-drying with 593 a mixture of mannitol/maltodextrin/leucine (MMAD of 3.3 µm, FPF of 56%). An assay in mice bearing 594 lung tumour demonstrated that inhaled nanocomposites induced a decrease of lung weight 595 comparing with the inhalation of free drugs, along with reduction of tumour size and levels of 596 angiogenic markers [112]. Another work proposed the modification of SLN surface with a chitosan 597 derivative that was previously added of folate moieties [113]. The authors hypothesised that both the 598 chitosan derivative and the folate engraftments would increase the retention of the nanoparticles 599 within the lungs, and activate the folate receptors, increasing the amount of drug delivered to cancer 600 cells. The nanocarriers (~250 nm; +32 mV) provided slower release of paclitaxel after coating (58% in 601 3 days) and demonstrated binding affinity to cell lines expressing the folate receptor. In *in vivo* assays, 602 higher lung paclitaxel concentration was observed for the inhaled chitosan-coated SLN compared 603 with the intravenous administration of the drug. Moreover, drug concentration was higher at 1 h and 604 6 h post-administration for the coated formulation compared with inhaled and intravenous delivered 605 paclitaxel. As a final remark for this study, the authors noticed that the SLN were distributed 606 throughout the solid lung tumours, with low interaction with the vessels, which occurs with systemic 607 delivery of anticancer agents. Paclitaxel was also loaded in PEG-polylactic acid (PLA) nanoparticles 608 that were further conjugated with the epithelial cell adhesion molecule (EpCAM, CD326), also 609 overexpressed in lung cancer. IT delivery of nanoparticles to c-Raf transgenic lung cancer mice 610 permitted reducing drug toxicity, with animal surviving increasing from 20% to 70% [114]. Another 611 approach proposed lipid polymeric nanoparticles (hydrophobic polymeric core, phospholipid layer 612 and an outer layer of epidermal growth factor (EGF), PEG and distearoylphosphoethanolamine) 613 targeting the EGF receptor (EGFR) [115], which is overexpressed in lung carcinoma [116,117]. 614 Cisplatin and Dox were the associated drugs. The presence of EGF in the outer part of the 615 nanoparticle promoted the interaction with EGFR, leading to the release of drugs at the cancer site. 616 An in vivo assay revealed tumour inhibition ratio of ~75%.

617 Chitosan-coated PLGA nanoplexes were proposed to carry an antisense oligonucleotide against 618 the human telomerase RNA component, as telomerase activity is detected in most NSCLC. The 619 potential of the oligonucleotide as a telomerase inhibitor has been described [118], although its poor 620 cellular uptake hinders its use in cancer therapy. The nanoplexes (±160 nm) were delivered IT to 621 healthy mice, using the model of the isolated perfused and ventilated lung, and provided increased 622 uptake of the oligonucleotide by the epithelium than that observed for the free form of the 623 oligonucleotide. Although no specific study was performed, the authors justified the results with the 624 potential ability of nanoparticles to escape pulmonary clearance mechanisms [119].

625 A solution towards a resistant form of cancer was proposed as inhalable self-assembled 626 nanoparticles comprised of human serum albumin (HSA), tumor necrosis factor (TNF)-related 627 apoptosis-inducing ligand (TRAIL) and Dox [120]. The latter was conjugated to HSA and formed 628 nanoparticles, which were then coated with TRAIL (342 nm). Initial tests in H226 cells, which are 629 representative of NSCLC, have shown that the simultaneous presence of Dox and TRAIL enabled 630 increased cytotoxic potential, as cell viability after 3 days of exposure decreased from approximately 631 60% when only one of the molecules was present in HSA nanoparticles, to 20-30% after dual 632 association. An in vivo assay was then performed in lung tumour-bearing mice, delivering the 633 nanoparticles in the form of micron-sized liquid droplets. The tumours of mice treated with HSA 634 nanoparticles combining TRAIL and Dox were much smaller and lighter than those of mice treated 635 with the corresponding nanoparticles containing only one of the molecules, TRAIL or Dox. 636 Haloperidol was also used as ligand to enhance targeting ability of albumin-based nanoparticles (218

nm). Nanoparticles were prepared by desolvation of bovine serum albumin, previously conjugated
with haloperidol and loaded with Dox. Spray-drying with mannitol, trehalose and leucine resulted
in nano-in-microparticles with aerodynamic diameter of 4.6 μm and FPF of 66% [121].

640 Some works further report therapeutic approaches that rely on the ability of the carrier matrix 641 to respond to different stimuli [122,123]. Modification of temperature and pH are usual stimuli to be 642 used [124], setting the basis for the elaboration of the so called *smart polymers* or *systems*. The rationale 643 behind their use is that a certain stimulus (pH value or temperature that is reached), will trigger a 644 phase transition in the carrier matrix, leading to the release of the drug in a predetermined site. In 645 this context, a copolymer based on methoxy poly(ethylene glycol)-poly(ethylenimine)-poly(L-646 glutamate) was produced and nanoparticles prepared (< 75 nm), using electrostatic interaction and 647 chelate effect to encapsulate simultaneously Dox and cisplatin [125]. In vitro assays demonstrated 648 increased release of Dox at acidic pH, showing the capacity to release the drug in a cancer setting. 649 The pulmonary administration of the nanocarriers to mice with metastatic lung cancer was 650 performed using a Microsprayer aeroliser, resulting in increased accumulation of carriers within the 651 lungs, along with low concentration in other tissues, especially in the area surrounding tumour 652 lesions. It was hypothesised that the smaller size of the carriers benefitted their penetration in the 653 cancer mass, while an inneffective vessel arrangement prevented a systemic dissemination. The 654 results also shown decreased tumour masses, suggesting increased efficacy of the nanoformulation. 655 Dendrimers of poly(amidoamine) were also used in a similar pH stimulation strategy. Dox was 656 conjugated with the polymer and the dendrimers were spray-dried with mannitol to endow suitable aerodynamic characteristics (FPF was 40-60%). Dendrimers readily released from microparticles in 657 658 aqueous medium and drug release was only found to occur in response to intracellular pH drop [126]. 659 In another work, similar dendrimers showed strong time-dependent toxicity in Calu-3 cells, a model 660 of the respiratory epithelium, which was attributed to sustained drug release. It was also shown that 661 the conjugation of PEG molecules to the dendrimer improved their permeation across the cell layer, 662 in a concentration-dependent manner. In this case, the dendrimers were formulated in a pressurised 663 metered dose inhaler, leading to aerosols with 82% FPF and MMAD of 1.3 µm [127]. Dendrimers of 664 PEGylated polylysine, also conjugated with Dox, were IT administered to a syngenic rat model of 665 lung metastised breast cancer. An administration twice a week led to over 95% reduction in lung 666 tumour after two weeks, comparing with IV administration of Dox solution, which resulted in a 667 reduction of 30%-50% [128].

668 In some cases, a combination of the above-mentioned strategies is proposed, as happens in a 669 work reporting a stimuli-responsive core-shell nanoparticle conjugated with folic acid. The rationale 670 behind this formulation was to create a pH- and temperature-sensitive network, comprised by a 671 copolymer of poly(N-isopropylacrylamide) and carboxymethylchitosan, which comprises the shell 672 of the nanosystem. In turn, the core is comprised of PLGA and an image contrast agent 673 (superparamagnetic iron oxide, SPIO). While PLGA allows the controlled release of the encapsulated 674 molecules, gemcitabine in this case, SPIO serves the dual role of contrasting agent and inductor of 675 temperature change by external application of an alterning magnetic field. SPIO-induced 676 temperature alterations lead to conformational change of polymeric shell, allowing drug release. 677 Additionally, the shell of the system is pH-sensitive, providing drug release at the acidic pH 678 characteristic of cancer environment. Moreover, the delivery is even more targeted owing to the 679 surface conjugation with folic acid, benefitting from the overexpression of the folate receptor in 680 cancer cells [129]. The nanocarriers (~289 nm, -36 mV) evidenced increased cell uptake in the presence 681 of a magnet, as a consequence of the presence of SPIO in the formulation. In vivo assays in lung 682 tumour-bearing mice show decreased tumour volume comparing with the controls. Pulmonary 683 retention of nanoparticles was confirmed by magnetic ressonance imaging (MRI) and, when coupled 684 with radiotherapy, a synergic effect takes place to slow tumour growth.

