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Abstract

The Antarctic Zone, the southernmost belt of the Antarctic Circumpolar Current, plays an important role in the control of atmospheric carbon dioxide concentrations. In the last decade, a number of studies have highlighted the importance of diatom assemblage composition in influencing the magnitude of the organic carbon and biogenic silica fluxes exported out of the mixed layer in Southern Ocean ecosystems. Here we investigate the relationship between the makeup of the diatom assemblage, organic carbon and biogenic silica export and several significant environmental parameters using sediment trap records deployed in different sectors of the Antarctic Zone. The study is divided in two parts. We first present unpublished diatom species flux data collected by a sediment trap in the offshore waters of Prydz Bay (Station PZB-1) over a year. The results of this study revealed a major export peak of diatom valves in Austral summer and two small unexpected secondary flux pulses during full winter conditions. The summer diatom sinking assemblages were largely composed of small and rapidly dividing species such as *Fragilariopsis cylindrus*, *Fragilariopsis curta* and *Pseudo-nitzschia lineola*, while winter assemblages were dominated by *Fragilariopsis kerguelensis* most reflecting its persistent strategy and selective preservation. In the second part of the study, we compare the annual diatom assemblage composition and biogeochemical fluxes of Station PZB-1 with flux data documented in previous sediment trap studies conducted in other sectors of the Antarctic Zone in order to investigate how diatom floristics influence the composition and magnitude of particle fluxes in the Antarctic Zone. The lack of correlation between the annual diatom valve, organic carbon and biogenic silica fluxes across stations indicates that other factors aside from diatom abundance play a major role in the carbon and silica export in AZ. Among these factors, the composition of the diatom assemblage appears to be critical, as suggested by the strong and significant correlation between Bio-SiO₂ and the valve fluxes of *F. kerguelensis* alone, that this species is the main Bio-SiO₂ vector from the surface layer to the deep ocean in the AZ waters, regardless of its relative abundance. Lastly, the good correlation between the annual fluxes of the group of small *Fragilariopsis* species with satellite-derived chlorophyll-a concentration estimates over the study stations, suggest that high abundances of these species in the Southern Ocean paleorecords could be used as a proxy of high algal biomass accumulation.

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1 **Diatom species fluxes in the seasonally ice-covered Antarctic Zone: new data from**
2 **offshore Prydz Bay and comparison with other regions from the eastern Antarctic**
3 **and western Pacific sectors of the Southern Ocean.**

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15 **Abstract**

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19 composition in influencing the magnitude of the organic carbon and biogenic silica fluxes
20 exported out of the mixed layer in Southern Ocean ecosystems. Here we investigate the
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22 silica export and several significant environmental parameters using sediment trap
23 records deployed in different sectors of the Antarctic Zone. The study is divided in two
24 parts. We first present unpublished diatom species flux data collected by a sediment trap
25 in the offshore waters of Prydz Bay (Station PZB-1) over a year. The results of this study
26 revealed a major export peak of diatom valves in Austral summer and two small
27 unexpected secondary flux pulses during full winter conditions. The summer diatom
28 sinking assemblages were largely composed of small and rapidly dividing species such
29 as *Fragilariopsis cylindrus*, *Fragilariopsis curta* and *Pseudo-nitzschia lineola*, while

30 winter assemblages were dominated by *Fragilariopsis kerguelensis* most reflecting its
31 persistent strategy and selective preservation.

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33 and biogeochemical fluxes of Station PZB-1 with flux data documented in previous
34 sediment trap studies conducted in other sectors of the Antarctic Zone in order to
35 investigate how diatom floristics influence the composition and magnitude of particle
36 fluxes in the Antarctic Zone. The lack of correlation between the annual diatom valve,
37 organic carbon and biogenic silica fluxes across stations indicates that other factors aside
38 from diatom abundance play a major role in the carbon and silica export in AZ. Among
39 these factors, the composition of the diatom assemblage appears to be critical, as
40 suggested by the strong and significant correlation between Bio-SiO₂ and the valve fluxes
41 of *F. kerguelensis* alone, that this species is the main Bio-SiO₂ vector from the surface
42 layer to the deep ocean in the AZ waters, regardless of its relative abundance. Lastly, the
43 good correlation between the annual fluxes of the group of small *Fragilariopsis* species
44 with satellite-derived chlorophyll-*a* concentration estimates over the study stations,
45 suggest that high abundances of these species in the Southern Ocean paleorecords could
46 be used as a proxy of high algal biomass accumulation.

47

48 **1. Introduction**

49 **1. Southern Ocean phytoplankton**

50 The Southern Ocean is a crucial component of the global overturning circulation and
51 regulates the climate system through the uptake of heat, freshwater and atmospheric CO₂
52 (Sarmiento et al., 2004). Despite its relatively small size, it accounts for about 20% (0.47
53 GtCyr⁻¹) of the ocean CO₂ uptake flux (Takahashi et al., 2002). A significant fraction of
54 the CO₂ drawdown in the Southern Ocean waters is driven by phytoplankton (Siegel et
55 al., 2014)

56 Phytoplankton productivity is not sustained at its full capacity in the Southern Ocean in
57 spite of the fact that concentrations of most macronutrients remain ubiquitously high,
58 making the Southern Ocean the largest high-nutrient low-chlorophyll (HNLC) area of the
59 global ocean. Numerous studies have conclusively demonstrated that Fe-limitation plays
60 a critical role in restricting phytoplankton biomass and production within HNLC regions

61 of the Southern Ocean (Boyd and Law, 2001; Boyd et al., 2004; Coale et al., 2004 , among
62 others). Indeed, it is likely that iron availability had played an important role in the
63 variations of the atmospheric carbon dioxide levels over glacial cycles (Martínez-García
64 et al., 2009). Diatoms are the major primary producers in the Southern Ocean and are
65 often reported dominating high-productivity events in the Polar Frontal Zone, Antarctic
66 Zone (AZ) and coastal systems of Antarctica (e.g. Wilson et al., 1986; Bathmann et al.,
67 1997; Arrigo et al., 1999; Selph et al., 2001; Landry et al., 2002). Diatom blooms account
68 for a large fraction of the particulate organic carbon (POC) flux and for almost all of the
69 biogenic silica export out of the mixed layer. This export can be direct through the
70 formation of rapidly sinking aggregates of entangled cells and chains with fast sinking
71 rates (Boyd and Newton, 1999; Smetacek et al., 2012) or indirect via the production of
72 faecal material by zooplankton grazing of diatoms (Rembauville et al., 2014; Manno et
73 al., 2015; Belcher et al., 2016). The large accumulation of siliceous diatom remains in the
74 deep sea sediments between the winter sea ice edge and the Antarctic Polar Front is
75 responsible for the formation of a circumpolar Diatom Ooze Belt in the Southern Ocean
76 deep sea sediments (Burckle and Cirilli, 1987) that represents one of the most important
77 silica sinks in the world ocean (Tréguer, 2014).

78 As a more complete understanding of Southern Ocean ecosystems is developed,
79 it is becoming increasingly evident that the species composition of the plankton
80 communities plays a critical role in the regulation of ocean nutrient stoichiometry at
81 regional and global levels (e.g. Arrigo et al., 1999; Salter et al., 2014). More specifically,
82 recent studies provided conclusive evidence that the ecological traits and strategies of
83 different polar diatom species contribute to the regulation of the efficiency of the
84 biological pump and the degree of coupling of the carbon and silicon in the particles
85 sinking to the interior layers of the Southern Ocean (e.g. Salter et al., 2012; Assmy et al.,
86 2013; Rembauville et al., 2014; Rigual-Hernández et al., 2015b). Although ocean colour
87 satellites provide a circumpolar view of biological activity and some insights into the
88 fractional contribution of major phytoplankton functional groups (e.g. coccolithophores,
89 *Phaeocystis*-like, diatoms) to algal biomass accumulation (Alvain et al., 2008), they are
90 unable to resolve the species composition of phytoplankton communities and seasonal
91 species succession. *In situ* and year-round observations are therefore needed to refine the
92 interpretations based on satellite data, to relate surface chlorophyll to column-integrated
93 production and export and to determine the specific role of phytoplankton species in the

94 Southern Ocean ecosystems. Sediment trap mooring deployments are one direct method
95 of characterizing and quantifying the biogeochemical composition of particle fluxes and
96 allows for the determination of composition and seasonality of diatom sinking
97 assemblages. This technology is particularly useful in remote regions of the Southern
98 Ocean, such as those of AZ, which are often seasonally covered by sea ice.

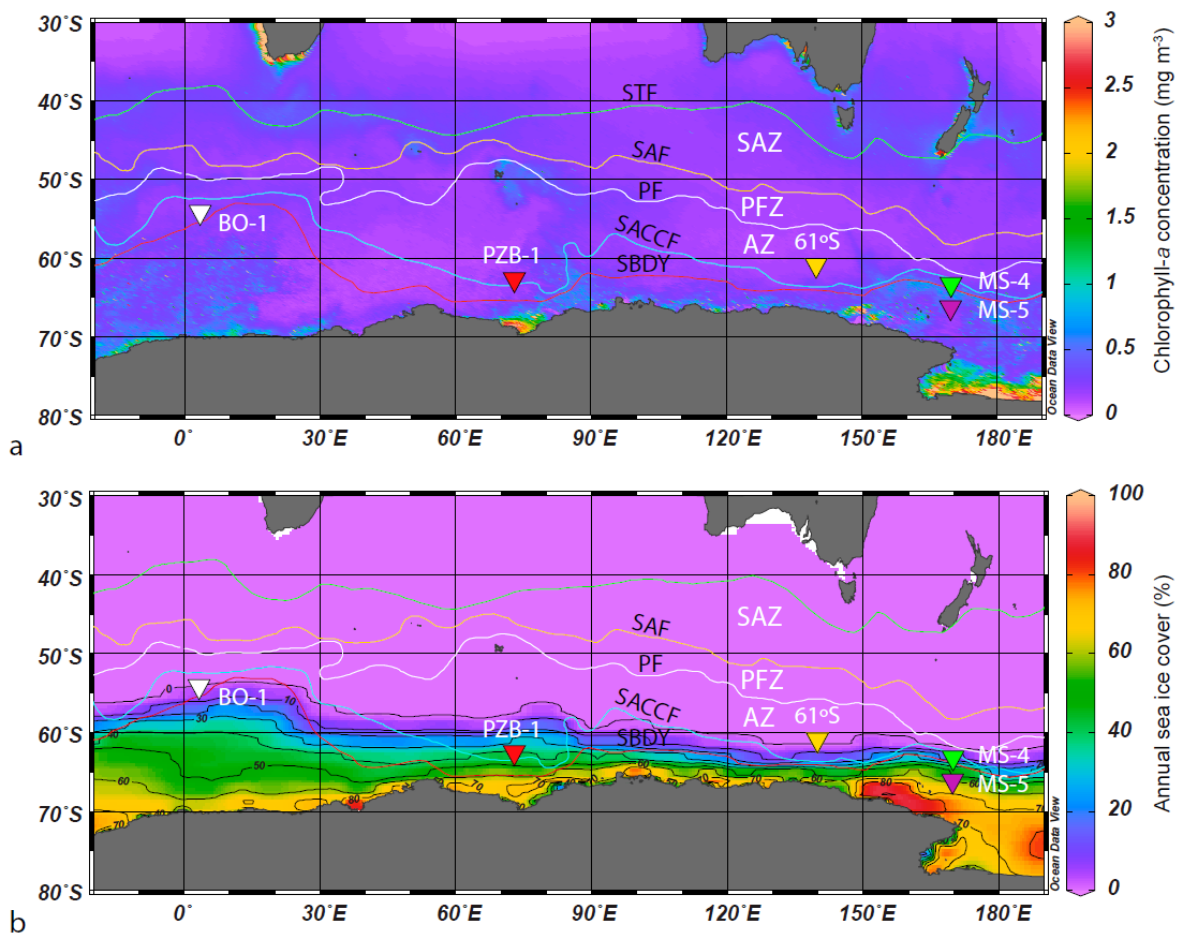
99 The main goal of this study is to investigate the relationship between the diatom
100 species flux assemblage, the POC and opal fluxes, and several significant environmental
101 parameters across key regions of the AZ of the Eastern Antarctic and Western Pacific
102 sectors of the Southern Ocean. The study is divided into two parts: firstly, we present
103 unpublished data on the diatom species fluxes intercepted by a sediment trap over a year
104 in the offshore waters of Prydz Bay. An improved understanding of the seasonality and
105 environmental preferences of diatom species captured by the trap will allow for better
106 interpretation of the paleorecords of the study region. Secondly, we compare the diatom
107 flux assemblage and biogeochemical fluxes of Station PZB-1 with already published
108 sediment trap data from other regions of the AZ. This comparison provides insight into
109 how diatom floristics influence the composition and magnitude of particle fluxes across
110 different sectors of the AZ.

111

112 **1.2 The Antarctic Zone**

113 The Southern Ocean is divided into a series of zonal systems characterized by
114 relatively uniform chemical and physical properties. These zonal systems are separated
115 by fronts that carry most of the transport of the Antarctic Circumpolar Current (ACC)
116 (Nowlin and Clifford, 1982). The AZ represents the southernmost zonal system of the
117 Antarctic Circumpolar Current and is delimited to the north by the Polar Front (PF) and
118 to the south by the Southern Boundary (SBDY; Fig. 1). South of Kerguelen, the AZ is at
119 its largest extent, while in other sectors it is only a few degrees wide (Fig. 1b). The AZ is
120 characterized by a well-mixed surface layer (down to 150 m in winter) of cold Winter
121 Water (WW) which is overlain by warmer and less saline Antarctic Surface Water
122 (AASW) in the summer (Orsi et al., 1995; Chaigneau et al., 2004). The Southern
123 Boundary of Upper Circumpolar Deepwater (SBDY or the Antarctic Divergence)
124 represents the southern limit of the AZ where the atmospheric wind regime reverses into
125 strong westerlies that drive a northward Ekman transport of deep, relatively warm and

126 nutrient-rich waters that upwell south of the APF (Speer et al., 2000; Pollard et al., 2006).
 127 The AZ can be broadly subdivided into a permanent ice-free zone (Permanently Open
 128 Ocean Zone, or POOZ) and a Seasonal Ice Zone (SIZ). The SIZ is limited in the north by
 129 the northern winter limit of the pack-ice and in the south by the northern summer limit of
 130 the pack-ice. The retreating sea ice edge during the summer exhibits one of the highest
 131 rates of primary production in the Southern Ocean (Arrigo et al., 2008) and accounts for
 132 an annual primary production of $86.7 \pm 12.6 \text{ Tg C yr}^{-1}$. Diatom production and export to
 133 the deep sea is thought to be largely responsible for the Si concentration gradient across
 134 the AZ (Dugdale and Wilkerson, 2001; Assmy et al., 2013), from $\sim 60 \mu\text{mol kg}^{-1}$ in the
 135 southern AZ due to the upwelling of Circumpolar Deep Waters, to $< 10 \mu\text{mol kg}^{-1}$ north
 136 the Antarctic Polar Front (Coale et al., 2004; Bostock et al., 2013).



