1 Article Type: Original Article

```
2 Intraspecific genetic variation matters when predicting seagrass distribution under climate change
```

3 Running Title: Intraspecific prediction of seagrass distribution

- 4 Zi-Min Hu¹, Quan-Sheng Zhang¹, Jie Zhang², Jamie M. Kass³, Stefano Mammola^{4,5}, Pablo Fresia⁶,
- 5 Stefano G. A. Draisma⁷, Jorge Assis⁸, Alexander Jueterbock⁹, Masashi Yokota¹⁰, and Zhixin Zhang¹⁰
 6
- 7 ¹Ocean School, YanTai University, Yantai, 264005, China
- 8 ²Key Laboratory of Experimental Marine Biology, Center for Ocean Mega-Science, Institute of
- 9 Oceanology, Chinese Academy of Sciences, Qingdao, 266071, China
- 10 ³Biodiversity and Biocomplexity Unit, Okinawa Institute of Science and Technology Graduate
- 11 University, Okinawa 904-0495, Japan
- 12 ⁴Laboratory for Integrative Biodiversity Research (LIBRe), Finnish Museum of Natural History
- 13 (LUOMUS), University of Helsinki, Pohjoinen Rautatiekatu 13, 00100 Helsinki, Finland
- 14 ⁵Molecular Ecology Group (MEG), Water Research Institute (IRSA), National Research Council of
- 15 Italy (CNR), Verbania Pallanza, 28922, Italy
- 16 ⁶Unidad de Bioinformatica, Institut Pasteur de Montevideo, Mataojo, 2020 Montevideo, Uruguay
- 17 ⁷Excellence Center for Biodiversity of Peninsular Thailand, Faculty of Science, Prince of Songkla
- 18 University, Hat Yai, Songkhla 90110, Thailand
- 19 ⁸CCMAR, University of Algarve, Campus de Gambelas, Faro, 8005–139, Portugal
- 20 9Algal and Microbial Biotechnology Division, Faculty of Biosciences and Aquaculture, Nord
- 21 University, Bodø, 8026, Norway
- 22 ¹⁰Graduate School of Marine Science and Technology, Tokyo University of Marine Science and
- 23 Technology, Konan, Minato, Tokyo 108-8477, Japan
- 24
- 25 Correspondence: Zhixin Zhang, Graduate School of Marine Science and Technology, Tokyo
- 26 University of Marine Science and Technology, Konan, Minato, Tokyo 108-8477, Japan. E-mail:
- 27 zhangzhixin1102@hotmail.com

29 Abstract

30 Seagrasses play a vital role in structuring coastal marine ecosystems, but their distributional range 31 and genetic diversity have declined rapidly over the past decades. In order to improve 32 conservation of seagrass species, it is important to predict how climate change may impact their 33 ranges. Such predictions are typically made with correlative species distribution models (SDMs), 34 which can estimate a species' potential distribution under present and future climatic scenarios given species' presence data and climatic predictor variables. However, these models are typically 35 36 constructed with species-level data, and thus ignore intraspecific genetic variability, which can 37 give rise to populations with adaptations to heterogeneous climatic conditions. Here, we explore 38 the link between intraspecific adaptation and niche differentiation in *Thalassia hemprichii*, a 39 seagrass broadly distributed in the tropical Indo-Pacific Ocean and a crucial provider of habitat for 40 numerous marine species. By retrieving and re-analyzing microsatellite data published previously, we delimited two distinct phylogeographical lineages within the nominal species and found an 41 42 intermediate level of differentiation in their multidimensional environmental niches, suggesting 43 the possibility for local adaptation. We then compared projections of the species' habitat 44 suitability under climate change scenarios using species-level and lineage-level SDMs. In the 45 Central Tropical Indo-Pacific region, models for both levels predicted considerable range contraction in the future, but the lineage-level models predicted more severe habitat loss. 46 47 Importantly, the two modelling approaches predicted opposite patterns of habitat change in the 48 Western Tropical Indo-Pacific region. Our results highlight the necessity of conserving distinct 49 populations and genetic pools to avoid regional extinction due to climate change and have 50 important implications for guiding future management of seagrasses.