Finally, magnetic nanoparticles have also been proposed several times within the scope of lung
cancer. Many reports explore an application in lung cancer diagnosis, using Fe₃O₄ paramagnetic cores
[130] or gadolinium-based particles [131,132], in the latter further enabling a radiosensitising effect.
Nevertheless, therapeutic actions are also proposed. Iron oxide nanoparticles (Fe₃O₄; 56 nm, -49 mV)

689 were spray-dried with lactose and doxorubicin, reaching an MMAD of 3.27 µm. An *in vitro* study 690 demonstrated that, comparing with a liquid suspension, the microencapsulated nanoparticles 691 provided more than twice deposition and retention of particles in regions under the influence of a 692 strong magnetic gradient [133].

693

694 As a whole, several different strategies are described that end up with positive results in lung 695 cancer treatment. Nevertheless, cancer research still has much ground to cover and the associated 696 therapeutics are growing at the rate of decipheration of new receptors and new molecular cascades. 697 Nanotechnology is progressing along these discoveries, to provide improved strategies of cancer 698 treatment. Those described above are dominated by the optimisation of the carriers surface, either by 699 engineering with specific ligands, by carefully selecting the matrix components or by combining all 700 the effects, in order to provide more targeted delivery of the drugs and an intimate contact with 701 cancer cells, which will thus result in improved therapeutics.

702

703 **3.** Expectations for the future

704 Pharmaceutical technology has been playing a vital role on medicine, as it allows exploring 705 different materials and their combinations to prepare drug carriers and to further endow these with 706 better properties that enable reaching the desired target sites, ending up with therapeutic success. 707 Many questions arise around the topic of nanocarrier-based lung drug delivery. Scientists have been 708 giving many and varied answers, in the attempt to address all the rising issues, finding alternatives 709 and engineering adequate systems to fulfill requirements and needs. One of the concerns is always 710 the fate of the drug. In lung drug delivery, the objective is sometimes to retain the drug in the lung, 711 as happens in local delivery approaches, thus minimising the systemic absorption. In other cases, a 712 systemic effect is the desired outcome and the carriers are engineered to avoid retention. The options 713 to address therapeutic demands are varied, as seen by the plethora of systems, alternatives and 714 engineering possibilities described through the review. As expected, the global analysis reveals that 715 most of the works focus on the use of the lung route to attain local rather than systemic effects. It was 716 also verified that, from the three main topics explored in the review, the delivery of anticancer drugs 717 is the one concentrating more proposals in the literature, which is justified by the severity of the 718 numbers associated with this disease, the increasing number of patients and the lack of therapeutic 719 options, apart from the marketing appeal of cancer therapies.

720 Whichever the specific topic, it has become clear that the use of nanocarriers is an added value 721 and may provide an evolution of therapeutic responses if used properly and the arising toxicological 722 concerns are addressed. Furthermore, the engineering of different strategies mainly involved surface 723 functionalisation of the carriers or at least took benefit of their components to provide specific effects. 724 The latter is perhaps the strategy still requiring further investment to bring out all its potential, as 725 smart polymers have emerged as a new range of powerful tools, but still need refinement. The use of 726 the different physiological conditions, such as pH, temperature and redox compounds or even light 727 at different wavelengths can be the answer to more targeted and efficent therapies. Objectivelly, the 728 field needs clinically feasible formulations, which possibly could combine some of the different 729 strategies that were described, certainly using some kind of surface engineering to reach specific 730 biological targets, but also adjusting the desired properties with the use of materials that may respond 731 specifically, either to stimuli or to the established characteristics of the target area. Recently, a very 732 interesting and promising approach was reported in this context, comprising a nanoparticle-in-733 microgel system that provides drug release triggered by the presence of proteases. As the presence 734 of these enzymes is greatly increased in the lung as a consequence of the inflammatory processs 735 related to asthma, COPD and cystic fibrosis, this may comprise a therapeutic strategy for the 736 treatment of these conditions [134].Despite the tremendous advancement, the evolution of the field 737 is in strong dependence of new knowledge being generated at more basic science, namely the 738 molecular mechanisms of the diseases, which are great indicators of tools to be used in the 739 development of new therapies. Additionally, it cannot be forgotten that many of the tools identified

740 as successful and providing improved therapeutic responses, that is, carriers and above all, the 741 materials, are not approved so far by the regulatory entities for an application in lung delivery. This 742 poses a great challenge itself. Addressing the toxicity of inhaled therapeutic nanocarriers is actually 743 a matter of inescapable importance. For many years now it has become clear that the biocompatibility 744 of nanomaterials is not that of the raw materials and its evaluation needs to go much beyond the 745 assessment of the isolated components. The nanomaterial must be considered a new entity instead, 746 within the context of a specific delivery route [135]. Therefore, generating data on the safety of the 747 nanocarriers and the new materials identified as potential adjuvants, in the framework of the lung 748 route, is currently understood as an urgent need to potentiate lung drug delivery applications. This 749 should involve toxicity tests that evaluate all the possible toxicity pathways, both in vitro and in vivo, 750 while ensuring that the 3Rs policy to reduce, refine and replace the use of animals in research is 751 followed. The initial in vitro tests should address cytotoxicity and genotoxicity, and should also 752 evaluate potential epigenetic toxicity [136]. The fate of the proper carriers after the delivery is often 753 disregarded. A very recent study comparing the clearance kinetics of liposomes and solid lipid 754 nanoparticles after IT delivery of suspensions to rats has shown similar clearance rates, despite 755 different deposition patterns [137]. Studies around this topic are, thus, imperative to provide data on 756 the safety of the materials and the kick-off to their clinical application.

757 It is important to point out that, when in vivo assays were described, which occurred in a 758 considerable number of the presented works, IT delivery of the nanocarriers was the predominant 759 technique for the assessment, which implies a high risk when establishing possible correlations with 760 human delivery. The fact that nanocarriers themselves do not exhibit suitable aerodynamics for 761 inhalation is also a matter of relevance, as this always implies an extra step, tipically proposed to 762 involve the spray-drying of nanocarriers to produce nano-in-microcarriers that can deposit in the 763 lung.

764 The area still needs to evolve in several topics before inhalable nanocarriers enter clinical trials. 765 Not only the question of performing more realistic in vivo assays is determinant, but also the 766 toxicological assessment plays a defining role. Helpful technologies have been arising, such as 3D 767 printing, which was used on the printing of artificial airways that enable the study of particle 768 flowability and dose assessment. Lim et al. describe this application in the neonate, showing a 769 powerful tool to improve the ethics associated with formulation testing and to provide solutions to 770 children born with respiratory complications [138].

771 All in all, this review highlighted an integrative process that considers progress made at the level 772 of basic science, which clarifies pathophysiological aspects of each clinical condition, and the 773 development of tools and strategies to reach the pharmacological targets. Many works were 774 described with inhalable nanocariers that have been showing potential, even with the existing 775 limitations. All the issues, however, point out to a common objective of providing the knowledge to 776 enable the engineering of nanocarriers that will promote improved lung therapeutics.