137 **Figure 1. a** Chlorophyll-*a* composite map (1997 to 2012) from Sea-viewing Wide Field-
 138 of-View Sensor (SeaWiFS) of the Southern Ocean. **b.** Area-Averaged of Percentage sea
 139 ice cover monthly between 1997 and 2012 from the Giovanni NASA portal. Location of
 140 the sediment trap moorings in the AZ discussed in this article: PZB-1, BO-1, 61°S, MS-
 141 the Antarctic Polar Front (Coale et al., 2004; Bostock et al., 2013).
 142 4 and MS-5 stations (inverted triangles). Abbreviations: STF – Subtropical Front, SAZ –

143 Subantarctic Zone, SAF – Subantarctic Front, PFZ – Polar Frontal Zone, PF – Polar Front,
144 AZ – Antarctic Zone, Southern ACC Front – SACCF, Southern Boundary of Upper
145 Circumpolar Deepwater – SBDY. Oceanic fronts after Orsi et al. (1995).

146

147

148 **2. Material and Methods**

149 **2.1 Field programs**

150 **2.1.1 Offshore Prydz Bay trap experiment**

151 As part of a field research collaboration between the US and China polar research
152 programs, a mooring equipped with three sediment traps (1400, 2400 and 3400 m depth)
153 was deployed at 62°29'S; 72°59'E in the seasonal ice zone of the AZ north of Prydz Bay
154 (station PZB-1) from December 1998 to December 1999 (Pilskaln et al., 2004; Fig. 1).
155 All the sediment traps were McLane, Inc. time-series traps, with a 0.5 m² diameter
156 opening and equipped with 13 cups (Honjo and Doherty, 1988). Due to a timer failure,
157 no samples beyond cup 2 were collected by the 2400 m PZB-1 trap. Additionally, the
158 material mass collected in the 3400 m PZB-1 trap cups was too small to allow for diatom
159 counts after the priority geochemical analyses were completed (see Pilskaln et al. 2004
160 for details). Therefore, only data from the 1400 m is discussed here.

161 The sample cups were filled prior to the deployment with a 4% density-adjusted formalin
162 solution in filtered seawater buffered to a pH of 7.8-8.1 (Honjo et al., 2000; Pilskaln et
163 al., 2004). Cup rotation and collection intervals were established based on anticipated
164 mass fluxes. The sampling intervals ranged from 17 days in austral spring and summer to
165 a maximum collection interval of 41 days during the winter ice-cover month. A
166 presentation and discussion of the geochemical fluxes and major planktonic contributions
167 to settling particles at several trap depths in the offshore Prydz Bay region can be found
168 in Pilskaln et al. (2004). Annual export fluxes at 1400 m were dominated by Bio-SiO₂
169 that represented 73% of the annual mass export. A detailed description of the diatom
170 species analysis of the 1400 m time-series trap samples is provided in section 3.2.

171 **2.1.2 Other sediment trap experiments in the AZ between 0-180°E**

172 In the second part of the paper, biogeochemical and diatom species flux data from Station
173 PZB-1 are compared with already published datasets from four sediment trap
174 deployments (Table 1) in distinct settings of the AZ of the Southern Ocean (Fischer et al.,
175 2002; Grigorov et al., 2014; Rigual-Hernández et al., 2015a). Next, we summarize the
176 field experiments and the environmental conditions at each of the stations compared.

177 Station BO-1 (54° 20'S; 3° 23'E) was located in the eastern Atlantic Sector of the
178 Southern Ocean, close to the northernmost winter sea-ice edge and about 4° north of the
179 ACC-Weddell Gyre Boundary. The surface waters around station BO are characterized
180 by high macronutrient concentrations (Fischer et al., 2002), low iron levels (Loscher et
181 al., 1997) and low algal accumulation (Antoine et al., 1996; Arrigo et al., 2008). A
182 mooring line equipped with two sediment traps, placed at ~500 and 2200 m below the
183 surface (water column of 2700 m), was deployed over five almost consecutive years
184 (1990-95). Fischer et al. (2002) reported biogeochemical data for the five deployments
185 but diatom composition was only documented for year 1991. Bio-SiO₂ dominated the
186 mass fluxes year-round, representing 64% of the annual total mass flux. Station BO-1
187 was under the influence of sea ice for about three months, from August to October 1991.

188 Station 61°S (60° 44.43'S; 139° 53.97'E) was located in the Australian sector of the
189 southern AZ north of the Seasonal Ice Zone (Massom et al., 2013). Despite the high
190 macronutrient concentrations (silicate, nitrate and phosphate), the waters in this region
191 exhibit very low algal concentrations year-round (< 0.5 µg l⁻¹; Popp et al., 1999; Parslow
192 et al., 2001; Trull et al., 2001) most likely due to the very low iron concentrations (0.1-
193 0.2 nM; Boyd et al., 2000). Station 61°S was equipped for a year (November 2001 to
194 September 2002) with a mooring line with three sediment traps placed at 1000, 2000 and
195 3700 m below the surface in a water column of 4393 m. Due to equipment malfunction
196 no samples were recovered from the 1000 m trap. Since the rest of the sediment traps
197 compared here were deployed at shallower depths, only data from the shallowest trap
198 (2000 m) are discussed here. Export fluxes were largely dominated by Bio-SiO₂ that
199 accounted for 76% of the annual mass flux at 2000 m. A detailed description of the diatom
200 valve and biogeochemical fluxes can be found in Rigual-Hernández et al. (2015a).

201 As part of the US-JGOFS Antarctic Environmental Southern Ocean Process Study
202 (AESOPS; Smith Jr et al., 2000) and array of sediment traps was deployed along the
203 170°W parallel, in the western Pacific sector of the Southern Ocean (Honjo et al., 2000).
204 Two sediment trap mooring lines with sediment traps placed at ~1000 and 2000 m depth

205 were deployed during a year at Stations MS-4 (63° 09'S; 169° 54'W) and MS-5 (66°
 206 10'S; 169° 40'W), located south of the Antarctic Polar Front within the AZ. Because
 207 diatom species flux data were only available for the 1000 m traps, only data from these
 208 traps are discussed here. Station MS-4 was located south of the ACC while MS-5 was
 209 placed within the north of Ross Sea Gyre. The pelagic waters south of the Antarctic Polar
 210 Front in this region are characterized by high nutrient concentrations year-round and by
 211 the highest annual primary production values of the pelagic province of the Southern
 212 Ocean (Arrigo et al., 2008). Station MS-4 was under the influence of sea ice during about
 213 three months in winter, while the MS-5 mooring line was covered by ice during most of
 214 the year, except during three months in summer (Honjo et al., 2000). The episodic release
 215 of iron from the melting sea ice has been suggested as an important mechanism
 216 stimulating algal growth (predominantly diatoms) in the waters around Stations MS-4 and
 217 MS-5 (Sedwick and DiTullio, 1997; Grigorov et al., 2014). Detailed information on the
 218 biogeochemical and diatom flux data measured at the Stations MS-4 and MS-5 can be
 219 found in Honjo et al. (2000) and Grigorov et al. (2014), respectively. Annual export fluxes
 220 were dominated by Bio-SiO₂ at both Stations, accounting for 69% and 58% of the annual
 221 total mass flux at MS-4 and MS-5, respectively.

222

Station	Latitude	Longitude	Water column depth (m)	Trap depth (m)	Sampling interval start	Sampling interval end
BO-1	54° 20' S	3° 23' E	2734	450	28/12/1990	01/04/1992
PZB-1	62°29' S	72°59' E	4000	1400	30/12/1998	13/12/1999
61°S	60° 44' S	139° 54' E	4393	2000	30/11/2001	29/09/2002
MS-4	63° 09' S	169° 54' W	2885	1031	28/11/1996	24/12/1997
MS-5	66° 10' S	169° 40' W	3015	937	28/11/1996	24/12/1997

223

224 **Table 1.** Summary of mooring deployment locations, sediment trap depths and anchor
 225 depths.

226 2.2 Sample processing for diatom analysis

227 A detailed explanation of the sample preparation for diatom analysis for the PFZ-1
 228 is provided next, together with brief summary of the sample processing used in the other
 229 sediment trap experiments. One quantitative wet-split fraction from each of the 1400 m
 230 trap cups was designated for diatom analysis. The sample aliquot was gently rinsed and
 231 filtered with distilled water to remove salt and formalin and then treated with potassium

232 permanganate and hydrochloric acid for the removal of organic and calcareous material
 233 according to the procedures detailed in Schrader and Gersonde (1978) and Romero
 234 (1998). A 150-300 μl drop of the remaining suspended siliceous material was placed on
 235 a coverslip in a settling chamber, using the random settling technique described in Moore
 236 (1973). Once dry, the coverslips were mounted on glass slides using a high refractive
 237 index mountant (Naphrax) for light microscopy counts and identification of diatoms. The
 238 total flux of diatoms to each cup in valves $\text{m}^{-2} \text{day}^{-1}$ was calculated by multiplying the
 239 valve count by the area counted as a fraction of the total area and the dilution volume, all
 240 multiplied by the split and then divided by the collection period in days and the trap
 241 collection surface area (Sancetta and Calvert, 1988). The diatom species fluxes of the
 242 most abundant diatom species at PZB-1 are listed in Table 2 and plotted in Figure 2.

243

244 **Table 2:** Diatom species fluxes of the most abundant diatom species for the deployment
 245 PZB-1 (1400 m trap).

Cup number	Mid-point date	Collection days	Total diatom flux ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Fragilariopsis curta</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Fragilariopsis cylindrus</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Fragilariopsis kerguelensis</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Fragilariopsis pseudonana</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Fragilariopsis rhombica</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Pseudo-nitzschia lineola</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Thalassiosira gracilis</i> var. <i>expecta</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)	<i>Thalassiosira lentiginosa</i> ($10^6 \times \text{valves m}^{-2} \text{d}^{-1}$)
1	07/01/1999	17.0	128.6	15.8	35.5	16.6	3.7	8.3	14.6	4.9	0.9
2	25/01/1999	19.0	284.6	28.2	101.6	30.1	16.9	23.8	41.1	12.5	0.6
3	12/02/1999	17.0	37.2	2.6	9.0	9.0	0.7	3.6	2.5	2.5	0.2
4	01/03/1999	17.0	23.4	2.2	2.2	9.3	0.3	1.8	1.5	2.5	0.5
5	18/03/1999	17.0	1.5	0.1	0.0	0.9	0.0	0.1	0.0	0.1	0.1
6	16/04/1999	40.0	1.5	0.1	0.0	0.8	0.0	0.1	0.0	0.2	0.0
7	26/05/1999	41.0	1.4	0.1	0.0	0.6	0.0	0.1	0.0	0.2	0.0
8	06/07/1999	41.0	2.9	0.2	0.0	1.5	0.0	0.1	0.0	0.4	0.0
9	16/08/1999	41.0	33.5	1.0	0.1	21.2	0.0	1.1	0.0	2.3	0.4
10	26/09/1999	41.0	5.1	0.0	0.0	3.3	0.0	0.1	0.0	0.3	0.2
11	06/11/1999	40.0	24.2	2.2	3.2	11.1	0.2	0.4	0.1	1.3	1.7
12	04/12/1999	17.0	2.8	0.4	0.3	1.1	0.0	0.1	0.0	0.2	0.2
13	21/12/1999	17.0	3.9	0.3	0.2	2.1	0.0	0.1	0.0	0.1	0.4

246

247 Sample processing and diatom species examination for the Station BO-1 are described in
 248 detail in Fischer et al. (2002). In short, samples were prepared according to the
 249 methodology described in Gersonde and Zielinski (2000) and glass slides were counted
 250 following the recommendations described by Schrader and Gersonde (1978). Sediment
 251 trap samples from Station 61°S were acid-cleaned following the methodology of Romero
 252 et al. (1999, 2000). Microscopic slides were prepared following the random-settling
 253 decantation method described in Flores and Sierro (1997). The recommendations of
 254 Schrader and Gersonde (1978) were followed for counting diatom valves.

255 Samples from the Stations MS-4 and MS-5 were cleaned using sequential hydrogen
 256 peroxide digestions (Grigorov et al., 2014). Then, slides were prepared following the

257 method described by Sancetta (1992) and counted according to established conventions
258 (Fenner, 1991; Schrader and Gersonde, 1978)

259 **2.3 Estimation of annual diatom species contribution to total diatom valve flux**

260 Annual relative contribution of diatom species for all stations (Table 3 and Fig. 3) were
261 estimated calculating the integrated flux of each species for a 365-day period. Then, the
262 annual relative contribution of each species was estimated as the ratio of the annual valve
263 flux of a given species and the total diatom valve flux multiplied by one hundred. Due to
264 differences in the sampling duration of the five sediment trap experiments analyzed here,
265 some adjustments of the data sets were made to enable comparison between stations. The
266 PZB-1 sediment trap collected particles for a total of 365 days, therefore no corrections
267 were needed for this trap. Since the collection period for the BO-1 sediment trap was 368
268 days, the proportional flux of 3 days of the last cup was removed for our calculations. For
269 Station 61°S, the collection was shorter (309 days) than a calendar year, therefore annual
270 estimates were calculated. These annual estimates take into consideration that the
271 unsampled days occurred during winter when particle fluxes were low. The valve fluxes
272 of the last winter cup were used to represent mean daily fluxes during the unobserved
273 period (see Rigual-Hernández et al. 2015a for more details). Since MS-4 and MS-5
274 sediment traps sampled a total of 425 days, the data set was reduced to 365 days to enable
275 comparison with the rest of the stations.

276 **2.4 Environmental parameters**

277 The monthly products of total chlorophyll-*a* concentration and contribution of major
278 functional phytoplankton types to total chlorophyll-*a* for the five study stations for the
279 period 1998 and 2015 were obtained from the NASA Ocean Biogeochemical model
280 (NOBM) (accessed at <https://giovanni.gsfc.nasa.gov/giovanni/>). The data were extracted
281 for a half degree area around each mooring location. Primary productivity values (mg C
282 m⁻² d⁻¹) for all stations were extracted from the Ocean Productivity website (accessed at
283 www.science.oregonstate.edu/ocean.productivity/index.php/), which provides estimates
284 of net primary productivity derived by applying the standard vertically generalized
285 production model (VGPM; Behrenfeld and Falkowski, 1997) and the *Eppley*-VGPM
286 productivity models to SeaWiFS chlorophyll-*a* data.

287

288 2.5 Correlation analysis

289 In order to investigate the relationship between satellite and environmental parameters
290 (i.e. sea ice cover, chlorophyll-*a* and primary productivity estimates) and annual
291 biogeochemical and diatom fluxes measured by the traps, a correlation matrix was
292 calculated. However, given the fact that no satellite information was available for the
293 collection period of some experiments, some assumptions had to be made for this analysis
294 as detailed next.

295 The BO-1 sediment trap was deployed before Sea-WIFS satellite began scientific
296 operations (September 1997). Moreover, the majority of the sampling intervals of MS-4
297 and MS-5 traps took place before the commencement of the Sea WIFS Chlorophyll-*a*
298 data record. Therefore, no chlorophyll-*a* data was available for the collection intervals of
299 these three stations. Nonetheless, the coefficient of variation - i.e., the standard deviation
300 of the mean divided by the mean in percentage - of chlorophyll-*a* concentration for the
301 17 years analyzed here (from 1998 until 2015) indicates little inter-annual variability for
302 the three stations: 13%, 6% and 8% for the BO-1, MS-4 and MS-5, respectively.
303 Therefore, interannual average for these stations was considered representative of the
304 sediment-trap deployment years and used in the correlation analysis. Since the sediment
305 trap deployments of stations PZB-1 and 61°S took place after 1997, the annual average
306 chlorophyll-*a* concentration for the collection year (from September to September) was
307 used in the correlation analysis.