51

Keywords: climate change scenario, genetic lineage, niche conservation, range shift, species
distribution model, *Thalassia hemprichii*

54 Introduction

55 Marine ecosystems worldwide are experiencing rapid shifts in environmental conditions due to 56 climate change, the most evident of which is a steady increase in sea surface temperature (SST) 57 (Cheung et al. 2013). These changes can affect marine organisms in different ways, such as by 58 altering the structure of trophic webs (e.g., Hyndes et al. 2016), biasing sex ratios in species with 59 temperature-dependent sex determination (e.g., Miyoshi et al. 2020), and redefining the geographical ranges of species (e.g., Pinsky et al. 2020). In order to guide natural resource 60 61 management under this changing marine landscape, it is crucial to make future predictions of 62 suitable habitat for target species as accurately as possible.

63 Species distribution models (SDMs), which estimate relationships between species' presence data and environmental predictors, have been used extensively to predict potential changes in 64 65 species' distributions under climate change scenarios (Guisan et al. 2017). The majority of SDMs are constructed at the species-level or even higher taxonomic levels, and this is particularly true 66 for applications to marine species (Robinson *et al.* 2011; Robinson *et al.* 2017; Chefaoui *et al.* 2018; 67 Jayathilake & Costello 2018; Melo-Merino et al. 2020). One fundamental and critical assumption 68 underlying species-level SDMs is niche conservatism, which assumes that all populations of a 69 70 species have analogous environmental requirements and respond in a similar way to a changing 71 environment (Guisan et al. 2017; Smith et al. 2019). But this assumption ignores intraspecific 72 variation, in particular local adaptation and phenotypic plasticity (Pazzaglia et al. 2021), which are frequently observed especially in broadly distributed taxa (e.g., Marín-Guirao et al. 2016; Duarte 73 et al. 2018; King et al. 2018; Benito Garzón et al. 2019; Peterson et al. 2019; Zhang et al. 2020b). 74 75 SDMs constructed with data for lineages below the species level can account for possible local adaptations and therefore can provide more reliable niche estimations and habitat suitability 76 77 projections for species with high intraspecific variation. For instance, a species-level SDM for the 78 threatened Japanese crayfish Cambaroides japonicus (De Haan 1841) predicted that this species

79 might lose a large proportion of its suitable habitat in the future, whereas lineage-level SDMs for 80 the same species predicted a weaker impact of climate change overall (Zhang et al. 2021). The importance of taxonomic units (i.e., above and below the species level) in distribution modelling 81 82 has recently been recognized (Benito Garzón et al. 2019; Peterson et al. 2019; Smith et al. 2019; 83 Collart *et al.* 2021), which has resulted in more SDM applications for terrestrial and freshwater 84 species that consider intra-specific variation (e.g., Ikeda et al. 2017; Razgour et al. 2019; Zhang et 85 al. 2021). Conversely, relatively few SDM studies have investigated this issue in the marine realm 86 (but see Assis et al. 2018a; Cacciapaglia & van Woesik 2018; Lowen et al. 2019). 87 Seagrasses are one of the most critical habitat engineers of tropical coastal marine 88 environments. They not only harbor rich marine biodiversity in seagrass meadows, but also 89 provide a number of ecosystem services, such as primary productivity, habitat restoration, 90 resources for marine life, and human recreation (Unsworth et al. 2018). Maintaining these services is key to achieving conservation and economic goals under global change. Yet, seagrass ecosystems 91 are declining worldwide at an annual rate of 7% due to multiple natural and human-mediated 92 93 disturbances (Orth et al. 2006; Waycott et al. 2009). It is noteworthy that climate change has received considerable attention as a major factor for the increasing loss of seagrass meadows (Jordà 94 95 et al. 2012; Thomson et al. 2015; Repolho et al. 2017; Duarte et al. 2018; Smale et al. 2019). This is 96 particularly true for the tropical Indo-Pacific bioregion, which supports the most seagrass diversity 97 and a high diversity of associated flora and fauna (Short et al. 2007) but has suffered from striking 98 degradation of seagrass coverage (Coles et al. 2011; Rasheed & Unsworth 2011; Grech et al. 2012; 99 Chefaoui et al. 2018; Olsen et al. 2018; Brodie et al. 2020). Given the global ecological roles of seagrasses, it is imperative to make accurate forecasts of their distribution patterns in the face of 100 101 climate change, but seagrasses are "among the least-studied groups" (Melo-Merino et al. 2020) 102 with respect to range shift projections. The majority (if not all) of SDM studies on seagrasses have 103 been at the species level and therefore did not incorporate potential intraspecific variation.