777

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784 References



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788

- Singh, L.; Kruger, H.G.; Maguire, G.E.M.; Govender, T.; Parboosing, R. The role of nanotechnology in the treatment of viral infections. *Therapeutic Advances in Infectious Diseases* 2017, *4*, 105-131, doi:10.1177/2049936117713593.
- Peres, C.; Matos, A.I.; Conniot, J.; Sainz, V.; Zupančič, E.; Silva, J.M.; Graça, L.; Sá
 Gaspar, R.; Préat, V.; Florindo, H.F. Poly(lactic acid)-based particulate systems are
 promising tools for immune modulation. *Acta Biomaterialia* 2017, 48, 41-57,
 doi:10.1016/j.actbio.2016.11.012.
- Movahedi, F.; Li, L.; Gu, W.; Xu, Z.P. Nanoformulations of albendazole as effective anticancer and antiparasite agents. *Nanomedicine* 2017, *12*, 2555-2574, doi:10.2217/nnm-2017-0102.
- Newman, S.P. Delivering drugs to the lungs: The history of repurposing in the treatment of respiratory diseases. *Advanced Drug Delivery Reviews* 2018, *133*, 5-18, doi:10.1016/j.addr.2018.04.010.
- Pushpakom, S.; Iorio, F.; Eyers, P.A.; Escott, K.J.; Hopper, S.; Wells, A.; Doig, A.;
 Guilliams, T.; Latimer, J.; McNamee, C., et al. Drug repurposing: Progress,
 challenges and recommendations. *Nature Reviews Drug Discovery* 2018, *18*, 41-58,
 doi:10.1038/nrd.2018.168.
- 806 6. *Tratado de Tecnología Farmacéutica, Volumen II: Operaciones básicas*; Pacheco,
 807 R.M., Ed. Editorial Síntesis S.A.: Spain, 2016; pp. 458.
- 808 7. Edgar, J.Y.C.; Wang, H. Introduction for design of nanoparticle based drug delivery
 809 systems. *Current Pharmaceutical Design* 2017, 23, 2108-2112,
 810 doi:10.2174/1381612822666161025154003.
- 8. Park, K. Controlled drug delivery systems: Past forward and future back. *Journal of Controlled Release* 2014, *190*, 3-8, doi:10.1016/j.jconrel.2014.03.054.
- 813 9. Rodrigues, S.; Alves, A.D.; Cavaco, J.S.; Pontes, J.F.; Guerreiro, F.; Rosa da Costa,
 814 A.M.; Buttini, F.; Grenha, A. Dual antibiotherapy of tuberculosis mediated by
 815 inhalable locust bean gum microparticles. *International Journal of Pharmaceutics*816 2017, 529, 433-441, doi:10.1016/j.ijpharm.2017.06.088.
- 817 10. Cunha, L.; Rodrigues, S.; Rosa da Costa, A.M.; Faleiro, M.L.; Buttini, F.; Grenha, A.
 818 Inhalable fucoidan microparticles combining two antitubercular drugs with potential 819 application in pulmonary tuberculosis therapy. *Polymers* 2018, 10, 636, 820 doi:10.3390/polym10060636.
- 11. Wang, Q.; Mi, G.; Hickey, D.; Li, Y.; Tu, J.; Webster, T.J.; Shen, Y. Azithromycinloaded respirable microparticles for targeted pulmonary delivery for the treatment of
 pneumonia. *Biomaterials* 2018, 160, 107-123,
 doi:10.1016/j.biomaterials.2018.01.022.
- Agüero, L.; Zaldivar-Silva, D.; Peña, L.; Dias, M.L. Alginate microparticles as oral
 colon drug delivery device: A review. *Carbohydrate Polymers* 2017, *168*, 32-43,
 doi:10.1016/j.carbpol.2017.03.033.
- Hu, Y.; Li, M.; Zhang, M.; Jin, Y. Inhalation treatment of idiopathic pulmonary fibrosis with curcumin large porous microparticles. *International Journal of Pharmaceutics* 2018, 551, 212-222, doi:10.1016/j.ijpharm.2018.09.031.
- Rivera Gil, P.; Hühn, D.; del Mercato, L.L.; Sasse, D.; Parak, W.J. Nanopharmacy:
 Inorganic nanoscale devices as vectors and active compounds. *Pharmacological Research* 2010, 62, 115-125, doi:10.1016/j.phrs.2010.01.009.
- International Organization for Standardization (ISO). ISO/TS 80004-2-2015
 Nanotechnologies Vocabulary Part 2: Nano-objects. 2015; Vol. ISO/TS 80004-22015.
- Food and Drug Administration (FDA). Guidance for Industry Considering wether
 an FDA-Regulated Product Involves the Application of Nanotechnology. U.S.
 Department of Health and Human Services, Ed. 2014; Vol. FDA-2010-D-0530.

- Bangham, A.D.; Standish, M.M.; Watkins, J.C. Diffusion of univalent ions across the
 lamellae of swollen phospholipids. *Journal of Molecular Biology* 1965, *13*, 238-252,
 doi:10.1016/s0022-2836(65)80093-6.
- 18. Abdelaziz, H.M.; Gaber, M.; Abd-Elwakil, M.M.; Mabrouk, M.T.; Elgohary, M.M.;
 Kamel, N.M.; Kabary, D.M.; Freag, M.S.; Samaha, M.W.; Mortada, S.M., et al.
 Inhalable particulate drug delivery systems for lung cancer therapy: Nanoparticles,
 microparticles, nanocomposites and nanoaggregates. *Journal of Controlled Release*2018, 269, 374-392, doi:10.1016/j.jconrel.2017.11.036.
- Mishra, V.; Bansal, K.K.; Verma, A.; Yadav, N.; Thakur, S.; Sudhakar, K.;
 Rosenholm, J.M. Solid lipid nanoparticles: Emerging colloidal nano drug delivery
 systems. *Pharmaceutics* 2018, *10*, 191, doi:10.3390/pharmaceutics10040191.
- 851 20. Huynh Mai, C.; Thanh Diep, T.; Le, T.T.T.; Nguyen, V. Advances in colloidal
 852 dispersions: A review. *Journal of Dispersion Science and Technology* 2019,
 853 10.1080/01932691.2019.1591970, 1-16, doi:10.1080/01932691.2019.1591970.
- Garcês, A.; Amaral, M.H.; Sousa Lobo, J.M.; Silva, A.C. Formulations based on solid
 lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) for cutaneous use:
 A review. *European Journal of Pharmaceutical Sciences* 2018, *112*, 159-167,
 doi:10.1016/j.ejps.2017.11.023.
- Jaiswal, P.; Gidwani, B.; Vyas, A. Nanostructured lipid carriers and their current application in targeted drug delivery. *Artificial Cells, Nanomedicine, and Biotechnology* 2014, 44, 27-40, doi:10.3109/21691401.2014.909822.
- 23. Cordani, M.; Somoza, Á. Targeting autophagy using metallic nanoparticles: A
 promising strategy for cancer treatment. *Cellular and Molecular Life Sciences* 2019,
 76, 1215-1242, doi:10.1007/s00018-018-2973-y.
- Hoang Thi, T.T.; Cao, V.D.; Nguyen, T.N.Q.; Hoang, D.T.; Ngo, V.C.; Nguyen, D.H. 864 24. 865 Functionalized mesoporous silica nanoparticles and biomedical applications. 866 Materials Science Engineering 2019, 631-656, and С 99, doi:10.1016/j.msec.2019.01.129. 867
- 868 25. Hillaireau, H.; Couvreur, P. Nanocarriers' entry into the cell: Relevance to drug delivery. *Cellular and Molecular Life Sciences* 2009, 66, 2873-2896, doi:10.1007/s00018-009-0053-z.
- 871 26. Csaba, N.; Garcia-Fuentes, M.; Alonso, M.J. The performance of nanocarriers for 872 transmucosal drug delivery. *Expert Opinion on Drug Delivery* 2006, *3*, 463-478, 873 doi:10.1517/17425247.3.4.463.
- Singh, R.; Lillard Jr., J.W. Nanoparticle-based targeted drug delivery. *Experimental and Molecular Pathology* 2009, *86*, 215-223, doi:10.1016/j.yexmp.2008.12.004.
- Weissig, V.; Pettinger, T.K.; Murdock, N. Nanopharmaceuticals (part 1): Products on
 the market. *International Journal of Nanomedicine* 2014, 9, 4357-4373,
 doi:10.2147/IJN.S46900.
- 879 29. Bernabeu, E.; Cagel, M.; Lagomarsino, E.; Moretton, M.; Chiappetta, D.A.
 880 Paclitaxel: What has been done and the challenges remain ahead. *International* 881 *Journal of Pharmaceutics* 2017, *526*, 474-495, doi:10.1016/j.ijpharm.2017.05.016.
- Weissig, V.; Guzman-Villanueva, D. Nanopharmaceuticals (part 2): Products in the
 pipeline. *International Journal of Nanomedicine* 2015, 10, 1245-1257,
 doi:10.2147/IJN.S65526.
- 885 31. Global Health Estimates 2016: Disease burden by Cause, Age, Sex, by Country and
 886 by Region, 2000-2016; World Health Organization (WHO): Geneva, 2018.
- 887 32. Hillery, A.M.; Park, K. Drug Delivery Fundamentals & Applications, 2nd ed.; CRC
 888 Press: 2017.
- 889 33. Newman, S.P. Drug delivery to the lungs: Challenges and opportunities. *Therapeutic Delivery* 2017, *8*, 647-661, doi:10.4155/tde-2017-0037.