308 Since primary productivity data is only available from 1997, a similar approach than that
309 followed for the chlorophyll-*a* concentration was followed. In this case, the coefficient of
310 variation of Eppley model for Stations BO-1, MS-4 and MS-5 was 18%, 16% and 42%,
311 respectively. For the VGMP model, the coefficient of variation for Stations BO-1, MS-4
312 and MS-5 was 21%, 20%, and 42%. Given the high interannual variability of both primary
313 productivity models, the interannual average was not considered representative of the
314 collection interval and therefore, unacceptable for the correlation analysis.

315 Sea-ice data over the sediment trap deployment locations was obtained from Pilskaln et
316 al. (2004) for Station PZB-1, Fischer et al. (2002) for Station BO-1, Rigual-Hernández et
317 al. (2015a) for Station 61°S and Grigorov et al. (2014) for Stations MS-4 and MS-5.

318 Given the low sample size (five stations) and large differences in the magnitude of
319 variables compared, correlations were considered significant at p -value < 0.1 .

320

321 3. Results

322 3.1 PZB-1 diatom fluxes

323 The seasonality of diatom valve export at 1400 m at Station PFZ-1 closely
324 followed the temporal variability of total mass fluxes (Pilskałn et al., 2004) exhibiting
325 two distinct periods of enhanced diatom valve export (Fig. 2). The first and major peak
326 (up to 2.8×10^8 valves $\text{m}^{-2} \text{d}^{-1}$) occurred between late December 1998 and early February
327 1999, i.e. during the austral summer. Additionally, the sediment trap recorded two
328 unexpected peaks of particle export increase during the austral winter (August; up to 3.3
329 $\times 10^7$ valves $\text{m}^{-2} \text{d}^{-1}$) and spring (November; up to 2.4×10^7 valves $\text{m}^{-2} \text{d}^{-1}$) when the
330 mooring location was covered by seasonal sea ice (Fig. 2).

331 A total of 60 diatom species and subspecies were identified (Table 3). At 1400 m,
332 five species accounted for 75% of the annual diatom assemblage which was co-dominated
333 by *Fragilariopsis cylindrus* (25%) and *Fragilariopsis kerguelensis* (24%), followed by
334 *Pseudo-nitzschia lineola* (10%), *Fragilariopsis curta* (9%) and *Fragilariopsis rhombica*
335 (7%).

336 Most of the diatom species fluxes at 1400 m broadly followed similar seasonal
337 trends with the highest annual fluxes during the peak of the summer bloom period (late
338 December and January) and low or negligible fluxes during the remainder of the year
339 with the exception of the two short export pulses in August and November 1999 (Fig. 2b
340 and c). Nonetheless, some important differences can be seen among species. During the
341 early period of the summer bloom (January 1999), small-sized species of the genus
342 *Fragilariopsis*, such as *F. cylindrus*, *F. pseudonana* and *F. rhombica*, *F. curta* together
343 with *Pseudo-nitzschia lineola* rapidly increased their fluxes (three-fold, five-fold, three-
344 fold, two-fold and three-fold, respectively) dominating the sinking diatom assemblage.
345 The small *Fragilariopsis* species mentioned above represented 74% of the sinking diatom
346 assemblage by the end of January (Fig. 2d). The relative abundance of this group rapidly
347 decreased in the later stages of the summer bloom, representing 34% of the assemblage
348 by late February. *Fragilariopsis kerguelensis*, that has larger and more heavily silicified
349 valves than the rest of the members of the genus *Fragilariopsis*, with major contributions
350 identified in this study, was also an important contributor to the summer bloom. However,
351 its development during the bloom differed from that of former group of species in that

352 the contribution of *F. kerguelensis* to the sinking assemblage collected by the trap
353 increased steadily during the bloom, from ~10% in January to 40% by late February.

354 The two diatom valve export pulses documented in August and November
355 exhibited a different diatom assemblage composition than that of the summer bloom. The
356 winter peak (August) was mainly caused by an increased flux of *F. kerguelensis* that
357 represented 63% of the sinking diatom assemblage, followed by *Fragilariopsis*
358 *separanda* (8%) and the two varieties of *Thalassiosira gracilis*: *T. gracilis* var. *expecta*
359 (7%) and *T. gracilis* var. *gracilis* (5%) (plotted in Figure 2 as *Thalassiosira gracilis*
360 group). The main constituent of the November peak was also *F. kerguelensis* (46%), but,
361 interestingly the secondary components of the flux were mainly composed by the small
362 *F. cylindrus* (13%) and *F. curta* (9%), *Thalassiosira lentiginosa* (7%), *F. separanda* (6%)
363 and *T. gracilis* var. *expecta* (5%).

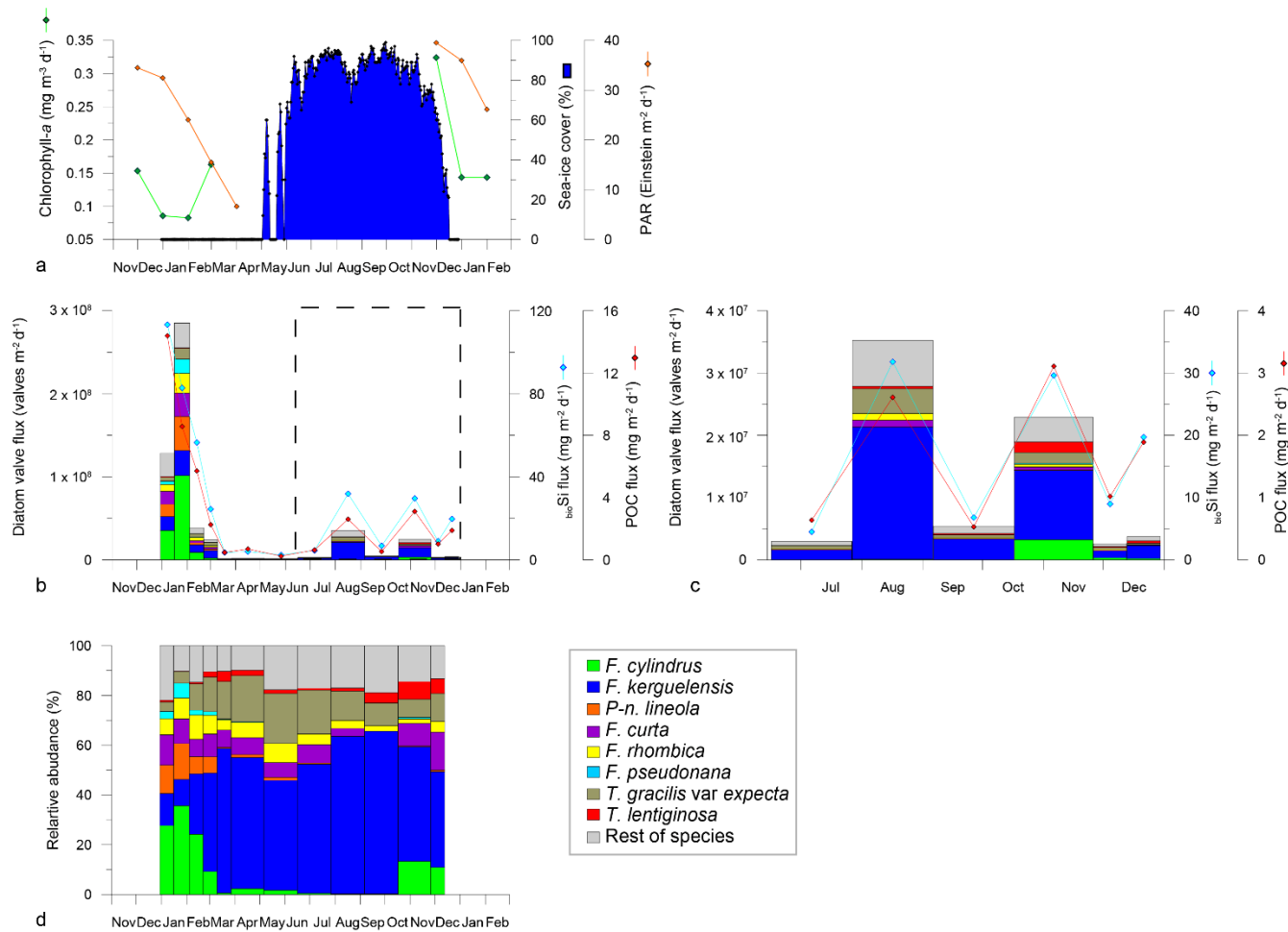
364 **3.2 Correlation analysis**

365 The results of the correlation matrix are presented in Table 4. Total chlorophyll-*a* is
366 positively correlated with annual total diatom ($r = 0.891$) and small *Fragilariopsis* group
367 fluxes ($r = 0.994$). Both annual *F. kerguelensis* and Bio-SiO₂ fluxes are negatively
368 correlated with the annual percentage of sea-ice coverage ($r = -0.960$ and -0.958 ,
369 respectively). Lastly, annual *F. kerguelensis* flux shows a positive correlation with annual
370 Bio-SiO₂ export ($r = 0.869$).

371

372

373



374

375 **Figure 2. a.** Satellite observations at PZB-1 location: monthly averaged chlorophyll-*a*
 376 concentration and photosynthetically available radiation (PAR) derived from Giovanni
 377 NASA website and daily percentage of sea ice cover over the trap (modified from Pilskaln
 378 et al., 2004) for the period November 1998 to February 1999. **b.** Annual variability of the
 379 total diatom, biogenic silica and POC fluxes at PZB-1, 1400 m (from Pilskaln et al.,
 380 2004). The dashed box represents the winter period, which is shown magnified in Figure
 381 2c. **c.** Annual variability of the total diatom, biogenic silica and POC fluxes plotted for
 382 June-December at PZB-1, 1400 m (from Pilskaln et al., 2004). **d.** Seasonal variability of
 383 the relative abundance of the main diatom taxa at PZB-1, 1400 m.

384

385 **Table 3.** List of diatom species and their annual relative contribution to total diatom flux
 386 recorded from time-series sediment trap studies in the AZ of the East Antarctic Southern
 387 Ocean: PZB-1 at 1400 m (this study), 61°S at 2000 m (Rigual-Hernández et al., 2015a),
 388 MS-4 at 1030 m, MS-5 at 930 m (Grigorov et al., 2014) and BO1 at 450 m (Fischer et al.,

389 2002). Relative abundances < 0.1 are represented by an asterisk (*), whereas the absence
390 of a taxon in a given station is represented by an empty circle (\circ). Only major species
391 (nine) are shown for station BO1.