The seagrass Thalassia hemprichii (Ehrenberg) Ascherson (Hydrocharitaceae) is a perennial 104 climax species that is widely distributed in the tropical Indo-Pacific bioregion (Green & Short 105 106 2003), extending from Australia, the peripheral limit of its eastern range (Hernawan et al. 2017), 107 to East Africa in the West Indian Ocean (Jahnke et al. 2019a). It reproduces sexually via seeds and 108 asexually via vegetative growth of rhizomes. Uprooted adult plants can potentially float for 109 months and hence colonize distant areas (Wu et al. 2016). In addition, this seagrass forms buoyant seeds that remain afloat for long enough to disperse a few hundreds of kilometers (Lacap et al. 110 111 2002). A recent survey revealed that seedlings can also disperse for over a month due to the accumulation of oxygen in the body tissue (Wu et al. 2016). Thus, T. hemprichii has excellent 112 113 long-distance dispersal potential that may play a significant role in shaping population genetic 114 structure (Lowe & Allendorf 2010). This species may be particularly vulnerable to climate change 115 because it exhibits spatial separation of the sexes (dioecious), reinforced by physiological and morphological differentiation of each sex to variable microhabitats (Hultine et al. 2016). Recent 116 genetic studies of T. hemprichii detected genetic lineage divisions in the East and West Indo-117 Pacific Ocean (Hernawan et al. 2017; Jahnke et al. 2019a), but we still do not have a clear 118 understanding of the distribution of lineages across the entire tropical Indo-Pacific region, or 119 120 whether these diverged lineages are expected to respond differentially to climate change. In the present study, we used *T. hemprichii* as a model to: (i) examine divergence of genetic 121 lineages in the tropical Indo-Pacific Ocean; (ii) test if phylogeographical lineages exist, and if so, 122 quantify niche differentiation between distinct lineages; (iii) predict climate change impacts on 123 the species' range with species-level and lineage-level SDMs. By incorporating potential 124 intraspecific variation, our SDMs can provide more realistic predictions on how climate change 125 126 will shift future distributions of a habitat-forming seagrass, thus generating valuable knowledge 127 for guiding the long-term management of this species in the tropical Indo-Pacific coast.

129 Materials and methods

130 Datasets and intraspecific genetic clustering

We retrieved and compiled molecular datasets of two recently published regional studies of T. 131 hemprichii, i.e., 17 populations in the East Indo-Pacific (Hernawan et al. 2017) and 11 populations 132 in the Western Indian Ocean (Jahnke et al. 2019a). We used twelve microsatellites (i.e., Thh3, 133 Thh15, Thh34, Thh41, TH07, TH34, TH37, TH43, TH52, TH66, TH73) for population structuring 134 and lineage sorting of 1021 individuals from 28 populations across the tropical Indo-Pacific (Fig. 135 136 1a). We then estimated pairwise genetic differences among populations using the Cavalli-Sforza and Edwards chord distance and represented them in a network using the R package IGRAPH 137 138 (Csardi & Nepusz 2006) with the addition of a custom script by Johansson *et al.* (2015). To visually inspect the relationships within and between the main genetic clusters inferred by STRUCTURE 139 140 (Pritchard et al. 2000), we pruned the full network by sequentially removing edges (i.e., network pairwise links among sampling sites) of decreasing genetic distance until the point at which the 141 main groups of tightly connected nodes still remained connected (in order to avoid the split of any 142 large network cluster from the main network). We estimated the classification of sampling sites 143 within network communities at each step of the pruning process with the "fastgreedy" community 144 145 detection algorithm implemented in IGRAPH (Clauset et al. 2004, Blondel et al. 2008). Network analysis (Fig. 1b), Bayesian-based STRUCTURE (Fig. 1c), and molecular variation (AMOVA) 146 (Supporting Information Table S1) revealed strong overall genetic differentiation among two 147 distinct lineages occupying the Tropical Indo-Pacific. Based on the landscape genetic analysis of 148 Cushman et al. (2014) and the definitions of global marine ecoregions (Spalding et al. 2007), we 149 classified these two lineages as distinct genotypes encompassed within two biogeographic regions: 150 the Western Tropical Indo-Pacific (WTIP) and the Central Tropical Indo-Pacific (CTIP). We then 151 152 used the two lineages in subsequent ecological niche modelling.