- 891 34. Borghardt, J.M.; Kloft, C.; Sharma, A. Inhaled therapy in respiratory disease: The complex interplay of pulmonary kinetic processes. *Canadian Respiratory Journal* 2018, 2018, 2732017, doi:10.1155/2018/2732017.
- 894 35. Mack, G.S. Pfizer dumps Exubera. *Nature Biotechnology* 2007, 25, 1331-1332, doi:10.1038/nbt1207-1331.
- Zaykov, A.N.; Mayer, J.P.; DiMarchi, R.D. Pursuit of a perfect insulin. *Nature Reviews Drug Discovery* 2016, 15, 425-439, doi:10.1038/nrd.2015.36.
- 898 37. Almeida, A.J.; Grenha, A. Chapter 22 - Technosphere[®]: An inhalation system for 899 pulmonary delivery of biopharmaceuticals. In Mucosal Delivery of 900 Biopharmaceuticals: Biology, Challenges and Strategies, Springer: 2014; 901 10.1007/978-1-4614-9524-6.
- 38. Jesus, S.; Schmutz, M.; Som, C.; Borchard, G.; Wick, P.; Borges, O. Hazard
 assessment of polymeric nanobiomaterials for drug delivery: What can we learn from
 literature so far. *Frontiers in Bioengineering and Biotechnology* 2019, 7, 261,
 doi:10.3389/fbioe.2019.00261.
- 39. Grenha, A.; Seijo, B.; Remuñán-Lopez, C. Microencapsulated chitosan nanoparticles
 907 for lung protein delivery. *European Journal of Pharmaceutical Sciences* 2005, 25,
 908 427-437, doi:10.1016/j.ejps.2005.04.009.
- Gaspar, D.P.; Vital, J.; Leiva, M.C.; Goncalves, L.M.D.; Taboada, P.; Remuñán-Lopez, C.; Vítor, J.; Almeida, A.J. Transfection of pulmonary cells by stable pDNA-polycationic hybrid nanostructured particles. *Nanomedicine* 2019, *14*, 407-429, doi:10.2217/nnm-2018-0270.
- Shi, M.; Zhao, X.; Zhang, J.; Pan, S.; Yang, C.; Wei, Y.; Hu, H.; Qiao, M.; Chen, D.;
 Zhao, X. pH-responsive hybrid nanoparticle with enhanced dissociation characteristic
 for siRNA delivery. *International Journal of Nanomedicine* 2018, *13*, 6885-6902,
 doi:10.2147/IJN.S180119.
- 42. Abd Elwakil, M.M.; Mabrouk, M.T.; Helmy, M.W.; Abdelfattah, E.-Z.A.; Khiste,
 S.K.; Elkhodairy, K.A.; Elzoghby, A.O. Inhalable lactoferrin-chondroitin
 nanocomposites for combined delivery of doxorubicin and ellagic acid to lung
 carcinoma. *Nanomedicine* 2018, *13*, 2015-2035, doi:10.2217/nnm-2018-0039.
- 43. Gaspar, D.P.; Gaspar, M.M.; Eleutério, C.V.; Grenha, A.; Blanco, M.; Goncalves,
 L.M.D.; Taboada, P.; Almeida, A.J.; Remuñán-Lopez, C. Microencapsulated solid
 lipid nanoparticles as a hybrid platform for pulmonary antibiotic delivery. *Molecular Pharmaceutics* 2017, *14*, 2977-2990, doi:10.1021/acs.molpharmaceut.7b00169.
- 44. Gao, W.; Chen, Y.; Zhang, Y.; Zhang, Q.; Zhang, L. Nanoparticle-based local antimicrobial drug delivery. *Advanced Drug Delivery Reviews* 2018, 127, 46-57, doi:10.1016/j.addr.2017.09.015.
- 928 45. Tratado de Tecnología Farmacéutica, Volumen III: Formas de dosificación;
 929 Pacheco, R.M., Ed. Editorial Síntesis S.A.: Spain, 2017; pp. 458.
- Grenha, A.; Remuñán-López, C.; Carvalho, E.L.S.; Seijo, B. Microspheres containing
 lipid/chitosan nanoparticles complexes for pulmonary delivery of therapeutic
 proteins. *European Journal of Pharmaceutics and Biopharmaceutics* 2008, 69, 83933 93, doi:10.1016/j.ejpb.2007.10.017.
- 47. Al-Qadi, S.; Grenha, A.; Carrion-Recio, D.; Seijo, B.; Remuñán-López, C.
 Microencapsulated chitosan nanoparticles for pulmonary protein delivery: *In vivo*evaluation of insulin-loaded formulations. *Journal of Controlled Release* 2012, *157*,
 383-390, doi:10.1016/j.jconrel.2011.08.008.
- 938 48. Grenha, A.; Grainger, C.I.; Dailey, L.A.; Seijo, B.; Martin, G.P.; Remuñán-Lopez,
 939 C.; Forbes, B. Chitosan nanoparticles are compatible with respiratory epithelial cells
 940 *in vitro. European Journal of Pharmaceutical Sciences* 2007, *31*, 73-84,
 941 doi:10.1016/j.ejps.2007.02.008.