Species	BOI	PZB-1	61°S	MS-4	MS-5	Species	BOI	PZB-1	61°S	MS-4	MS-5	
<i>Actinocyclus actinophilus</i> (Ehrenberg) Simonsen		0.12	*	*	*	<i>P. truncata</i> (G. Karsten) Nöthig et Ligowski		*	o	o	o	
<i>Actinocyclus curvatus</i> Janisch		*	o	*	o	<i>Proboscia</i> spp.		*	o	o	o	
<i>Actinocyclus exiguus</i> Fryxell et Semina		o	o	o	o	<i>Psammiodictyon panduriforme</i> (Gregory) Mann		o	o	o	o	
<i>Actinocyclus octonarius</i> Ehrenberg		o	o	*	o	<i>Pseudo-nitzschia</i> cf. <i>lineola</i>		o	0.4	o	o	
<i>Actinocyclus</i> spp.		o	*	*	*	<i>P-n. lineola</i> (Cleve) Hasle		9.57	o	2.30	3.18	
<i>Alveus marinus</i> (Grunow) Kaczmarek et Fryxell		o	o	o	o	<i>P-n. turgiduloides</i> (Hasle) Hasle		1.58	o	o	o	
<i>Asteromphalus hookeri</i> Ehrenberg		0.21	0.2	0.16	1.11	<i>P-n. prolongatoides</i> (Hasle) Hasle		0.22	o	o	o	
<i>A. hyalinus</i> Karsten		*	0.2	*	o	<i>P-n. heimit</i> Manguin		o	*	o	o	
<i>A. parvulus</i> Karsten		0.34	0.2	*	*	<i>Pseudo-nitzschia</i> spp.		*	0.1	o	0.13	
<i>Asteromphalus</i> spp.		o	*	o	o	<i>Rhizosolenia antennata</i> (Ehrenberg) Brown f. <i>antennata</i>		*	o	o	o	
<i>Apeitia subularis</i> (Grunow) Fryxell et Sims		0.33	*	0.7	0.15	<i>R. antennata</i> (Ehrenberg) Brown f. <i>semispina</i> Sundström		0.70	o	o	o	
<i>Banquetia belgicae</i> (Van Heurck) Padlock		0.38	o	o	o	<i>R. bergonii</i> Pergallo		o	o	o	o	
<i>Chaetoceros aequatorialis</i> var. <i>antarcticus</i> Manguin		o	*	o	o	<i>Rhizosolenia</i> cf. <i>costata</i>		o	o	o	o	
<i>C. atlanticus</i> Cleve		0.95	0.2	0.19	*	<i>Rhizosolenia</i> cf. <i>chunii</i>		0.23	o	o	o	
<i>C. criophilus</i> Castracane		o	o	*	*	<i>R. curvata</i> Zacharias		o	o	o	o	
<i>C. diadema</i> Ehrenberg		o	o	o	*	<i>R. polydactyla</i> Castracane f. <i>polydactyla</i>		o	o	o	o	
<i>C. dichæta</i> Ehrenberg		*	0.1	o	o	<i>R. polydactyla</i> Castracane f. <i>squamosa</i>		*	o	o	o	
<i>C. peruvianus</i> Brightwell		*	o	o	o	<i>R. simplex</i> Karsten		o	o	*	o	
<i>Chaetoceros</i> subgenus <i>Hyalochaete</i> spp.		0.49	0.2	*	0.13	<i>R. sima</i> Castracane		o	o	o	*	
<i>Chaetoceros</i> subgenus <i>Phaeoceros</i> spp.			0.2			<i>R. styliformis</i> Brightwell		o	o	*	o	
<i>Chaetoceros</i> resting spores		2.59	0.1	o	o	<i>Rhizosolenia</i> sp. f. 1A <i>sensu</i> Armand et Zielinski		*	*	o	o	
<i>Cocconeis</i> spp.		o	o	*	o	<i>Rhizosolenia</i> spp.		o	0.1	o	o	
<i>Corethron</i> sp.		0.25	*	*	*	<i>Roperia tessellata</i> (Roper) Grunow		o	o	o	o	
<i>Coscinodiscus asteromphalus</i> Ehrenberg		*	o	o	o	<i>Stellarima stellaris</i> (Roper) Hasle et Sims		o	o	o	o	
<i>Cyclotella</i> spp.		o	o	o	o	<i>Synedropsis</i> sp.		*	o	o	o	
<i>Dactylosolen antarcticus</i> Castracane		o	o	o	o	<i>Thalassionema nitzschioides</i> var. <i>capitulata</i> (Castracane) Moreno-Ruiz		o	0.1	o	o	
<i>Diploneis bombus</i> (Ehrenberg) Ehrenberg		o	o	o	o	<i>T. nitzschioides</i> var. <i>lancoolata</i> (Grunow) Pergallo et Pergallo		o	0.1	*	o	
<i>Eucampia antarctica</i> (Castracane) Manguin (summer form)		*	o	*	o	<i>T. nitzschioides</i> var. <i>parvum</i> Moreno-Ruiz		o	o	o	o	
<i>E. antarctica</i> (Castracane) Manguin (winter form)		o	0.1	o	o	<i>T. nitzschioides</i> var. 1 <i>sensu</i> Zielinski et Gersonde		3.45	o	o	* 0.16	
<i>Fragilariopsis curta</i> (Van Heurck) Hustedt		37.05	9.05	0.6	18.70	35.21	<i>Thalassiosira antarctica</i> Comber		*	o	* 0.09	
<i>F. cylindrus</i> (Grunow) Krieger		3.04	24.83	0.2	36.52	47.04	<i>T. decipiens</i> (Grunow ex Van Heurck) Jørgensen		o	o	o 0.17	
<i>F. doliohus</i> (Wallich) Medlin et Sims		o	o	o	o	<i>T. eccentrica</i> (Ehrenberg) Cleve		o	0.2	o	o	
<i>F. kerguelensis</i> (O'Meara) Hustedt		28.63	24.29	79.9	13.83	4.13	<i>T. ferlineata</i> Hasle et Fryxell		o	o	o	
<i>F. obliquecostata</i> (van Heurck) Heiden		*	*	*	0.16	<i>T. gracilis</i> var. <i>expecta</i> (Van Landingham) Fryxell et Hasle		5.20	0.4	7.09	o	
<i>F. pseudonana</i> (Hasle) Hasle		3.55	2	12.05	2.96	<i>T. gracilis</i> var. <i>gracilis</i> (Karsten) Hustedt		1.40	3.6	o	o	
<i>F. rhombica</i> (O'Meara) Hustedt		1.47	6.67	0.9	1.43	0.79	<i>T. gracilis</i> group		o	4.1	o 1.67	
<i>F. ritscherii</i> Hustedt		0.94	0.1	0.73	1.33	<i>T. gravis</i> Cleve		6.97	0.43	* 0.14	0.12	
<i>F. separanda</i> Hustedt		2.72	1.76	2.1	4.53	0.18	<i>T. lentiginosa</i> (Janisch) Fryxell		2.79	1.26	5 0.81	0.27
<i>Fragilariopsis sublinearis</i> (Van Heurck) Heiden et Kolbe		0.57	o	o	o	<i>T. leptopus</i> (Grunow ex Van Heurck) Hasle et Fryxell		o	*	o	o	
<i>F. cf. sublineata</i> (Van Heurck) Heiden		o	*	o	o	<i>T. lineata</i> Jousé		o	o	o	o	
<i>Fragilariopsis vanheurckii</i> (Pergallo) Hustedt		0.26	o	o	o	<i>T. maculata</i> Fryxell et Johans		o	*	o	o	
<i>Fragilariopsis</i> spp.		o	o	o	o	<i>T. ostrupii</i> (Ostenfeld) Hasle var. <i>ostrupii</i> Fryxell et Hasle		*	*	o 0.13	0.02	
<i>Gubardia</i> spp.		o	o	o	*	<i>T. ostrupii</i> (Ostenfeld) Hasle var. <i>wenckae</i> Fryxell et Hasle		o	o	o	o	
<i>Gyrosigma</i> spp.		o	o	o	o	<i>T. oliveriana</i> (O'Meara) Makarova et Nikolaev		*	0.7	o	0.07	
<i>Haslea trompii</i> (Cleve) Simonsen		0.16	*	o	o	<i>T. symmetrica</i> Fryxell et Hasle		o	o	o	o	
<i>Hemidiscus cuneiformis</i> Wallich		o	o	o	o	<i>T. trifulta</i> Fryxell		0.11	o	o	o	
<i>Manguinea</i> spp.		*	o	o	o	<i>T. tumida</i> (Janisch) Hasle		*	0.1	o	o	
<i>Membranella</i> spp.		0.33	o	o	o	<i>Thalassiosira</i> sp. 1		o	*	o	o	
<i>Navicula directa</i> (Smith) Ralfs in Pritchard		0.23	0.3	o	o	<i>Thalassiosira</i> sp. 2		o	o	o	o	
<i>Navicula</i> spp.		*	o	0.55	0.55	<i>Thalassiosira</i> sp. 3		o	o	o	o	
<i>Nitzschia bicapitata</i> Cleve		o	o	o	o	<i>Thalassiosira</i> eccentric group		o	o	o	o	
<i>N. braarudii</i> (Hasle)		o	o	o	o	<i>Thalassiosira. linear</i> group		o	0.1	o	o	
<i>N. kolaczekii</i> Grunow		o	o	o	o	<i>T. trifulta</i> group		o	o	o	o	
<i>N. sicula</i> (Castracane) Hustedt var. <i>bicuneata</i> Grunow		o	0.1	o	o	<i>Thalassiosira</i> spp. < 20 µm		o	0.4	*	o	
<i>N. sicula</i> (Castracane) Hustedt var. <i>rostrata</i> Hustedt		0.10	o	o	o	<i>Thalassiosira</i> spp. > 20 µm		o	*	o	o	
<i>Nitzschia</i> spp.		o	o	o	o	<i>Thalassiothrix antarctica</i> Schimper ex Karsten		o	0.2	0.20	0.23	
<i>Paralia</i> spp.		0.04	o	o	o	<i>Thalassiothrix longissima + antarctica</i>		0.46	o	o	o	
<i>Pleurosigma</i> spp.		o	*	0.01	o	<i>Trachyneis aspera</i> (Ehrenberg) Cleve		o	o	o	o	
<i>Pleurosigma directum</i>		0.03	o	o	o	<i>Trichotoxon reinboldii</i> (Van Heurck) Reid et Round		*	o	*	0.10	
<i>Porosira pseudodenticulata</i> (Hustedt) Jousé		o	*	o	o	<i>Tropidoneis</i> group		o	*	o	o	
<i>Proboscia alata</i> (Brightwell) Sundström		*	o	o	o	Other centrics		*	*	o	o	
<i>P. inermis</i> (Castracane) Jordan et Ligowski		*	o	o	o	Other pennates		*	0.1	o	o	

394

395 **Table 4.** Correlation matrix (r) for the main environmental parameters, diatom and
 396 biogeochemical fluxes measured at the study stations. Correlation in red are significant
 397 at $p < 0.1$.

	Total Chl- <i>a</i> concentration	Sea ice cover	Total diatom flux	<i>F. kerguelensis</i> flux	Small <i>Fragilariopsis</i> group flux	Biogenic Silica flux	POC flux	POC flux normalized at 2000m
Total Chl- <i>a</i> concentration	1.000 p= ---							
Sea ice cover	-0.029 p=.964	1.000 p= ---						
Total diatom flux	0.891 p=.043	-0.433 p=.467	1.000 p= ---					
<i>F. kerguelensis</i> flux	0.218 p=.724	-0.960 p=.010	0.588 p=.297	1.000 p= ---				
Small <i>Fragilariopsis</i> group flux	0.944 p=.016	-0.235 p=.703	0.977 p=.004	0.409 p=.494	1.000 p= ---			
Biogenic Silica flux	-0.056 p=.929	-0.958 p=.010	0.379 p=.529	0.869 p=.056	0.199 p=.749	1.000 p= ---		
POC flux	0.236 p=.702	-0.055 p=.930	0.381 p=.527	-0.008 p=.990	0.433 p=.466	0.297 p=.627	1.000 p= ---	
POC flux normalized at 2000m	0.419 p=.482	-0.637 p=.247	0.730 p=.162	0.787 p=.114	0.650 p=.236	0.588 p=.297	0.244 p=.692	1.000 p= ---

398

399

400 5. Discussion

401 5.1 Seasonal drivers of diatom community development and valve fluxes at PZB-1

402 Increasing irradiance and air temperature in springtime, along with sea ice melting
 403 processes, resulted in a moderate increase in the algal biomass, as inferred from the
 404 satellite derived chlorophyll-*a* concentration (Fig. 2a). As sea ice melts, it releases
 405 nutrients and dust particles intercepted during ice formation that stimulate phytoplankton
 406 growth (Sedwick and DiTullio, 1997; van der Merwe et al., 2011). Partitioning of the
 407 chlorophyll-*a* signal between major phytoplankton groups by the NASA Ocean
 408 Biogeochemical Model (NOBM) suggests that diatoms largely dominate the
 409 phytoplankton communities at the study station, accounting for > 99% of the chlorophyll-
 410 *a* production (Fig. 4). The high contribution of diatoms to the total chlorophyll-*a*
 411 concentration at PZB-1 station most likely represents an overestimation of their real
 412 abundance as a result of the NOBM limitations (Hirata et al., 2011; Rousseaux and Gregg,
 413 2012). Nonetheless, these results reinforce the idea that diatoms dominate the
 414 phytoplankton assemblages in the PZB-1 region and are consistent *in situ* observations of
 415 phytoplankton community composition in pelagic Southern Ocean systems under the
 416 influence of recent sea ice melt (e.g. DiTullio and Smith, 1996; Kang et al., 2001) where
 417 mix layers are often shallow and stratified. These conditions favor the development of

418 diatoms over *Phaeocystis antarctica*, a colonial haptophyte known to regularly develop
419 large blooms in seasonal ice zones and coastal Antarctic waters with deep mixed layer
420 due to its ability to sustain near-maximal photosynthetic rates at much lower solar
421 irradiance levels than do diatoms (Arrigo et al., 1999; Kropuenske et al., 2010; Mills et
422 al., 2010).

423 Diatom export at PZB-1 1400 m basically followed the seasonal trend of
424 chlorophyll-*a* concentrations in the surface layer as documented by monthly composites
425 of SeaWifs data from the study region and summarized in Pilskalns et al. (2004).

426 About two thirds of the annual diatom valve export occurred during a short
427 interval of 36 days (Dec. 30 - Feb. 4; Pilskalns et al., 2004). The pronounced diatom valve
428 export was coupled with a strong pulse of organic carbon and biogenic silica that
429 accounted for about half of the annual carbon and silica export at 1400 m (Pilskalns et al.,
430 2004). This pronounced and short-lived pulse of diatom production and export is a
431 common feature in the circumpolar AZ (Fischer et al., 2002; Grigorov et al., 2014;
432 Rigual-Hernández et al., 2015a) and likely the primary driver of atmospheric CO₂
433 sequestration in this zonal region of the Southern Ocean.

434

435 **5.2 Diatom species succession at 1400 m at PZB-1**

436 Ice melting processes in SIZ systems have been suggested to seed the surface
437 waters with phytoplankton cells accumulated in the sea ice that grow in the stable surface
438 waters after their release (Mangoni et al., 2009; Riaux-Gobin et al., 2011). This situation
439 seems to be the case for the PFZ-1 system as suggested by the large contribution of
440 *Fragilariopsis cylindrus* and *Fragilariopsis curta* to the summer bloom, as both species
441 have been reported to thrive abundantly in the waters adjacent to the ice and in the ice
442 itself (Kang and Fryxell, 1992; Scott et al., 1994; Leventer, 1998; Kang et al., 2001;
443 Riaux-Gobin et al., 2011). Despite the large contribution of sea-ice affiliated species to
444 the annual bloom, the strong contribution of open ocean species (mainly represented by
445 *F. kerguelensis*, *Pseudo-nitzschia lineola* and *Thalassiosira gracilis* group) also suggests
446 a significant input of pelagic phytoplankton communities over station PZB-1.

447 Regardless of their sea-ice preference, most of the diatom species contributing to
448 the peak in valve flux at PZB-1 are characterized by a boom-and-bust life strategy (i.e. r-

449 selected strategists). This group of diatoms, mainly represented by small *Fragilariopsis*
450 species and *Pseudo-nitzschia lineola*, posses a suite of functional traits such as small cell
451 sizes, weakly silicified frustules and high rates of nutrient acquisition, growth and
452 reproduction (Assmy et al., 2007; Durkin et al., 2012) that favor their rapid development
453 at the initial stages of the diatom bloom (Quéguiner, 2013). The high abundance of the
454 needle-shaped *Pseudo-nitzschia lineola* in the PZB-1 traps is consistent with previous
455 studies that reported species of this genus as one of the major components of diatom
456 blooms in the pelagic waters of the ACC (Hasle and Syvertsen, 1997; Kopczynska et al.,
457 2001; Smetacek et al., 2002; Assmy et al., 2007). Moreover, *Pseudo-nitzschia* species
458 represent important vectors of both carbon and silica flux to the deep ocean since they
459 have been reported to reach meso- and bathypelagic depths of the water column with a
460 significant fraction of their cellular organic content intact (Smetacek et al., 2012;
461 Rembauville et al., 2014).

462 The three- to four-fold increase in *F. cylindrus*, *F. rhombica* and *F. pseudonana*
463 fluxes during the growth phase of the bloom (i.e. from cup 1 to 2) and rapid decline in
464 their relative contribution from cup 3 are in contrast with the less pronounced two-fold
465 flux increase of *F. kerguelensis* but gradual increase in its relative abundance from mid-
466 summer throughout autumn (Fig. 2). The distinct seasonal trend of *F. kerguelensis* flux
467 suggests an ecological strategy different than the rest of the main components of the
468 diatom bloom. *Fragilariopsis kerguelensis* life strategy has been described as “persistent”
469 by Assmy et al. (2013) based on the lower growth rates of this species compared to that
470 of other major components of the bloom and its strong mechanical protection (Hamm et
471 al., 2003) against the heavy zooplankton grazing in the ACC (Pollard et al., 2002;
472 McLeod et al., 2010).

473 The two small flux peaks measured by the traps occurring in August and October-
474 November were unexpected given the fact that sea ice coverage started to build-up in
475 May, covering the mooring location completely from June until November (Fig. 2a).
476 Based on the large contribution of *F. kerguelensis* in the first peak, Pilskaln et al. (2004)
477 attributed the August pulse to an injection of material advected laterally into the trap from
478 an area of open, ice-free water. Indeed, NOAA Advanced Very High Resolution
479 Radiometer (AVHRR) imagery suggests the incursion of winds from mid-latitudes over
480 the study region between the 15th and 19th of August (supplementary Figure 1a). These
481 winds could have helped to propel warmer surface water masses towards the Station PZB-

482 1. Additionally, SSM/I ice concentration images from the National Snow and Ice Data
483 Center (supplementary Figure 1b) support this interpretation indicating the reduction of
484 sea ice concentration over the Station PZB-1 during a period of 12 days in mid-August,
485 probably as a result of the southward injection of warmer waters from this major
486 atmospheric event. This idea is further supported by the rest of the diatom assemblage
487 collected by the trap that is mainly composed of open ocean species, such as the
488 *Thalassiosira gracilis* group (12%), considered a cool open ocean taxon with peak
489 abundances within the maximum winter sea-ice edge (Crosta et al., 2005), and
490 *Fragilariopsis separanda* (8%) reported to display a similar distribution to *F.*
491 *keruelensis* in the Indian Ocean (Mohan et al., 2006). Based on this evidence, we
492 speculate that first winter export peak could have been the result of the transport of an
493 ice-free water masses by a major atmospheric event over the mooring location.

494 The secondary export pulse in all components measured in late October through
495 late November likely had a different origin than the August peak. The onset of the sea-
496 ice retreat around the PZB-1 trap in November together with the increase in light levels
497 resulted in the initiation of the phytoplankton bloom as suggested by the increase in
498 chlorophyll-*a* concentration (Fig. 2). The development of a diatom bloom under sea-ice
499 conditions is consistent with previous investigations that have reported large under-ice
500 phytoplankton blooms in polar environments (e.g. Arrigo et al., 2012; Lowry et al., 2018;
501 Nomura et al., 2018) The high contribution of the sea-ice affiliated species *F. cylindrus*
502 and *F. curta* (13% and 9%, respectively) during the November supports the idea of an
503 early initiation of the phytoplankton bloom triggered by the receding sea ice as previously
504 suggested by Pilskalns et al. (2004).