155 We collected a total of 62,465 presence records of *T. hemprichii* from a recently assembled and 156 cleaned dataset of global marine forests (Assis et al. 2020) and published literature (see Data 157 availability). In SDM studies, it is critical to correct for sampling bias and remove clustered 158 records, which may over-represent environmental conditions in better-surveyed regions (Kramer-159 Schadt *et al.* 2013). Therefore, presence records were filtered by: i) removing duplicated records at the resolution of our environmental predictors (i.e., keeping only one record per 5 arcmin grid 160 161 cell); ii) removing records on land or with distance to land > 370 km (following other SDM studies for coastal species; e.g., Zhang et al. 2020a), and iii) performing spatial thinning using a distance of 162 163 20 km using the R package spThin (Aiello-Lammens et al. 2015). This distance is a reasonable approximation of the dispersal potential for this plant traveling via floating propagules (Lacap *et al.* 164 2002), and it can also reduce potential effects of sampling bias while retaining sufficient numbers 165 of presence records for our analyses. As significant clustering was present in the data (particularly 166 around Australia), these procedures removed a large proportion (up to 99%) of the presence data. 167 Ultimately, we kept 519 records for the species-level model (hereafter "species model", records 168 from the entire region), 479 records for the CTIP lineage-level model (hereafter "CTIP model", 169 170 records within CTIP region only), and 26 records for the WTIP lineage-level model (hereafter "WTIP model", records within WTIP region only) (Fig. 1a). 171

172It is important to properly select the extent of the study area used to sample background173records when constructing presence-background SDMs for target species (Barve *et al.* 2011; Vale174*et al.* 2014). For coastal marine species, it is common practice to develop SDMs within the175Exclusive Economic Zone (i.e., within 370 km from the coast) (e.g., Lins *et al.* 2018; Stephenson *et al.* 2020; Zhang *et al.* 2020a). Besides, given the distributional range and records of *T. hemprichii*177from online repositories and literature (Fig. 1a), we restricted our study to the areas within 370 km178of land between 25°E and 180°E, and between 50°S and 40°N. Please note that our study extent

includes southern Australia and New Zealand, where this species does not naturally occur. It is
always challenging to estimate an appropriate study extent for a species (Barve *et al.* 2011), but the
extent we selected should represent the plausible accessible areas to *T. hemprichii* over
evolutionary time. We subsetted this main study extent to create separate study extents for the
WTIP and CTIP lineages (Fig. 1a) based on our molecular results (see details in the Lineage
genetic diversity in the Results section).