- 942 49. Shi, K.; Liu, Y.; Ke, L.; Fang, Y.; Yang, R.; Cui, F. Epsilon-poly-L-lysine guided
 943 improving pulmonary delivery of supramolecular self-assembled insulin
 944 nanospheres. *International Journal of Biological Macromolecules* 2015, 72, 1441945 1450, doi:10.1016/j.ijbiomac.2014.10.023.
- 50. Liu, J.; Gong, T.; Fu, H.; Wang, C.; Wang, X.; Chen, Q.; Zhang, Q.; He, Q.; Zhang,
 Solid lipid nanoparticles for pulmonary delivery of insulin. *International Journal of Pharmaceutics* 2008, *356*, 333-344, doi:10.1016/j.ijpharm.2008.01.008.
- 949 51. Buttini, F.; Colombo, G.; Kwok, P.C.L.; Wui, W.T. Chapter 6 Aerodynamic
 950 assessment for inhalation products: Fundamentals and current pharmacopoeial
 951 methods. In *Inhalation Drug Delivery*, 2013; 10.1002/9781118397145.ch6pp. 91952 119.
- 52. Sinsuebpol, C.; Chatchawalsaisin, J.; Kulvanich, P. Preparation and *in vivo* absorption evaluation of spray dried powders containing salmon calcitonin loaded chitosan nanoparticles for pulmonary delivery. *Drug Design, Development and Therapy* 2013, 7, 861-873, doi:10.2147/DDDT.S47681.
- 53. Kaye, R.S.; Purewal, T.S.; Alpar, H.O. Simultaneously manufactured nano-in-micro
 (SIMANIM) particles for dry-powder modified-release delivery of antibodies. *Journal of Pharmaceutical Sciences* 2009, *98*, 4055-4068, doi:10.1002/jps.21673.
- 960 54. De Backer, L.; Cerrada, A.; Pérez-Gil, J.; De Smedt, S.C.; Raemdonck, K. Bioinspired materials in drug delivery: Exploring the role of pulmonary surfactant in siRNA inhalation therapy. *Journal of Controlled Release* 2015, 220, 642-650, doi:10.1016/j.jconrel.2015.09.004.
- 55. Lipka, J.; Semmler-Behnke, M.; Wenk, A.; Burkhardt, J.; Aigner, A.; Kreyling, W.
 Biokinetic studies of non-complexed siRNA versus nano-sized PEI F25LMW/siRNA polyplexes following intratracheal instillation into mice. *International Journal of Pharmaceutics* 2016, *500*, 227-235, doi:10.1016/j.ijpharm.2016.01.038.
- 56. Hu, Q.; Bai, X.; Hu, G.; Zuo, Y.Y. Unveiling the molecular structure of pulmonary
 surfactant corona on nanoparticles. *ACS Nano* 2017, *11*, 6832-6842,
 doi:10.1021/acsnano.7b01873.
- 57. Hu, G.; Jiao, B.; Shi, X.; Valle, R.P.; Fan, Q.; Zuo, Y.Y. Physicochemical properties
 of nanoparticles regulate translocation across pulmonary surfactant monolayer and
 formation of lipoprotein corona. *ACS Nano* 2013, 7, 10525-10533,
 doi:10.1021/nn4054683.
- 58. Nguyen, J.; Reul, R.; Betz, T.; Dayyoub, E.; Schmehl, T.; Gessler, T.; Bakowsky, U.;
 Seeger, W.; Kissel, T. Nanocomposites of lung surfactant and biodegradable cationic
 nanoparticles improve transfection efficiency to lung cells. *Journal of Controlled Release* 2009, *140*, 47-54, doi:10.1016/j.jconrel.2009.07.017.
- 979 59. Agnoletti, M.; Bohr, A.; Thanki, K.; Wan, F.; Zeng, X.; Boetker, J.P.; Yang, M.;
 980 Foged, C. Inhalable siRNA-loaded nano-embedded microparticles engineered using
 981 microfluidics and spray drying. *European Journal of Pharmaceutics and*982 *Biopharmaceutics* 2017, 120, 9-21, doi:10.1016/j.ejpb.2017.08.001.
- 983 60. Johler, S.M.; Rejman, J.; Guan, S.; Rosenecker, J. Nebulisation of IVT mRNA
 984 complexes for intrapulmonary administration. *PLoS One* 2015, *10*, e0137504,
 985 doi:10.1371/journal.pone.0137504.
- 986 Kunda, N.K.; Alfagih, I.M.; Miyaji, E.N.; Figueiredo, D.B.; Goncalves, V.M.; 61. 987 Ferreira, D.M.; Dennison, S.R.; Somavarapu, S.; Hutcheon, G.A.; Saleem, I.Y. Pulmonary dry powder vaccine of pneumococcal antigen loaded nanoparticles. 988 989 International Journal of **Pharmaceutics** 2015. 495. 903-912. 990 doi:10.1016/j.ijpharm.2015.09.034.
- 991 62. Neuhaus, V.; Chichester, J.A.; Ebensen, T.; Schwarz, K.; Hartman, C.E.; Shoji, Y.;
 992 Guzman, C.A.; Yusibov, V.; Sewald, K.; Braun, A. A new adjuvanted nanoparticle-

- based H1N1 influenza vaccine induced antigen-specific local mucosal and systemic
 immune responses after administration into the lung. *Vaccine* 2014, *32*, 3216-3222,
 doi:10.1016/j.vaccine.2014.04.011.
- Scherließ, R.; Etschmann, C. DPI formulations for high dose applications Challenges and opportunities. *International Journal of Pharmaceutics* 2018, 548, 49doi:10.1016/j.ijpharm.2018.06.038.
- 999 64. Scherließ, R. Future of nanomedicines for treating respiratory diseases. *Expert*1000 *Opinion on Drug Delivery* 2019, *16*, 59-68, doi:10.1080/17425247.2019.1553955.
- Brinkac, L.; Voorhies, A.; Gomez, A.; Nelson, K.E. The threat of antimicrobial resistance on the human microbiome. *Microbial Ecology* 2017, *74*, 1001-1008, doi:10.1007/s00248-017-0985-z.
- 1004 66. Woods, A.; Rahman, K.M. Antimicrobial molecules in the lung: Formulation
 1005 challenges and future directions for innovation. *Future Medicinal Chemistry* 2018, 10, 575-604, doi:10.4155/fmc-2017-0162.
- 1007 67. Luyt, C.-E.; Hékimian, G.; Bréchot, N.; Chastre, J. Aerosol therapy for pneumonia in
 1008 the intensive care unit. *Clinics in Chest Medicine* 2018, *39*, 823-836,
 1009 doi:10.1016/j.ccm.2018.08.005.
- 1010 68. Alves, A.D.; Cavaco, J.S.; Guerreiro, F.; Lourenco, J.P.; Rosa da Costa, A.M.;
 1011 Grenha, A. Inhalable antitubercular therapy mediated by locust bean gum
 1012 microparticles. *Molecules* 2016, *21*, 702, doi:10.3390/molecules21060702.
- 1013 69. *Global tuberculosis report 2018*; World Health Organization (WHO): Geneva, 2018.
 1014 70. Guerreiro, F.; Pontes, J.F.; Rosa da Costa, A.M.; Grenha, A. Spray-drying of konjac
 1015 glucomannan to produce microparticles for an application as antitubercular drug
 1016 carriers. *Powder Technology* 2019, *342*, 246-252, doi:10.1016/j.powtec.2018.09.068.
- 1017 71. Maretti, E.; Costantino, L.; Buttini, F.; Rustichelli, C.; Leo, E.; Truzzi, E.; Iannuccelli,
 1018 V. Newly synthesized surfactants for surface mannosylation of respirable SLN
 1019 assemblies to target macrophages in tuberculosis therapy. *Drug Delivery and*1020 *Translational Research* 2018, 9, 298-310, doi:10.1007/s13346-018-00607-w.
- 1021 72. Auger, M.J.; Ross, J.A. Chapter 1 The biology of the macrophage. In *The macrophage: The natural immune system*, 1st ed.; Lewis, C.E., McGee, J.O.D., Eds.
 1023 Oxford University Press: New York, 1992.
- Maretti, E.; Costantino, L.; Rustichelli, C.; Leo, E.; Croce, M.A.; Buttini, F.; Truzzi,
 E.; Iannuccelli, V. Surface engineering of Solid Lipid Nanoparticle assemblies by
 methyl α-d-mannopyranoside for the active targeting to macrophages in antituberculosis inhalation therapy. *International Journal of Pharmaceutics* 2017, *528*,
 440-451, doi:10.1016/j.ijpharm.2017.06.045.
- Manca, M.L.; Castangia, I.; Caddeo, C.; Pando, D.; Escribano, E.; Valenti, D.;
 Lampis, S.; Zaru, M.; Fadda, A.M.; Manconi, M. Improvement of quercetin
 protective effect against oxidative stress skin damages by incorporation in
 nanovesicles. *Colloids and Surfaces B: Biointerfaces* 2014, 123, 566-574,
 doi:10.1016/j.colsurfb.2014.09.059.
- Melis, V.; Manca, M.L.; Bullita, E.; Tamburini, E.; Castangia, I.; Cardia, M.C.;
 Valenti, D.; Fadda, A.M.; Peris, J.E.; Manconi, M. Inhalable polymer-glycerosomes as safe and effective carriers for rifampicin delivery to the lungs. *Colloids and Surfaces B: Biointerfaces* 2016, *143*, 301-308, doi:10.1016/j.colsurfb.2016.03.044.
- 1038 76. Dicker, K.T.; Gurski, L.A.; Pradhan-Bhatt, S.; Witt, R.L.; Farach-Carson, M.C.; Jia,
 1039 X. Hyaluronan: A simple polysaccharide with diverse biological functions. *Acta*1040 *Biomaterialia* 2014, 10, 1558-1570, doi:10.1016/j.actbio.2013.12.019.
- 1041 77. Kwon, M.Y.; Wang, C.; Galarraga, J.H.; Pure, E.; Han, L.; Burdick, J.A. Influence of
 1042 hyaluronic acid modification on CD44 binding towards the design of hydrogel