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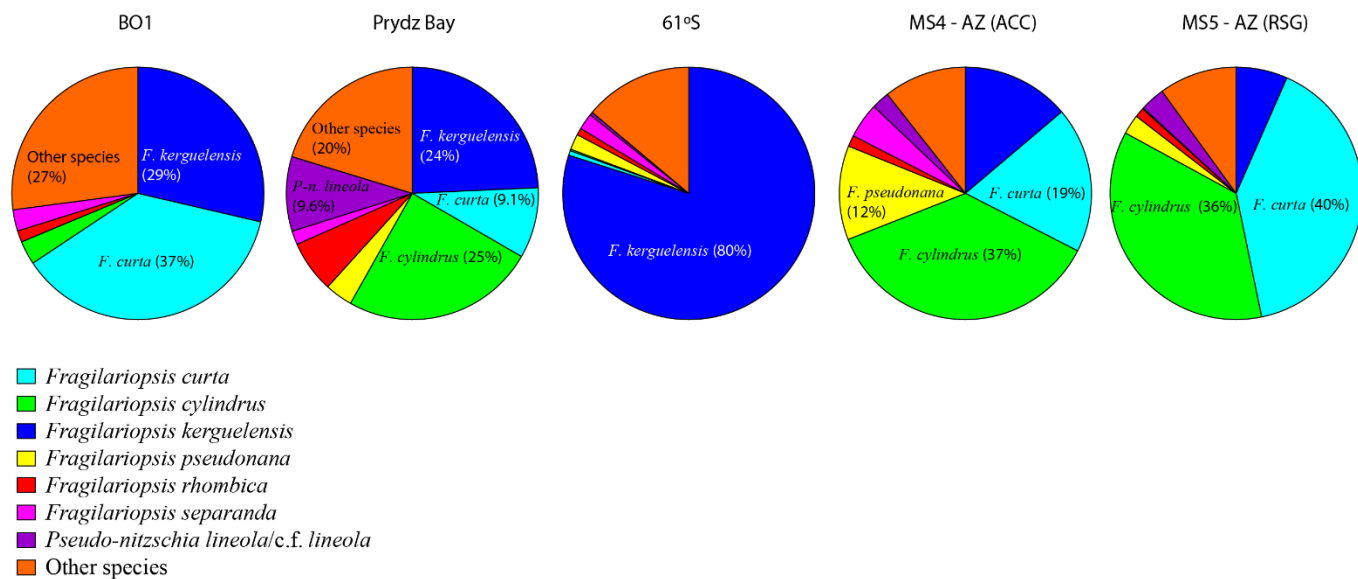
506 **5.3 Distribution of diatom species across different sectors of the Antarctic Zone**

507

508 Diatom assemblages at all stations were overwhelmingly dominated by species of
509 the genus *Fragilariopsis* with secondary contribution by *Pseudo-nitzschia lineola*/cf.
510 *lineola* (Fig. 3). Despite the fact that these species are often important contributors to the
511 living diatom assemblages in the surface layer in the AZ, they are most likely
512 overrepresented in the sediment traps due to selective dissolution in the upper water
513 column and mechanical breakage by zooplankton of weakly silicified species that can

514 cause profound modifications in the original diatom assemblage (McMinn, 1995; Jordan
 515 and Stickley, 2010; Rigual-Hernández et al., 2016). The variations in the composition of
 516 the diatom sinking assemblages across the stations seem to be largely determined by the
 517 influence of sea ice cover, a critical factor determining the distribution of diatom
 518 assemblages in the modern sediments of the Southern Ocean (Zielinski and Gersonde,
 519 1997; Armand et al., 2005). Maximum annual fluxes and relative contribution of *F.*
 520 *kerguelensis* are observed at Station 61°S, the only station not affected by sea ice, while
 521 minimum fluxes and relative contribution of this species are displayed at the MS-5
 522 station, characterized by the highest annual sea ice cover of all stations (Figs. 3 and 4).
 523 The negative effect of sea ice on *F. kerguelensis* distribution is supported by the negative
 524 and significant correlation between both variables (Table 4). Our observation on sediment
 525 trap fluxes is consistent with many previous reports on diatom assemblages from
 526 Southern Ocean sediments that described high abundance of this species as a proxy for
 527 the iron limited waters of the ACC (Burckle and Cirilli, 1987; Taylor et al., 1997; Crosta
 528 et al., 2005; Pike et al., 2008, among others).

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Figure 3. Annual relative contribution of the major diatom species recorded in AZ sediment traps of the East Antarctic Southern Ocean.

535 In contrast, *Fragilariopsis curta* and *Fragilariopsis cylindrus* display peak
536 relative abundances at those stations under the seasonal influence of sea ice cover (Fig. 3
537 and 4). The sea-ice affinity of these species is consistent with their biogeographical
538 distribution in the modern Southern Ocean where they are often found in high numbers
539 in the surface waters near the marginal ice-edge zones (e.g. Kang and Fryxell, 1992; Kang
540 et al., 2001), as part of sea ice communities (Scott et al., 1994; Ugalde et al., 2016) and
541 in the surface sediments in regions under the influence of sea ice (Zielinski and Gersonde,
542 1997; Armand et al., 2005). *Fragilariopsis pseudonana* exhibited its highest relative
543 contribution at MS-4, a station also characterized by peak diatom biomass accumulation
544 and diatom valve fluxes of all stations. Although *F. pseudonana* is often reported as an
545 important contributor to the phytoplankton communities in the Southern Ocean waters
546 (Kang and Lee, 1995; Villafañe et al., 1995; Kopczynska et al., 2001; Kopczyńska et al.,
547 2007; Cefarelli et al., 2010), it is often not preserved in the sedimentary record (Grigorov
548 et al., 2014) most likely due to its weak silicification. High diatom biomass accumulation
549 is known to facilitate mass aggregation, increase sinking rates and promote the deep-
550 water delivery of diatom frustules (Alldredge and Goltschalk, 1989; Passow et al., 2003).
551 Therefore, it is possible that the relatively high abundance of *F. pseudonana* at MS-4 trap
552 was due to high diatom biomass accumulation in the upper surface waters with subsequent
553 aggregation and sinking, thus increasing the probability of *F. pseudonana* transiting
554 through the mixed layer and reaching the sediment trap depth intact. Lastly, it is important
555 to note that annual fluxes of the small *Fragilariopsis* species group - composed by *F.*
556 *curta*, *F. cylindrus*, *F. rhombica* and *F. separanda* – display a robust and significant
557 correlation with total chlorophyll-*a* concentration ($r = 0.944$, $p = 0.016$) which highlights
558 the potential of this group species as a proxy of high algal biomass accumulation in
559 Southern Ocean paleorecords. Nonetheless, we acknowledge that caution should be taken
560 in the interpretation of the results of our correlation matrix because low sample size of
561 the current study limits ability to make strong statistical statements.

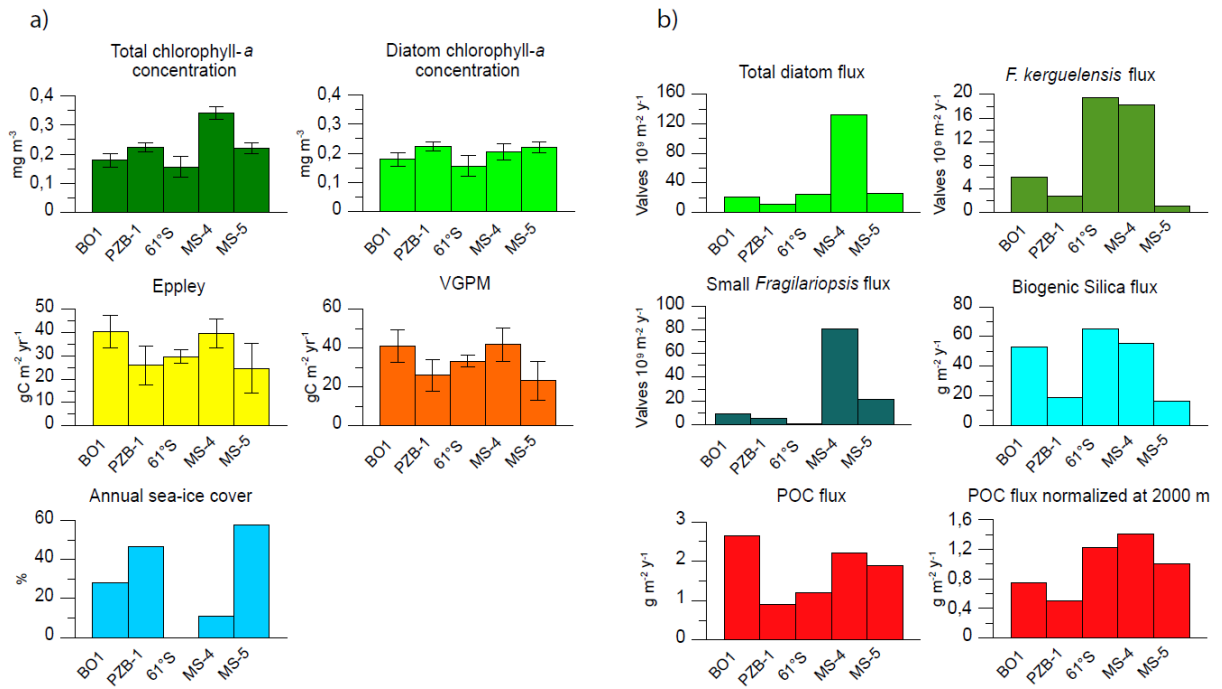
562

563 The genus *Pseudo-nitzschia* is cosmopolitan in the Southern Ocean waters but their
564 frustules are often not represented in the sedimentary record presumably due to
565 dissolution at the sediment-water interface and in the uppermost bottom sediments
566 (Rigual-Hernández et al., 2016). Nonetheless, despite their poor preservation *Pseudo-*
567 *nitzschia* species have been reported in the surface sediments of the Pacific and Prydz Bay

568 sectors of the AZ (Taylor et al., 1997; Grigorov et al., 2014) which coincide with stations
 569 displaying peak relative abundance of species of this genus (i.e. MS-4, MS-5 and PZB-1;
 570 Fig. 3).

571

572



573

574 **Figure 4.** Antarctic Zone station comparisons: BO1, PZB-1, 61°S, MS-4 and MS-5. **a.**
 575 Annual average total chlorophyll-*a* concentration, diatom chlorophyll-*a* concentration,
 576 two model estimates of primary productivity (VGPM and Eppley), and percent sea ice
 577 cover for the 5 stations obtained from the NOBM model. **b.** Annual fluxes of total diatom
 578 valves, major diatom species (*F. kerguelensis* and small *Fragilariopsis* group*),
 579 biogenic silicate, POC at sediment trap depths (BO-1, 450 m; PZB-1, 1400 m; 61°S, 2000
 580 m; MS-4, 1031 m; MS-5, 937) and 2000 m-normalized POC (after Honjo et al., 2008).
 581 *Small *Fragilariopsis* group includes *F. curta*, *F. cylindrus*, *F. rhombica* and *F.*
 582 *separanda*.

583

584 **5.4 Variations in annual biogenic particle export between sectors of the Antarctic**
 585 **Zone particle composition and their relationship with diatom assemblages**

586

587 All the sediment traps discussed in this study (Table 1) were deployed far below
588 the ventilation depths of the AZ (approximately 100 m; Trull et al., 2001), thereby
589 providing a direct measurement of the annual extraction of carbon from the atmosphere
590 by biological processes. In order to objectively compare the AZ trap data sets from
591 different mesopelagic depths, annual POC fluxes were normalized to 2 km after the Honjo
592 et al. (2008) study using the Berelson (2001) empirical POC flux reduction formula with
593 a power constant of 0.87 (Figure 4). Taking into account that the majority of biogenic
594 silicate dissolution occurs in the upper water column with minimal dissolution loss in the
595 mesopelagic zone (Ragueneau et al., 2002; Rigual-Hernández et al., 2015a; Rigual-
596 Hernández et al., 2016), diatom valve and biogenic silicate fluxes were not normalized.

597 Annual Bio-SiO₂ fluxes at mesopelagic depths at the five AZ stations were
598 between 2 to 9 times higher than the mean opal export fluxes at the 2 km
599 mesopelagic/bathypelagic boundary in the global ocean of 6.9 g SiO₂ m⁻² yr⁻¹ reported in
600 Honjo et al. (2008). The peak Bio-SiO₂ fluxes measured in the Australian and New
601 Zealand sectors of the AZ (i.e. 61°S and MS-4 stations, respectively) represent two of the
602 largest annual opal exports ever measured in the global ocean (Honjo et al., 2008; Rigual-
603 Hernández et al., 2015b) rivalled only by the North Pacific Boreal Gyres (48 g SiO₂ m⁻²
604 yr⁻¹ at the Aleutian-Bering Station; Takahashi et al., 2000). The large Bio-SiO₂ export
605 fluxes in the pelagic AZ are linked to the vicinity of the Antarctic Divergence where the
606 upwelling of deep nutrient-rich waters (Pollard et al., 2006) enhance diatom growth and
607 facilitate opal preservation in underlying sediments. Given the fact that diatoms are by
608 far the main biogenic silica exporters in the AZ waters (Grigorov et al., 2014; Rigual-
609 Hernández et al., 2015a), the weak and non-significant correlation between annual diatom
610 valve and Bio-SiO₂ fluxes ($r = 0.379$, $p = 0.529$) is somewhat surprising. This poor
611 correlation is most likely due to the large differences in biogenic silica (BSi) content per
612 cell of the dominant diatom species at each location. Particularly important is the
613 pronounced difference in BSi content between the different species of the genus
614 *Fragilariopsis* that dominated the sinking assemblages at all stations. While diatom
615 assemblages at Station 61°S were mainly composed of the relatively large and heavily
616 silicified *Fragilariopsis kerguelensis* (80% of the annual diatom export), MS-4 and MS-
617 5 assemblages were dominated by *Fragilariopsis curta* (19% and 35%, respectively) and
618 *Fragilariopsis cylindrus* (37% and 47% at both stations) characterized by smaller cell
619 sizes and weak silicification. Indeed, when comparing Bio-SiO₂ and valve fluxes of *F.*

620 *kerguelensis* alone, they yield a much stronger correlation ($r = 0.869$, $p = 0.056$)
621 suggesting that *F. kerguelensis* is the main vector of BSi to the deep ocean at all the study
622 stations regardless its relative contribution to the total assemblage. This result highlights,
623 once again, the critical role of *F. kerguelensis* in the silica cycle of the Southern Ocean,
624 particularly in the AZ where it is the main species responsible for the formation of the
625 silica-rich deposits that encircle Antarctica between the winter sea ice edge and the
626 Antarctic Polar Front (Burckle and Cirilli, 1987; Crosta et al., 2005).