A number of marine predictors have been demonstrated to influence the geographical 185 distribution of marine species (Bosch et al. 2018). Based on previous studies (including seagrasses; 186 e.g., Jayathilake & Costello 2018; Zhang et al. 2020a), we initially considered twenty such 187 188 predictors for modeling, including two geographical predictors (water depth and distance to land) 189 from the Global Marine Environment Datasets (http://gmed.auckland.ac.nz; Basher et al. 2018) 190 and eighteen environmental predictors (including annual mean, maximum, minimum, range, average of the minimum records per year, and average of the maximum records per year) for SST, 191 sea surface salinity, and sea surface current velocity from the Bio-ORACLE database v2.1 192 (https://www.bio-oracle.org; Assis et al. 2018b). In SDM studies, highly collinear predictors can 193 lead to spurious interpretations of variable importance and unexpected predictions if correlations 194 195 change in different projection scenarios (Dormann et al. 2013). Hence, we checked collinearity by 196 calculating the pairwise Pearson's correlation coefficients (r) among the twenty predictors (Supporting Information Fig. S1) and selected one among highly correlated predictors (|r| > 0.7) 197 (Dormann et al. 2013) based on present-day and future data availability, biological importance, 198 and previous findings on important variables for estimating seagrass distribution (Javathilake & 199 Costello 2018). In the end, we retained the two geographical predictors and six environmental 200 201 predictors: annual mean current velocity, minimum current velocity, annual mean sea surface 202 salinity, annual range of sea surface salinity, annual mean SST, and annual range of SST.

To project future habitat suitability of *T. hemprichii*, we considered four representative concentration pathway (RCP) scenarios (i.e., RCP 2.6, RCP 4.5, RCP 6.0, and RCP 8.5), and two time periods (i.e., 2050s: the average for 2040–2050s, and 2100s: the average for 2090–2100). We obtained the corresponding projections of future marine environmental layers from the Bio-ORACLE database v2.1. We assumed that the two geographical predictors would remain unchanged for future projections (Zhang *et al.* 2020a).

209

210 Niche differentiation estimation

To estimate whether the two lineages of *T. hemprichii* occupy different niche spaces, we 211 212 characterized their realized niches using Hutchinsonian *n*-dimensional hypervolumes (Hutchinson 1957) sensu Blonder et al. (2018). We quantified the realized niches of the WTIP and 213 CTIP lineages using the eight selected marine predictor variables (see previous section). In short, 214 we extracted and standardized (i.e., zero means and unit variance) marine predictor values 215 associated with the presence records for the two lineages. We then determined the volumes and 216 shapes of the realized niches with the R package hypervolume using the Gaussian method 217 (Blonder 2019). We measured the extent of niche differentiation between the two lineages with 218 219 the *kernel.beta* function (Mammola & Cardoso 2020) in the R package *BAT* (Cardoso *et al.* 2015, 220 2020). Following Carvalho & Cardoso (2020), niche differentiation between hypervolumes was 221 partitioned into the following two processes: niche shift (replacement of space between hypervolumes) and niche contraction/expansion (net difference between hypervolumes). The 222 niche differentiation index ranges from 0 (niches overlap entirely) to 1 (niches are fully dissimilar) 223 (Carvalho & Cardoso 2020; Mammola & Cardoso 2020). In addition, to ascertain whether the 224 realized niches of the two lineages were still different after considering the environmental space 225 226 available, we conducted a niche similarity test (Broennimann et al. 2012) using the R package

227 *ecospat* (Di Cola *et al.* 2017). This test compares the empirical realized environmental niche of