1043		biomaterials.	Biomaterials	2019,	222,	119451,
1044	70	doi:10.1016/j.biom	aterials.2019.119451.	(C '1	ית ייו יי	1 · A /
1045	/8.	East, L.; Isacke, C. (PPA) 2002 1572	264 286 doi:10.1016	$\frac{1}{1000}$	BIOCNIMICA EL BI (02)00210-7	opnysica Acta
1040	70	(DDA) 2002, 1572, Shah K : Chan I W	504-580, 001.10.1010 V · Wong TW Critic	al physicoche	(02)00319-7.	rical attributes
1047	1).	of nanoemulsions for	or pulmonary delivery	<i>v</i> of rifampici	n by nebulization	n technique in
1049		tuberculosis tre	atment. Drug	Deliverv	2017 . 24.	1631-1647.
1050		doi:10.1080/107175	544.2017.1384298.	200000	,,	1001 1017,
1051	80.	Pourshahab, P.S.; (Gilani, K.; Moazeni,	E.; Eslahi, H	I.; Fazeli, M.R.;	Jamalifar, H.
1052		Preparation and cha	racterization of spray	dried inhalab	le powders conta	ining chitosan
1053		nanoparticles for p	ulmonary delivery of	isoniazid. J	ournal of Micro	encapsulation
1054		2011 , 28, 605-613,	doi:10.3109/0265204	8.2011.59943	37.	
1055	81.	Rawal, T.; Patel, S	.; Butani, S. Chitosan	nanoparticle	s as a promising	approach for
1056		pulmonary delivery	v of bedaquiline. Euro	opean Journa	al of Pharmaceu	tical Sciences
1057	0.2	2018 , <i>124</i> , 273-287	, doi:10.1016/j.ejps.20	018.08.038.		
1058	82.	Cunha, L.; Rodrigu	les, S.; Rosa da Costa	i, A.M.; Fale	iro, L.; Buttini, F	A.; Grenha, A.
1059		Innalable chitosan r	nicroparticles for simi	ultaneous del	ivery of isoniazio	and rifabutin
1060		1313 1320 doi:10	treatment. <i>Drug Deve</i>	<i>16</i> 08231	Inaustriai Pharn	<i>1acy</i> 2019 , 43,
1062	83	$W_{11} T \cdot I_{120} W \cdot W$	Wang $W \cdot 7hou$ I \cdot Ta	1006231. m. W · Xiana	W · Zhang I · (Suo I · Chen
1062	05.	$T \cdot Ma D$ et al	Geninin-crosslinked	carboxymeth	vl chitosan nanc	ogel for lung-
1063		targeted delivery of	isoniazid and rifampi	in <i>Carbohyd</i>	rate Polymers 20	18 197 403-
1065		413. doi:10.1016/j.	carbpol.2018.06.034.			
1066	84.	Ong, V.; Mei, V.;	Cao, L.; Lee, K.; Chu	ung, E.J. Nar	nomedicine for c	ystic fibrosis.
1067		SLAS Technology 2	019 , 24, 169-180, doi	:10.1177/247	2630318824334	
1068	85.	Kozakova, J.; Altay	, A.; Zdimal, V.; Mas	kova, L.; Sor	vico, F.; Quarta,	E.; Rossi, A.;
1069		Buttini, F.; Colom	bo, G. Dry powder	inhaler of co	olistimethate soc	lium for lung
1070		infections in cystic	fibrosis: optimization	of powder co	onstruction. Drug	Development
1071		and Industrial	<i>Pharmacy</i> 2019 ,	10.1080/036	539045.2019.165	2636, 1-25,
1072	96	doi:10.1080/036390)45.2019.1652636.	11 . 4 0	M ('0)	
1073	86.	Akdag Cayli, Y.; Sa	inin, S.; Buttini, F.; Ba	ulducci, A.G.;	Montanari, S.; V	ural, I.; Oner,
1074		L. Dry powders for	the inhalation of cipi	tionta Drug	Development	noined with a
1075		Pharmacy 2017 13	1378 1380 doi:10.1	080/036300/	Development a	na mausimai
1070	87	Moreno-Sastre M	· Pastor M · Fequisal	hel $\Delta \cdot \text{Sans}$	F · Viñas M ·	Fleischer Δ·
1077	07.	Palomino, E : Bach	iller D: Pedraz II.	Pulmonary	delivery of tobra	mycin-loaded
1079		nanostructured lipic	l carriers for <i>Pseudon</i>	ionas aerugir	<i>iosa</i> infections as	ssociated with
1080		cystic fibrosis. In	ternational Journal	of Pharmad	ceutics 2016, 4	98, 263-273,
1081		doi:10.1016/j.ijpha	rm.2015.12.028.	5	,	, , ,
1082	88.	Schneider, C.S.; X	u, Q.; Boylan, N.J.;	Chisholm, J.	; Tang, B.C.; S	chuster, B.S.;
1083		Henning, A.; Ensig	n, L.M.; Lee, E.; Ads	tamongkonk	ul, P., et al. Nan	oparticles that
1084		do not adhere to m	ucus provide uniform	n and long-la	sting drug delive	ery to airways
1085		following inhal	ation. Science	Advances	2017 , <i>3</i> ,	e1601556,
1086		doi:10.1126/sciadv.	.1601556.			
1087	89.	d'Angelo, I.; Casc	iaro, B.; Miro, A.;	Quaglia, F.;	Mangoni, M.L.	; Ungaro, F.
1088		Overcoming barrie	ers in <i>Pseudomonas</i>	aeruginosa	lung infections	Engineered
1089		nanoparticles for lo	ocal delivery of a ca	tionic antimi	crobial peptide.	Colloids and
1090	00	Surfaces B: Biointe	rjaces 2015, 135, 717	-/25, doi:10.	1016/J.colsurfb.2	2015.08.027.
1091	90.	Digaro, F.; d'Ange	10, 1.; Coletta, C.; dE	La Potonda	M L: Operlie E	Dry poyedore

1092R.; Perfetto, B.; Tufano, M.A.; Miro, A.; La Rotonda, M.I.; Quaglia, F. Dry powders1093based on PLGA nanoparticles for pulmonary delivery of antibiotics: Modulation of

1094encapsulation efficiency, release rate and lung deposition pattern by hydrophilic1095polymers. Journal of Controlled Release2012, 157, 149-159,1096doi:10.1016/j.jconrel.2011.08.010.