627 POC export fluxes at 2000 m varied across the AZ stations with values about two-fold
628 lower in the Eastern Atlantic (Station BO 1) and offshore Prydz Bay (Station PZB-1) than
629 to those observed in the Western Pacific sector, where station MS-4 displayed the highest
630 POC fluxes of all stations ($1.4 \text{ g C m}^{-2} \text{ yr}^{-1}$; Fig. 4). The high POC export fluxes at the
631 MS-4 coincide with the highest chlorophyll-*a* concentration and primary productivity
632 estimates of all stations. These results are in agreement with Arrigo et al. (2008) who
633 described the Ross Sea sector as the most productive region of the Southern Ocean and
634 with Nelson et al. (2002) who documented high algal accumulation and high POC export
635 fluxes around the MS-4 deployment site. The temporal variations between the POC, Bio-
636 SiO₂ and diatom fluxes were strongly correlated at all stations (Fischer et al., 2002;
637 Grigorov et al., 2014; Rigual-Hernández et al., 2015a and this study). However, when the
638 magnitude of the annual fluxes of these three parameters is compared (Fig. 4), the
639 correlation between them is not significant (Table 4), indicating that the amount of
640 organic carbon sequestered to the ocean interior across the AZ is not directly proportional
641 to the amount of opal or diatom cells sinking out the mixed layer. These results suggest
642 that although diatoms represent a major vector of organic carbon to the ocean interior in
643 the AZ, other factors aside from the ballast effect of biogenic silica and diatom abundance
644 must play a major role in the efficiency by which POC is transported to the deep sea in
645 AZ ecosystems. Many factors including ecosystem structure and composition, as well as
646 physical (e.g. insolation, water column stability, annual sea ice cover, etc.) and chemical
647 parameters (e.g. availability of macro- and micronutrients, etc.) are known to influence
648 the magnitude and composition of the particles sinking to the deep ocean (e.g. Wassmann,
649 1998; Honjo et al., 2008; Ebersbach et al., 2011; Laurenceau-Cornec et al., 2015). Among
650 them, the makeup of the diatom community (i.e., diatom floristics) has been suggested to
651 be one of the main factors responsible for setting the different degree of coupling between
652 carbon and silicon in the particles sinking in the AZ (Assmy et al., 2013; Boyd, 2013).

653 The poor correlation between diatom valve and POC fluxes recorded in the traps is known
654 to be largely determined by the pronounced differences in organic carbon content between
655 diatom species and differences in the full cell: empty cell ratios of the diatom cells
656 reaching the trap depths. Indeed, previous sediment trap studies in other Southern Ocean
657 settings have demonstrated that the ratio of full to empty diatoms cells largely vary
658 between species, representing a first-order control in the BSi : POC export stoichiometry
659 of a given ecosystem. For example, Salter et al. (2012), Rembauville et al. (2014) and
660 Rembauville et al. (2016) highlighted the important role of resting stages from *Eucampia*
661 *antarctica* var. *antarctica*, *Chaetoceros* and *Thalassiosira antarctica* in the export of
662 organic carbon from the surface to the deep ocean. Moreover, organisms of higher trophic
663 levels are known to exert a significant influence in the efficiency of the transfer of organic
664 matter from the euphotic zone into the deep layers of the water column either through the
665 formation of faecal pellets (e.g. Lampitt et al., 2009; Ebersbach et al., 2011; Rembauville
666 et al., 2014; Manno et al., 2015; Belcher et al., 2017) or by vertical migrations (e.g.
667 Jackson and Burd, 2001; Davison et al., 2013). Thus, different zooplankton composition
668 at each of the studied stations may also account for part of the POC variability export
669 across regions of the AZ.

670 Lastly, the importance of heterotrophic bacteria in determining the degradation depths of
671 POC is becoming increasingly evident. A recent study by Edwards et al. (2015)
672 demonstrated how carbon-rich diatom aggregates sinking out the surface layer can be hot
673 spots for production of polyunsaturated aldehydes (PUAs), highly bioactive molecules
674 that can stimulate bacteria respiration at adequate concentrations. The stimulation of the
675 metabolism of the heterotrophic bacteria by the PUAs results in the faster
676 remineralization of phytoplankton-derived sinking organic matter leading to the shoaling
677 of remineralization depths of carbon and other nutrients. Taking into that the diatom
678 species dominating each of the sectors of the AZ exhibit significantly different life
679 strategies and metabolisms, variations in PUAs production between different polar
680 diatom species is likely, and therefore it could potentially play an important role in the
681 export efficiency in diatom dominated ecosystems such as the ones compared here.

682

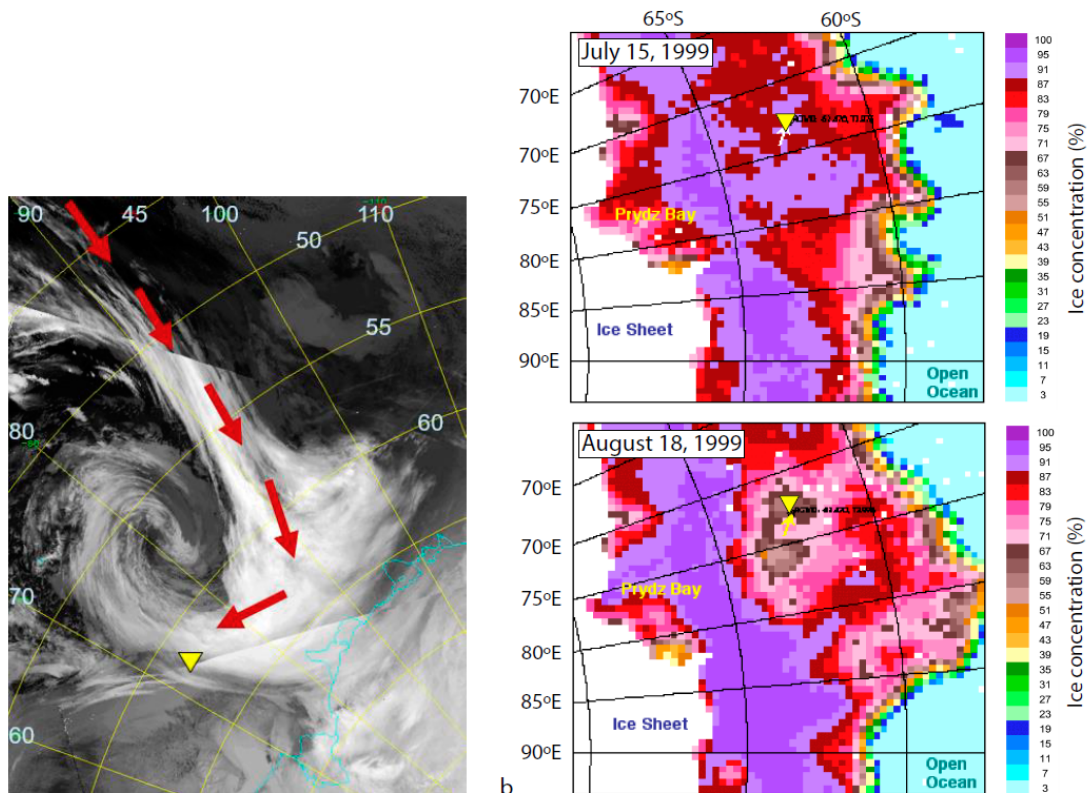
683 **Conclusions**

684

685 In this study we analyzed diatom floristics, POC, Bio-SiO₂ fluxes, and environmental
686 parameters across key regions of the AZ. In the first part of the study we documented the
687 seasonal variability of diatom species and its relationship with biogeochemical fluxes
688 intercepted by a sediment trap over a year in the offshore waters of Prydz Bay. A short
689 and pronounced diatom valve export period during the summer season co-occurred with
690 the highest annual POC fluxes at the PZB-1 station, suggesting that diatoms represent the
691 main organic carbon vectors to ocean interior in this region. Two peaks of enhanced
692 export were recorded during winter and autumn, a period characterized by extensive sea
693 ice cover. Taxonomic analysis of the diatom assemblages of these two export events
694 suggest different origins of the materials collected by the trap: the August peak seems to
695 be caused by lateral advection event while the November maximum seems to be the result
696 of the initiation of the sea-ice retreat and consequent onset of the spring phytoplankton
697 bloom.

698 In the second part of the study, five sediment trap experiments reporting diatom
699 assemblage composition and biogeochemical fluxes conducted in different regions of the
700 AZ were compared. The relative abundance of *F. kerguelensis* at all the study stations
701 seems to be largely influenced by the presence of sea ice, with maximum abundance of
702 this species in the only station not affected by sea ice and minima in the southernmost site
703 of the Ross Sea sector, characterized by the highest annual sea ice cover of all stations.
704 Annual fluxes of the small *Fragilariopsis* species group exhibited a strong and significant
705 correlation with total chlorophyll-*a* concentration across stations. This observation
706 highlights the potential of these species as a proxy of high algal biomass accumulation in
707 Southern Ocean paleorecords. Despite the fact that diatoms are the main Bio-SiO₂
708 producer at all stations, the annual biogenic silica and diatom valve export fluxes did not
709 covary across sites. This is most likely due to the pronounced differences in biogenic
710 silica content per cell of the dominant diatom species at each study station. Interestingly,
711 annual Bio-SiO₂ and *F. kerguelensis* fluxes alone exhibited a much stronger correlation
712 which underscores, once again, the major role of this species for selective silicon export
713 into the ocean interior of the AZ.

714 **Supplementary material**



715

716 **Supplementary Figure 1:** a. NOAA Advanced Very High Resolution Radiometer
 717 Channel 4 mosaic (thermal IR) between 15th -19th of August 1999, 4 km resolution for
 718 the study region. b. Daily ice concentration maps for the study region during July 15,
 719 1999 (typical winter conditions) and August 18, 1999 (showing small open water areas
 720 in the sea-ice around the sediment trap location). Yellow triangle shows the position of
 721 the sediment trap PZB-1.

722 **Supplement II.** Diatom species flux data of PZB-1 (excel file).

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732

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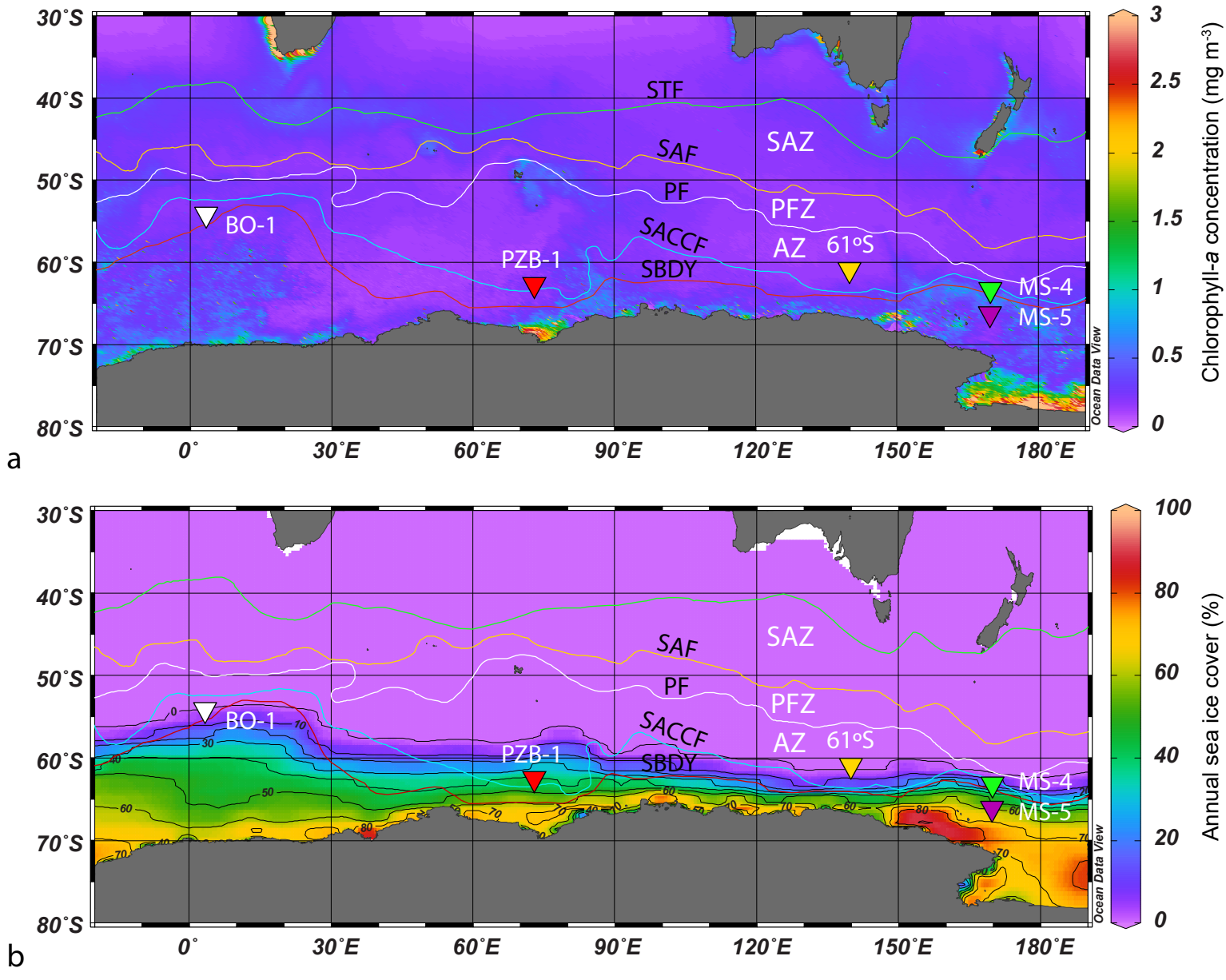