- 1097 91. Du, J.; El-Sherbiny, I.M.; Smyth, H.D. Swellable ciprofloxacin-loaded nano-in-micro
 1098 hydrogel particles for local lung drug delivery. *AAPS PharmSciTech* 2014, *15*, 15351099 1544, doi:10.1208/s12249-014-0176-x.
- Wu, S.; Liu, J.; Liu, C.; Yang, A.; Qiao, J. Quorum sensing for population-level control of bacteria and potential therapeutic applications. *Cellular and Molecular Life Sciences* 2019, 10.1007/s00018-019-03326-8, doi:10.1007/s00018-019-03326-8.
- Nafee, N.; Husari, A.; Maurer, C.K.; Lu, C.; de Rossi, C.; Steinbach, A.; Hartmann,
 R.W.; Lehr, C.-M.; Schneider, M. Antibiotic-free nanotherapeutics: Ultra-small,
 mucus-penetrating solid lipid nanoparticles enhance the pulmonary delivery and antivirulence efficacy of novel quorum sensing inhibitors. *Journal of Controlled Release* **2014**, *192*, 131-140, doi:10.1016/j.jconrel.2014.06.055.
- 1108 94. Cheng, T.-Y.D.; Cramb, S.M.; Baade, P.D.; Youlden, D.R.; Nwogu, C.; Reid, M.E.
 1109 The international epidemiology of lung cancer: Latest trends, disparities, and tumor
 1110 characteristics. *Journal of Thoracic Oncology* 2016, *11*, 1653-1671,
 1111 doi:10.1016/j.jtho.2016.05.021.
- 1112 95. *Fact sheet on lung cancer*; The Global Cancer Observatory, World Health 1113 Organization (WHO): 2019.
- Wu, L.; Leng, D.; Cun, D.; Foged, C.; Yang, M. Advances in combination therapy of
 lung cancer: Rationales, delivery technologies and dosage regimens. *Journal of Controlled Release* 2017, 260, 78-91, doi:10.1016/j.jconrel.2017.05.023.
- 1117 97. Lee, W.-H.; Loo, C.-Y.; Ghadiri, M.; Leong, C.-R.; Young, P.M.; Traini, D. The
 potential to treat lung cancer via inhalation of repurposed drugs. *Advanced Drug Delivery Reviews* 2018, *133*, 107-130, doi:10.1016/j.addr.2018.08.012.
- 1120 98. Tomoda, K.; Ohkoshi, T.; Hirota, K.; Sonavane, G.S.; Nakajima, T.; Terada, H.;
 1121 Komuro, M.; Kitazato, K.; Makino, K. Preparation and properties of inhalable
 1122 nanocomposite particles for treatment of lung cancer. *Colloids and Surfaces B:*1123 *Biointerfaces* 2009, 71, 177-182, doi:10.1016/j.colsurfb.2009.02.001.
- 1124 99. Lemjabbar-Alaoui, H.; Hassan, O.U.; Yang, Y.-W.; Buchanan, P. Lung cancer:
 1125 Biology and treatment options. *Biochimica et Biophysica Acta* 2015, *1856*, 189-210,
 1126 doi:10.1016/j.bbcan.2015.08.002.
- 1127 100. Rosière, R.; Hureaux, J.; Levet, V.; Amighi, K.; Wauthoz, N. La chimiothérapie
 1128 inhalée partie 1: Concept et challenges technologiques actuels. *Revue des Maladies*1129 *Respiratoires* 2018, *35*, 357-377, doi:10.1016/j.rmr.2018.02.001.
- 1130 101. Silva, C.O.; Pinho, J.O.; Lopes, J.M.; Almeida, A.J.; Gaspar, M.M.; Reis, C. Current
 1131 trends in cancer nanotheranostics: Metallic, polymeric, and lipid-based systems.
 1132 *Pharmaceutics* 2019, *11*, doi:10.3390/pharmaceutics11010022.
- 1133 102. Mottaghitalab, F.; Farokhi, M.; Fatahi, Y.; Atyabi, F.; Dinarvand, R. New insights
 1134 into designing hybrid nanoparticles for lung cancer: Diagnosis and treatment. *Journal*1135 of Controlled Release 2019, 295, 250-267, doi:10.1016/j.jconrel.2019.01.009.
- 1136 103. Rejman, J.; Oberle, V.; Zuhorn, I.S.; Hoekstra, D. Size-dependent internalization of particles via the pathways of clathrin- and caveolae-mediated endocytosis. *Biochemical Journal* 2004, *377*, 159-169, doi:10.1042/BJ20031253.
- 1139 104. Gadoue, S.M.; Toomeh, D. Radio-sensitization efficacy of gold nanoparticles in inhalational nanomedicine and the adverse effect of nano-detachment due to coating inactivation. *Physica Medica* 2019, *60*, 7-13, doi:10.1016/j.ejmp.2019.02.013.
- 1142105.Sztandera, K.; Gorzkiewicz, M.; Klajnert-Maculewicz, B. Gold nanoparticles in1143cancer treatment. Molecular Pharmaceutics2019, 16, 1-23,1144doi:10.1021/acs.molpharmaceut.8b00810.

- 1145 106. Kumar, R.; Korideck, H.; Ngwa, W.; Berbeco, R.I.; Makrigiorgos, G.M.; Sridhar, S.
 1146 Third generation gold nanoplatform optimized for radiation therapy. *Translational*1147 *Cancer Research* 2013, 2, doi:10.3978/j.issn.2218-676X.2013.07.02.
- 1148 107. Di Pietro, P.; Zaccaro, L.; Comegna, D.; Del Gatto, A.; Saviano, M.; Snyders, R.;
 1149 Cossement, D.; Satriano, C.; Rizzarelli, E. Silver nanoparticles functionalized with a
 1150 fluorescent cyclic RGD peptide: A versatile integrin targeting platform for cells and
 1151 bacteria. *RSC Advances* 2016, 6, 112381-112392, doi:10.1039/c6ra21568h.
- 108. Ganipineni, L.P.; Ucakar, B.; Joudiou, N.; Riva, R.; Jérôme, C.; Gallez, B.; Danhier,
 F.; Préat, V. Paclitaxel-loaded multifunctional nanoparticles for the targeted
 treatment of glioblastoma. *Journal of Drug Targeting* 2019, 27, 614-623,
 doi:10.1080/1061186X.2019.1567738.
- 109. Herter-Sprie, G.S.; Korideck, H.; Christensen, C.L.; Herter, J.M.; Rhee, K.; Berbeco,
 R.I.; Bennett, D.G.; Akbay, E.A.; Kozono, D.; Mak, R.H., et al. Image-guided
 radiotherapy platform using single nodule conditional lung cancer mouse models. *Nature Communications* 2014, *5*, 5870, doi:10.1038/ncomms6870.
- 1160 110. Ngwa, W.; Kumar, R.; Moreau, M.; Dabney, R.; Herman, A. Nanoparticle drones to target lung cancer with radiosensitizers and cannabinoids. *Frontiers in Oncology* 1162 2017, 7, 208, doi:10.3389/fonc.2017.00208.
- 1163 111. Hamzawy, M.A.; Abo-Youssef, A.M.; Salem, H.F.; Mohammed, S.A. Antitumor activity of intratracheal inhalation of temozolomide (TMZ) loaded into gold nanoparticles and/or liposomes against urethane-induced lung cancer in BALB/c mice. *Drug Delivery* 2017, 24, 599-607, doi:10.1080/10717544.2016.1247924.
- 1167 112. Kabary, D.M.; Helmy, M.W.; Abdelfattah, E.-Z.A.; Fang, J.-Y.; Elkhodairy, K.A.;
 1168 Elzoghby, A.O. Inhalable multi-compartmental phospholipid enveloped lipid core
 1169 nanocomposites for localized mTOR inhibitor/herbal combined therapy of lung
 1170 carcinoma. *European Journal of Pharmaceutics and Biopharmaceutics* 2018, *130*,
 1171 152-164, doi:10.1016/j.ejpb.2018.06.027.
- 1172 113. Rosière, R.; Van Woensel, M.; Gelbcke, M.; Mathieu, V.; Hecq, J.; Mathivet, T.;
 1173 Vermeersch, M.; Van Antwerpen, P.; Amighi, K.; Wauthoz, N. New folate-grafted
 1174 chitosan derivative to improve delivery of paclitaxel-loaded solid lipid nanoparticles
 1175 for lung tumour therapy by inhalation. *Molecular Pharmaceutics* 2018, *15*, 899-910,
 1176 doi:10.1021/acs.molpharmaceut.7b00846.
- 1177 114. Karra, N.; Nassar, T.; Laenger, F.; Benita, S.; Borlak, J. Safety and proof-of-concept efficacy of inhaled drug loaded nano- and immunonanoparticles in a c-Raf transgenic lung cancer model. *Current Cancer Drug Targets* 2012, *13*, 11-29, doi:10.2174/1568009611309010011.
- 1181 115. Nan, Y. Lung carcinoma therapy using epidermal growth factor receptortargeted lipid
 polymeric nanoparticles coloaded with cisplatin and doxorubicin. *Oncology Reports*2019, 42, 2087-2096, doi:10.3892/or.2019.7323.
- 1184 116. Pancewicz-Wojtkiewicz, J. Epidermal growth factor receptor and notch signaling in non-small-cell lung cancer. *Cancer Medicine* 2016, 5, 3572-3578, doi:10.1002/cam4.944.
- 1187 117. Yuan, M.; Huang, L.-L.; Chen, J.-H.; Wu, J.; Xu, Q. The emerging treatment
 1188 landscape of targeted therapy in non-small-cell lung cancer. *Signal Transduction and*1189 *Targeted Therapy* 2019, *4*, 61, doi:10.1038/s41392-019-0099-9.
- 1190 118. Pitts, A.E.; Corey, D.R. Inhibition of human telomerase by 2'-O-methyl-RNA.
 1191 *Proceedings of the National Academy of Sciences of the United States of America*1192 **1998**, 95, 11549-11554, doi:10.1073/pnas.95.20.11549.
- 1193 119. Dong, M.; Murdter, T.E.; Philippi, C.; Loretz, B.; Schaefer, U.F.; Lehr, C.M.;
 1194 Schwab, M.; Ammon-Treiber, S. Pulmonary delivery and tissue distribution of
 1195 aerosolized antisense 2'-O-Methyl RNA containing nanoplexes in the isolated