Figure 1

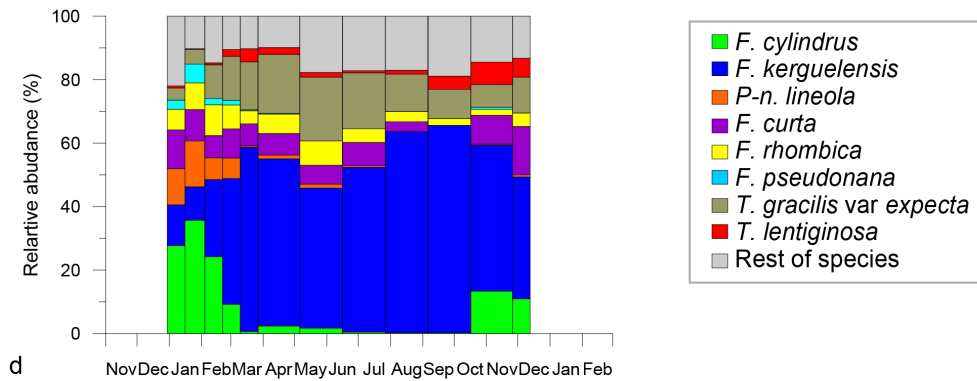
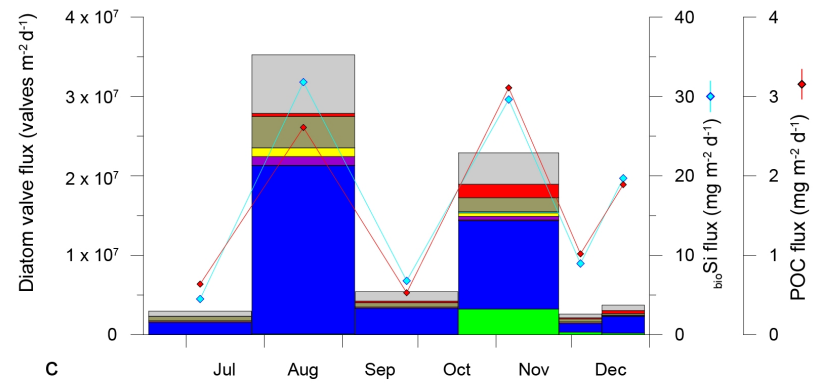
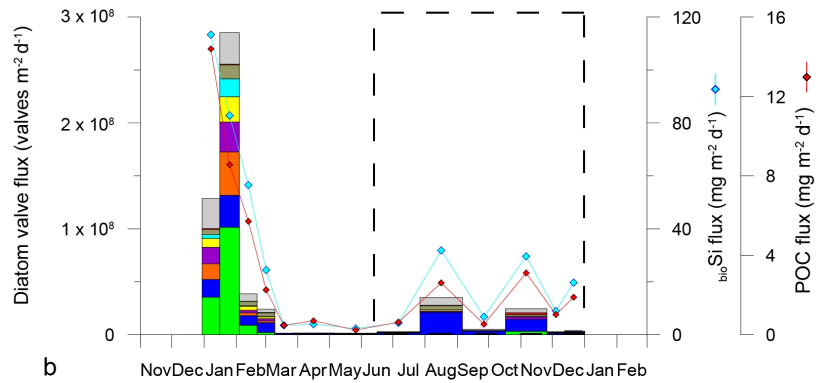
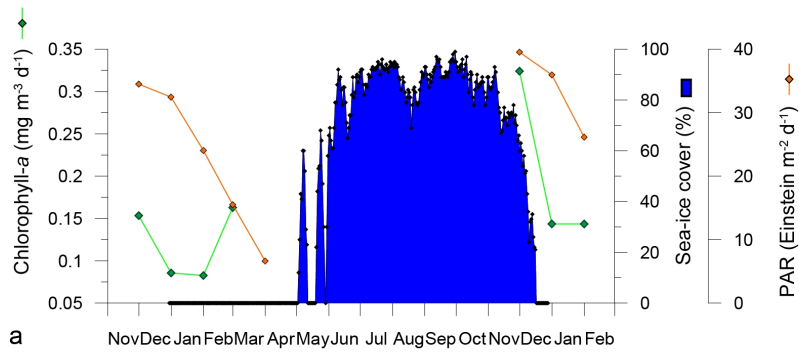


Figure 2

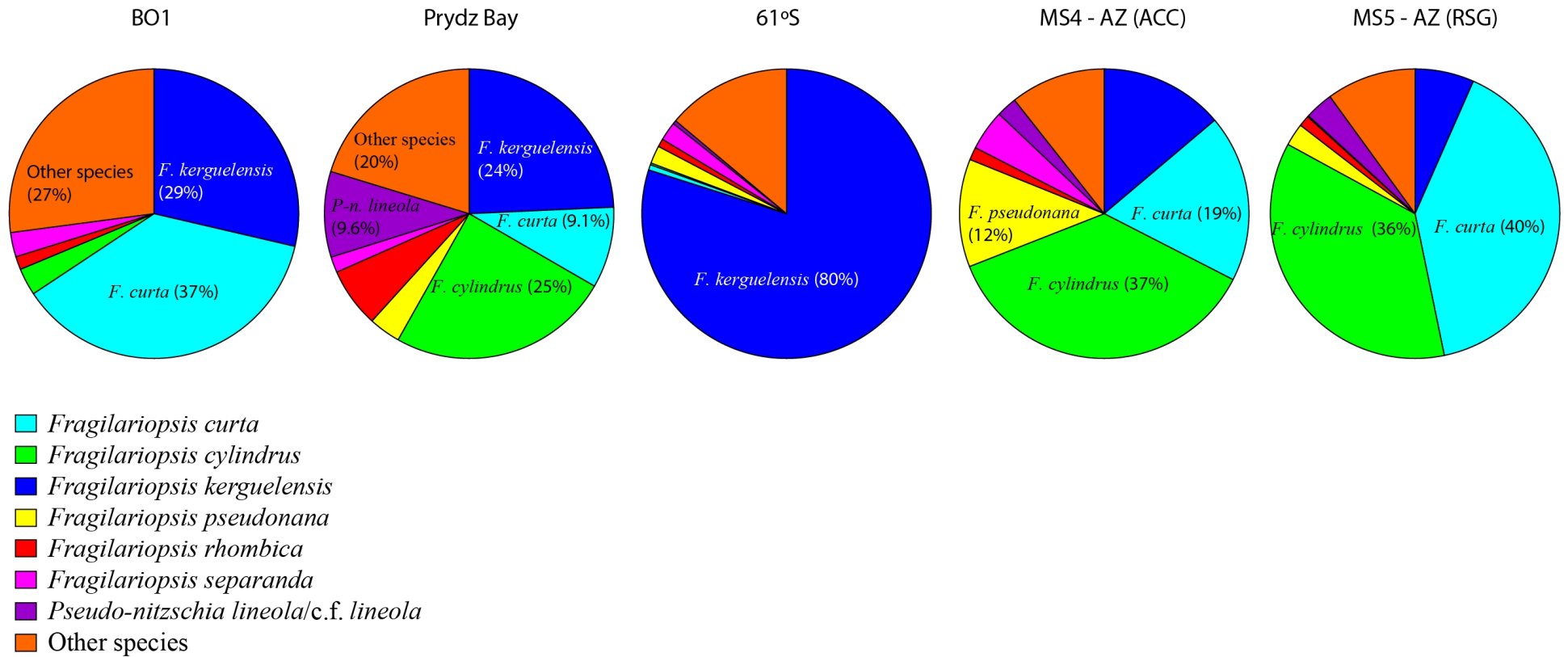
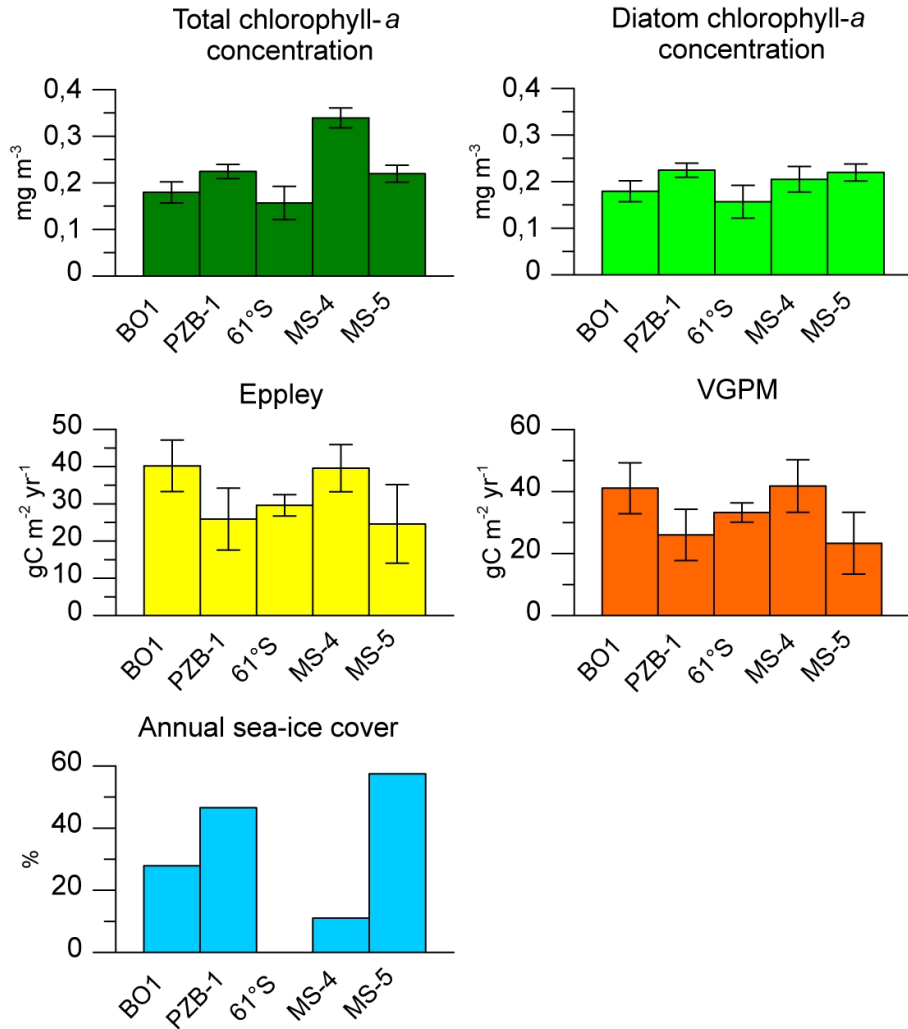
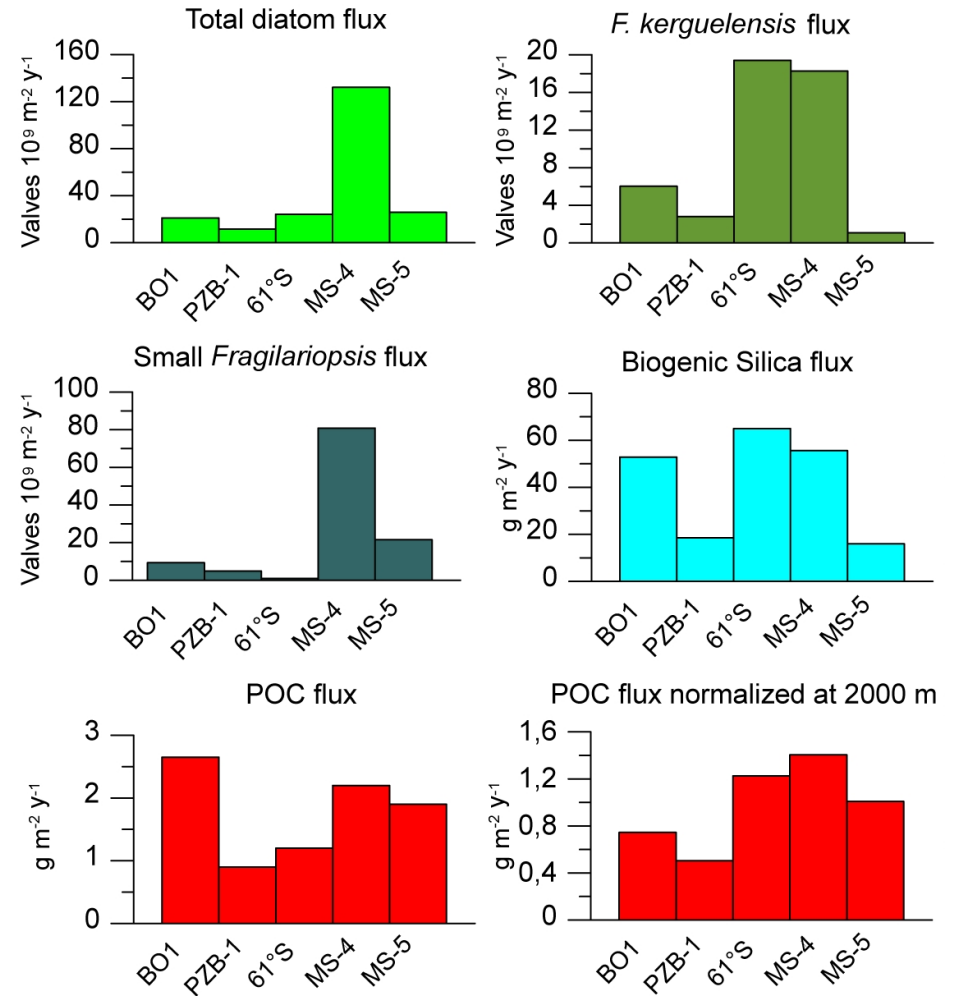


Figure 3.

a)



b)



Station	Latitude	Longitude	Water column depth (m)	Trap depth (m)	Sampling interval start	Sampling interval end
BO-1	54° 20'S	3° 23'E	2734	450	28/12/1990	01/04/1992
PZB-1	62°29'S	72°59'E	4000	1400	30/12/1998	13/12/1999
61°S	60° 44'S	139° 54'E	4393	2000	30/11/2001	29/09/2002
MS-4	63° 09'S	169° 54'W	2885	1031	28/11/1996	24/12/1997
MS-5	66° 10'S	169° 40'W	3015	937	28/11/1996	24/12/1997

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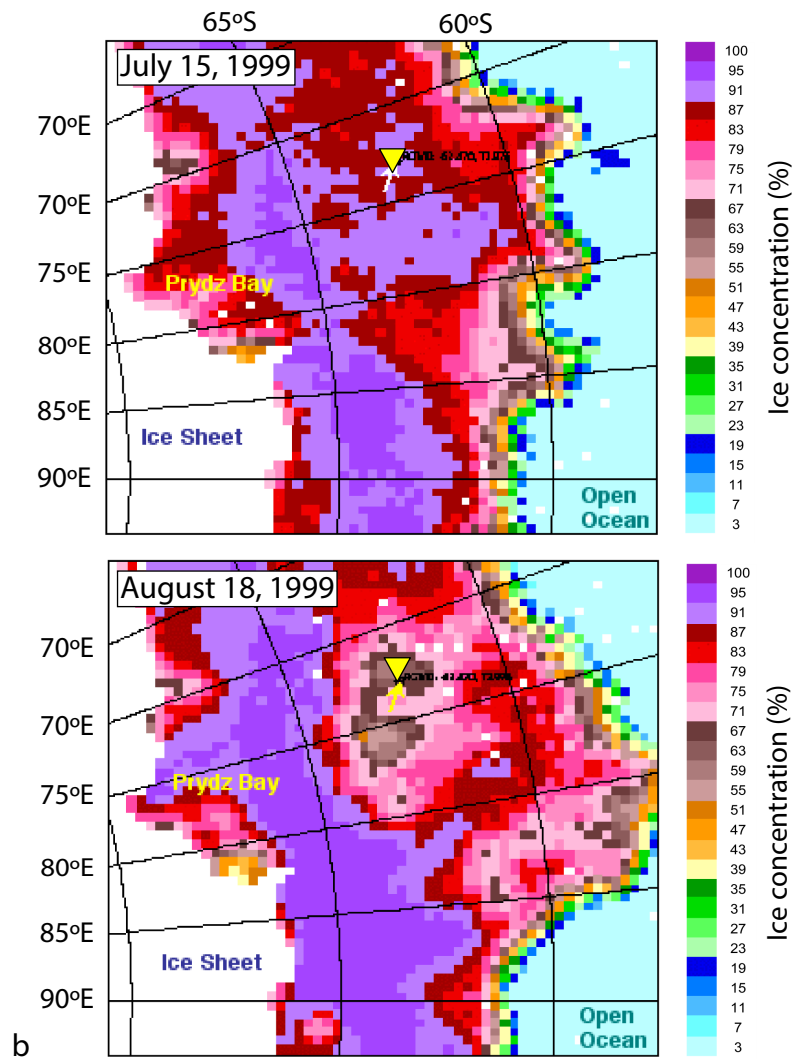
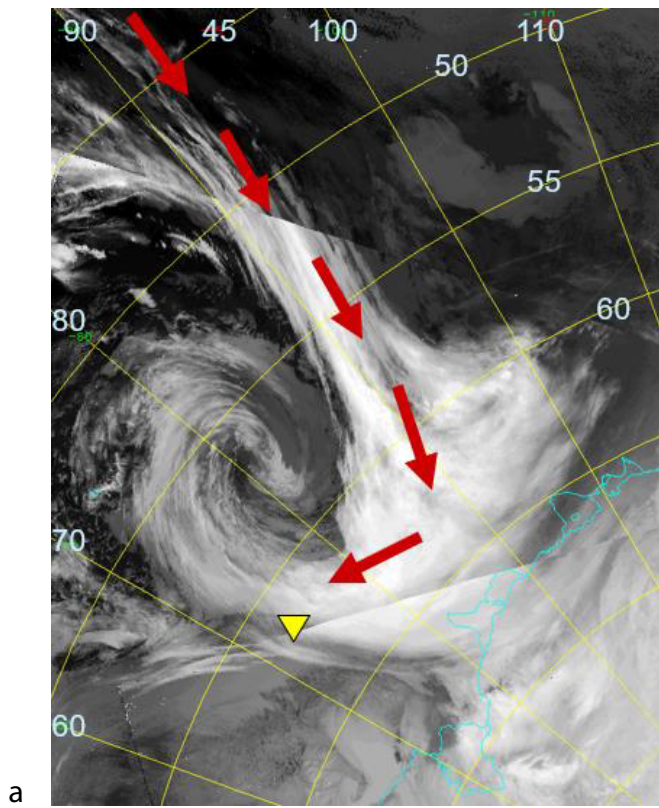
Honjo et al. (2000) and Grigorov et al. (2014)