1196		perfused and ventilated rat lung. European Journal of Pharmaceutics and
1197		Biopharmaceutics 2012, 81, 478-485, doi:10.1016/j.ejpb.2012.04.022.
1198	120.	Choi, S.H.; Byeon, H.J.; Choi, J.S.; Thao, L.; Kim, I.; Lee, E.S.; Shin, B.S.; Lee,
1199		K.C.; Youn, Y.S. Inhalable self-assembled albumin nanoparticles for treating drug-
1200		resistant lung cancer. Journal of Controlled Release 2015, 197, 199-207,
1201		doi:10.1016/j.jconrel.2014.11.008.
1202	121.	Varshosaz, J.; Hassanzadeh, F.; Mardani, A.; Rostami, M. Feasibility of haloperidol-
1203		anchored albumin nanoparticles loaded with doxorubicin as dry powder inhaler for
1204		pulmonary delivery. Pharmaceutical Development and Technology 2015, 20, 183-
1205		196, doi:10.3109/10837450.2013.852576.
1206	122.	Lakkadwala, S.; Nguyen, S.; Nesamony, J.; Narang, A.S.; Boddu, S.H. Chapter 7 -
1207		Smart Polymers in Drug Delivery. In Excipient Applications in Formulation Design
1208		and Drug Delivery, Narang, A.S., Boddu, S.H., Eds. Springer: 2015; 10.1007/978-3-
1209		319-20206-8.
1210	123.	Liu, M.; Du, H.; Zhang, W.; Zhai, G. Internal stimuli-responsive nanocarriers for drug
1211		delivery: Design strategies and applications. Materials Science and Engineering C
1212		2017 , <i>71</i> , 1267-1280, doi:10.1016/j.msec.2016.11.030.
1213	124.	Hoffman, A.S. Stimuli-responsive polymers: Biomedical applications and challenges
1214		for clinical translation. Advanced Drug Delivery Reviews 2013, 65, 10-16,
1215		doi:10.1016/j.addr.2012.11.004.
1216	125.	Xu, C.; Wang, Y.; Guo, Z.; Chen, J.; Lin, L.; Wu, J.; Tian, H.; Chen, X. Pulmonary
1217		delivery by exploiting doxorubicin and cisplatin co-loaded nanoparticles for
1218		metastatic lung cancer therapy. Journal of Controlled Release 2019, 295, 153-163,
1219		doi:10.1016/j.jconrel.2018.12.013.
1220	126.	Zhong, Q. Co-spray dried mannitol/poly(amidoamine)-doxorubicin dry-powder
1221		inhaler formulations for lung adenocarcinoma: Morphology, in vitro evaluation, and
1222		aerodynamic performance. AAPS PharmSciTech 2018, 19, 531-540,
1223		doi:10.1208/s12249-017-0859-1.
1224	127.	Zhong, Q.; Humia, B.V.; Punjabi, A.R.; Padilha, F.F.; da Rocha, S.R.P. The
1225		interaction of dendrimer-doxorubicin conjugates with a model pulmonary epithelium
1226		and their cosolvent-free, pseudo-solution formulations in pressurized metered-dose
1227		inhalers. European Journal of Pharmaceutical Sciences 2017, 109, 86-95,
1228		doi:10.1016/j.ejps.2017.07.030.
1229	128.	Kaminskas, L.M.; McLeod, V.M.; Ryan, G.M.; Kelly, B.D.; Haynes, J.M.;
1230		Williamson, M.; Thienthong, N.; Owen, D.J.; Porter, C.J.H. Pulmonary
1231		administration of a doxorubicin-conjugated dendrimer enhances drug exposure to
1232		lung metastases and improves cancer therapy. Journal of Controlled Release 2014,
1233		183, 18-26, doi:10.1016/j.jconrel.2014.03.012.
1234	129.	Menon, J.U.; Kuriakose, A.; Iyer, R.; Hernandez, E.; Gandee, L.; Zhang, S.;
1235		Takahashi, M.; Zhang, Z.; Saha, D.; Nguyen, K.T. Dual-drug containing core-shell
1236		nanoparticles for lung cancer therapy. Scientific Reports 2017, 7, 13249,
1237		doi:10.1038/s41598-017-13320-4.
1238	130.	Smulders, S.; Ketkar-Atre, A.; Luyts, K.; Vriens, H.; Nobre, S.D.S.; Rivard, C.; Van
1239		Landuyt, K.; Baken, S.; Smolders, E.; Golanski, L., et al. Body distribution of SiO ₂ -
1240		Fe ₃ O ₄ core-shell nanoparticles after intravenous injection and intratracheal
1241		instillation. <i>Nanotoxicology</i> 2016 , <i>10</i> , 567-574,
1242	101	doi:10.3109/17435390.2015.1100761.
1243	131.	Dufort, S.; Bianchi, A.; Henry, M.; Lux, F.; Le Duc, G.; Josserand, V.; Louis, C.;
1244		Perriat, P.; Cremillieux, Y.; Tillement, O., et al. Nebulized gadolinium-based
1245		nanoparticles: a theranostic approach for lung tumor imaging and radiosensitization.
1246		Small 2015, 11, 215-221, doi:10.1002/smll.201401284.

- 1247 132. Bianchi, A.; Dufort, S.; Lux, F.; Fortin, P.-Y.; Tassali, N.; Tillement, O.; Coll, J.-L.; Cremillieux, Y. Targeting and in vivo imaging of non-small-cell lung cancer using 1248 1249 nebulized multimodal contrast agents. Proceedings of the National Academy of 1250 United States of America **2014**, 111. Sciences of the 9247-9252, 1251 doi:10.1073/pnas.1402196111.
- 1252 133. McBride, A.A.; Price, D.N.; Lamoureux, L.R.; Elmaoued, A.A.; Vargas, J.M.;
 1253 Adolphi, N.L.; Muttil, P. Preparation and characterization of novel magnetic nano1254 in-microparticles for site-specific pulmonary drug delivery. *Molecular*1255 *Pharmaceutics* 2013, 10, 3574-3581, doi:10.1021/mp3007264.
- 1256 134. Mejias, J.C.; Roy, K. *In-vitro* and *in-vivo* characterization of a multi-stage enzyme1257 responsive nanoparticle-in-microgel pulmonary drug delivery system. *Journal of*1258 *Controlled Release* 2019, *316*, 393-403, doi:10.1016/j.jconrel.2019.09.012.
- 1259 135. Gaspar, R.; Duncan, R. Polymeric carriers: Preclinical safety and the regulatory implications for design and development of polymer therapeutics. *Advanced Drug Delivery Reviews* 2009, *61*, 1220-1231, doi:10.1016/j.addr.2009.06.003.
- 1262 136. Dusinska, M.; Tulinska, J.; El Yamani, N.; Kuricova, M.; Liskova, A.; Rollerova, E.;
 1263 Rundén-Pran, E.; Smolkova, B. Immunotoxicity, genotoxicity and epigenetic toxicity
 1264 of nanomaterials: New strategies for toxicity testing? *Food Chemical Toxicology*1265 2017, 109, 797-811, doi:10.1016/j.fct.2017.08.030.
- 1266 137. Haque, S.; Whittaker, M.; McIntosh, M.P.; Pouton, C.W.; Phipps, S.; Kaminskas,
 1267 L.M. A comparison of the lung clearance kinetics of solid lipid nanoparticles and
 1268 liposomes by following the ³H-labelled structural lipids after pulmonary delivery in
 1269 rats. *European Journal of Pharmaceutics and Biopharmaceutics* 2018, *125*, 1-12,
 1270 doi:10.1016/j.ejpb.2018.01.001.
- 1271 138. Lim, S.H.; Kathuria, H.; Tan, J.J.Y.; Kang, L. 3D printed drug delivery and testing
 1272 systems a passing fad or the future? *Advanced Drug Delivery Reviews* 2018, *132*,
 1273 139-168, doi:10.1016/j.addr.2018.05.006.