Cup number	Mid-point date	Collection days (days)	Total diatom flux (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Fragilariopsis curta</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Fragilariopsis cylindrus</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Fragilariopsis kerguelensis</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Fragilariopsis pseudonana</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Fragilariopsis rhombica</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Pseudo-nitzschia lineola</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Thalassiosira gracilis</i> var. <i>expecta</i> (10 ⁶ x valves m ⁻² d ⁻¹)	<i>Thalassiosira lentiginosa</i> (10 ⁶ x valves m ⁻² d ⁻¹)
1	07/01/1999	17.0	128.6	15.8	35.5	16.6	3.7	8.3	14.6	4.9	0.9
2	25/01/1999	19.0	284.6	28.2	101.6	30.1	16.9	23.8	41.1	12.5	0.6
3	12/02/1999	17.0	37.2	2.6	9.0	9.0	0.7	3.6	2.5	2.5	0.2
4	01/03/1999	17.0	23.4	2.2	2.2	9.3	0.3	1.8	1.5	2.5	0.5
5	18/03/1999	17.0	1.5	0.1	0.0	0.9	0.0	0.1	0.0	0.1	0.1
6	16/04/1999	40.0	1.5	0.1	0.0	0.8	0.0	0.1	0.0	0.2	0.0
7	26/05/1999	41.0	1.4	0.1	0.0	0.6	0.0	0.1	0.0	0.2	0.0
8	06/07/1999	41.0	2.9	0.2	0.0	1.5	0.0	0.1	0.0	0.4	0.0
9	16/08/1999	41.0	33.5	1.0	0.1	21.2	0.0	1.1	0.0	2.3	0.4
10	26/09/1999	41.0	5.1	0.0	0.0	3.3	0.0	0.1	0.0	0.3	0.2
11	06/11/1999	40.0	24.2	2.2	3.2	11.1	0.2	0.4	0.1	1.3	1.7
12	04/12/1999	17.0	2.8	0.4	0.3	1.1	0.0	0.1	0.0	0.2	0.2
13	21/12/1999	17.0	3.9	0.3	0.2	2.1	0.0	0.1	0.0	0.1	0.4

Species	BO1	PZB-1	61°S	MS-4	MS-5	Species	BO1	PZB-1	61°S	MS-4	MS-5
<i>Actinocyclus actinochilus</i> (Ehrenberg) Simonsen		0.12	*	*	*	<i>P. truncata</i> (G.Karsten) Nöthig et Ligowski		*	o	o	o
<i>Actinocyclus curvatus</i> Janisch		*	o	*	o	<i>Proboscia</i> spp.		*	o	o	o
<i>Actinocyclus exiguus</i> Fryxell et Semina		o	o	o	o	<i>Psammmodicum panduriforme</i> (Gregory) Mann		o	o	o	o
<i>Actinocyclus octonarius</i> Ehrenberg		o	o	*	o	<i>Pseudo-nitzschia</i> cf. <i>lineola</i>		o	0.4	o	o
<i>Actinocyclus</i> spp.		o	*	*	*	<i>P-n. lineola</i> (Cleve) Hasle		9.57	o	2.30	3.18
<i>Alveus marinus</i> (Grunow) Kaczmarek et Fryxell		o	o	o	o	<i>P-n. turgiduloides</i> (Hasle) Hasle		1.58	o	o	o
<i>Asteromphalus hookeri</i> Ehrenberg		0.21	0.2	0.16	1.11	<i>P-n. prolongatoides</i> (Hasle) Hasle		0.22	o	o	o
<i>A. hyalinus</i> Karsten		*	0.2	*	o	<i>P-n. heimii</i> Manguin		o	*	o	o
<i>A. parvulus</i> Karsten		0.34	0.2	*	*	<i>Pseudo-nitzschia</i> spp.		*	0.1	o	0.13
<i>Asteromphalus</i> spp.		o	*	o	o	<i>Rhizosolenia antennata</i> (Ehrenberg) Brown f. <i>antennata</i>		*	o	o	o
<i>Azpeitia tabularis</i> (Grunow) Fryxell et Sims		0.33	*	0.7	0.15	<i>R. antennata</i> (Ehrenberg) Brown f. <i>semispina</i> Sundström		0.70	o	o	o
<i>Banquisia belgicae</i> (Van Heurck) Paddock		0.38	o	o	o	<i>R. bergonii</i> Peragallo		o	o	o	o
<i>Chaetoceros aequatorialis</i> var. <i>antarcticus</i> Manguin		o	*	o	o	<i>Rhizosolenia</i> cf. <i>costata</i>		o	o	o	o
<i>C. atlanticus</i> Cleve		0.95	0.2	0.19	*	<i>Rhizosolenia</i> cf. <i>chunii</i>		0.23	o	o	o
<i>C. criophilus</i> Castracane		o	o	*	*	<i>R. curvata</i> Zacharias		o	o	o	o
<i>C. diadema</i> Ehrenberg		o	o	o	*	<i>R. polydactyla</i> Castracane f. <i>polydactyla</i>		o	o	o	o
<i>C. dichæta</i> Ehrenberg		*	0.1	o	o	<i>R. polydactyla</i> Castracane f. <i>squamosa</i>		*	o	o	o
<i>C. peruvianus</i> Brightwell		*	o	o	o	<i>R. simplex</i> Karsten		o	o	*	o
<i>Chaetoceros</i> subgenus <i>Hyalochaete</i> spp.		0.49	0.2	*	0.13	<i>R. sima</i> Castracane		o	o	o	*
<i>Chaetoceros</i> subgenus <i>Phaeoceros</i> spp.		0.2				<i>R. styliformis</i> Brightwell		o	o	*	o
<i>Chaetoceros</i> resting spores		2.59	0.1	o	o	<i>Rhizosolenia</i> sp. f. 1A <i>sensu</i> Armand et Zielinski		*	*	o	o
<i>Cocconeis</i> spp.		o	o	o	o	<i>Rhizosolenia</i> spp.		o	0.1	o	o
<i>Corethron</i> sp.		0.25	*	*	*	<i>Roperia tessellata</i> (Roper) Grunow		o	o	o	o
<i>Coscinodiscus asteromphalus</i> Ehrenberg		*	o	o	o	<i>Stellarima stellaris</i> (Roper) Hasle et Sims		o	o	o	o
<i>Cyclotella</i> spp.		o	o	o	o	<i>Synedropsis</i> sp.		*	o	o	o
<i>Dactylosolen antarcticus</i> Castracane		o	o	o	o	<i>Thalassionema nitzschoides</i> var. <i>capitulata</i> (Castracane) Moreno-Ruiz		o	0.1	o	o
<i>Diploneis bombus</i> (Ehrenberg) Ehrenberg		o	o	o	o	<i>T. nitzschoides</i> var. <i>lanceolata</i> (Grunow) Pergallo et Pergallo		o	0.1	*	o
<i>Eucampia antarctica</i> (Castracane) Mangin (summer form)		*	o	*	o	<i>T. nitzschoides</i> var. <i>parvum</i> Moreno-Ruiz		o	o	o	o
<i>E. antarctica</i> (Castracane) Mangin (winter form)		o	0.1	o	o	<i>T. nitzschoides</i> var. 1 <i>sensu</i> Zielinski et Gersonde		3.45	o	o	* 0.16
<i>Fragilariopsis curta</i> (Van Heurck) Hustedt		37.05	9.05	0.6	18.70	35.21	<i>Thalassiosira antarctica</i> Comber		*	o	* 0.09
<i>F. cylindrus</i> (Grunow) Krieger		3.04	24.83	0.2	36.52	47.04	<i>T. decipiens</i> (Grunow ex Van Heurck) Jørgensen		o	o	o 0.17
<i>F. doliolus</i> (Wallich) Medlin et Sims		o	o	o	o	<i>T. eccentrica</i> (Ehrenberg) Cleve		o	0.2	o	o
<i>F. kerguelensis</i> (O'Meara) Hustedt		28.63	24.29	79.9	13.83	4.13	<i>T. ferelineata</i> Hasle et Fryxell		o	o	o 0
<i>F. obliquecostata</i> (van Heurck) Heiden		*	*	*	0.16	<i>T. gracilis</i> var. <i>expecta</i> (Van Landingham) Fryxell et Hasle		5.20	0.4	7.09	o
<i>F. pseudonana</i> (Hasle) Hasle		3.55	2	12.05	2.96	<i>T. gracilis</i> var. <i>gracilis</i> (Karsten) Hustedt		1.40	3.6	o	o
<i>F. rhombica</i> (O'Meara) Hustedt		1.47	6.67	0.9	1.43	0.79	<i>T. gracilis</i> group		o	4.1	o 1.67
<i>F. ritscherii</i> Hustedt		0.94	0.1	0.73	1.33	<i>T. gravida</i> Cleve		6.97	0.43	*	0.14 0.12
<i>F. separanda</i> Hustedt		2.72	1.76	2.1	4.53	0.18	<i>T. lentiginosa</i> (Janisch) Fryxell		2.79	1.26	5 0.81 0.27
<i>Fragilariopsis sublinearis</i> (Van Heurck) Heiden et Kolbe		0.57	o	o	o	<i>T. leptopus</i> (Grunow ex Van Heurck) Hasle et Fryxell		o	*	o	o
<i>F. cf. sublineata</i> (Van Heurck) Heiden		o	*	o	o	<i>T. lineata</i> Jousé		o	o	o	o
<i>Fragilariopsis vanheurckii</i> (Peragallo) Hustedt		0.26	o	o	o	<i>T. maculata</i> Fryxell et Johans		o	*	o	o
<i>Fragilariopsis</i> spp.		o	o	o	o	<i>T. oestrupii</i> (Ostenfeld) Hasle var. <i>oestrupii</i> Fryxell et Hasle		*	*	0.13	0.02
<i>Guinardia</i> spp.		o	o	o	*	<i>T. oestrupii</i> (Ostenfeld) Hasle var. <i>venrickae</i> Fryxell et Hasle		o	o	o	o
<i>Gyrosigma</i> spp.		o	o	o	o	<i>T. oliveriana</i> (O'Meara) Makarova et Nikolaev		*	0.7	o	0.07
<i>Haslea trompii</i> (Cleve) Simonsen		0.16	*	o	o	<i>T. symmetrica</i> Fryxell et Hasle		o	o	o	o
<i>Hemidiscus cuneiformis</i> Wallich		o	o	o	o	<i>T. trifulta</i> Fryxell		0.11	o	o	o
<i>Manguinea</i> spp.		*	o	o	o	<i>T. tumida</i> (Janisch) Hasle		*	0.1	o	o
<i>Membraneis</i> spp.		0.33	o	o	o	<i>Thalassiosira</i> sp. 1		o	*	o	o
<i>Navicula directa</i> (Smith) Ralfs in Pritchard		0.23	0.3	o	o	<i>Thalassiosira</i> sp. 2		o	o	o	o
<i>Navicula</i> spp.		*	o	0.55	0.55	<i>Thalassiosira</i> sp. 3		o	o	o	o
<i>Nitzschia bicapitata</i> Cleve		o	o	o	o	<i>Thalassiosira</i> eccentric group		o	o	o	o
<i>N. braarudii</i> (Hasle)		o	o	o	o	<i>Thalassiosira</i> linear group		o	0.1	o	o
<i>N. kolaceckii</i> Grunow		o	o	o	o	<i>T. trifulta</i> group		o	o	o	o
<i>N. sicula</i> (Castracane) Hustedt var. <i>bicuneata</i> Grunow		o	0.1	o	o	<i>Thalassiosira</i> spp. < 20 µm		o	0.4	*	o
<i>N. sicula</i> (Castracane) Hustedt var. <i>rostrata</i> Hustedt		0.10	o	o	o	<i>Thalassiosira</i> spp. > 20 µm		o	*	o	o
<i>Nitzschia</i> spp.		o	o	o	o	<i>Thalassiothrix antarctica</i> Schimper ex Karsten		o	0.2	0.20	0.23
<i>Paralia</i> spp.		0.04	o	o	o	<i>Thalassiothrix longissima</i> + <i>antarctica</i>		0.46	o	o	o
<i>Pleurosigma</i> spp.		o	*	0.01	o	<i>Trachyneis aspera</i> (Ehrenberg) Cleve		o	o	o	o
<i>Pleurosigma directum</i>		0.03	o	o	o	<i>Trichotoxon reinboldii</i> (Van Heurck) Reid et Round		*	o	*	0.10
<i>Porosira pseudodenticulata</i> (Hustedt) Jousé		o	*	o	o	<i>Tropidoneis</i> group		o	*	o	o
<i>Proboscia alata</i> (Brightwell) Sundström		*	o	o	o	Other centrics		*	*	o	o
<i>P. inermis</i> (Castracane) Jordan et Ligowski		*	o	o	o	Other pennates		*	0.1	o	o

	Total Chl- <i>a</i> concentration	Sea ice cover	Total diatom flux	<i>F. kerguelensis</i> flux	Small <i>Fragilariopsis</i> group flux	Biogenic Silica flux	POC flux	POC flux normalized at 2000m
Total Chl- <i>a</i> concentration	1.000 p= ---							
Sea ice cover	-0.029 p=.964	1.000 p= ---						
Total diatom flux	0.891 p=.043	-0.433 p=.467	1.000 p= ---					
<i>F. kerguelensis</i> flux	0.218 p=.724	-0.960 p=.010	0.588 p=.297	1.000 p= ---				
Small <i>Fragilariopsis</i> group flux	0.944 p=.016	-0.235 p=.703	0.977 p=.004	0.409 p=.494	1.000 p= ---			
Biogenic Silica flux	-0.056 p=.929	-0.958 p=.010	0.379 p=.529	0.869 p=.056	0.199 p=.749	1.000 p= ---		
POC flux	0.236 p=.702	-0.055 p=.930	0.381 p=.527	-0.008 p=.990	0.433 p=.466	0.297 p=.627	1.000 p= ---	
POC flux normalized at 2000m	0.419 p=.482	-0.637 p=.247	0.730 p=.162	0.787 p=.114	0.650 p=.236	0.588 p=.297	0.244 p=.692	1.000 p= ---

Table 4.



Supplementary Figure 1