228 CTIP with random niches permuted for WTIP over its available environmental space.

229

230 Species distribution modelling

231 We built SDMs using Maxent 3.4.4, a presence-background machine learning algorithm with two 232 main complexity tuning parameters: regularization multiplier, which penalizes complexity by removing predictors with low predictive ability, and feature class, which allows for increasing 233 234 complexity of the model response (Phillips et al. 2017). For each model (species model, WTIP model, and CTIP model), we randomly generated 10,000 background points within the 235 236 corresponding study region. As Maxent's default settings for the main tuning parameters can result 237 in overfit models (Radosavljevic & Anderson 2014), we used a version of the R package ENMeval 238 under expansion (1.9.0; https://github.com/jamiemkass/ENMeval) to tune our Maxent models over ranges of each parameter and chose models with optimal complexity based on performance 239 metrics calculated on withheld data (Muscarella et al. 2014). In brief, we considered a total of 32 240 candidate models with different combinations of regularization multipliers (RM; ranging from 0.5 241 to 4.0, at 0.5 interval), which penalize complexity more with higher values, and feature classes 242 243 (linear, quadratic, hinge), which allow responses with differing flexibility. Rather than using 244 conventional random cross-validation to judge model performance, we used a spatial block crossvalidation approach, which typically results in evaluations that better reflect the model's ability to 245 transfer to non-analog conditions (Roberts et al. 2017; Valavi et al. 2019). Briefly, each study 246 region was divided into four spatial blocks containing an equal number of presence records, three 247 blocks were used for model training and the remaining block for validation, then this procedure 248 was repeated until every block was used for model validation. As with previous studies (e.g., 249 250 Radosavljevic & Anderson 2014; Kass et al. 2020), the optimal model was selected by sequentially 251 considering a 10% omission rate (i.e., the percentage of validation presences with habitat

252 suitability predictions lower than that of the 10th quantile of training predictions), followed by 253 the area under the receiver operating characteristics curve (AUC) calculated on the validation data 254 (i.e., the model's ability to discriminate between presence and background records) to break ties. 255 We acknowledge that AUC is a poor measure for the absolute performance of presencebackground models (e.g., Jiménez-Valverde 2012), but nonetheless this metric can be used to make 256 257 relative comparisons of candidate models fitted with the same data (Lobo et al. 2008). 258 Predictive performances of the three best-performing Maxent models were further assessed 259 using the continuous Boyce index, a reliable evaluation measure of presence-only algorithms (Hirzel et al. 2006). The continuous Boyce index ranges from -1 to 1, where positive values 260 261 suggest that model predictions match well with the presence data, and negative values suggest a 262 poor match (Hirzel et al. 2006). Variable importance for each model was determined using 263 permutation importance calculated by Maxent. For this method, presence and background data values for each predictor variable in turn were randomly permuted and training AUC 264 recalculated—a large drop in AUC indicates higher importance (Phillips 2017). In addition, we 265 estimated the marginal response curves of important predictors (i.e., curves representing habitat 266 suitability along a range of the values of one predictor variable while keeping the other predictors 267 268 constant). We converted continuous habitat suitability predictions for *T. hemprichii* to binary 269 values using the same 10% omission thresholds that we used for model evaluation (Radosavljevic & Anderson 2014). We then transformed the binary habitat suitability projections to the Lambert 270 271 Cylindrical Equal Area projection at a resolution of 10 km and calculated areas of potential 272 distribution (Zhang et al. 2020a).

It is of great importance to consider species dispersal ability into SDMs when estimating climate change impacts (Araújo *et al.* 2006; Guisan *et al.* 2017). Given that species move across trans-regional across barriers in marine environments (Robinson *et al.* 2011; Pearman *et al.* 2020), and the relatively high dispersal ability of *T. hemprichii* (Lacap *et al.* 2002), we estimated range

277 size change under an unlimited dispersal scenario, which assumes that species have unrestricted

dispersal ability and can disperse to any suitable area (Araújo *et al.* 2006; Zhang *et al.* 2020c).

279 Range size change was calculated as follows:

280 range size change = $\frac{futuresuitablearea - presentsuitablearea}{presentsuitablearea} \times 100\%$,

281 where negative and positive values represent range contraction and expansion, respectively. We used the optimal species- and lineage-level models to make projections of future potential 282 distribution based on the different RCP scenarios for the two future time periods. Making 283 284 projections using SDMs into novel environmental space (i.e., outside the range of training data) results in some degree of extrapolations, which should be quantified to determine levels of 285 286 uncertainty (Elith et al. 2010). Therefore, we measured the similarity between present-day and future environmental conditions using multivariate environmental similarity surfaces (MESS) 287 288 (Elith et al. 2010). In practice, we calculated the MESS with the R package *rmaxent* (Baumgartner & Wilson 2021) for each model using the top three most important predictors via permutation 289 290 importance: positive MESS values indicate conditions more similar to the training data, while 291 negative values indicate conditions more different (i.e., novel).

292

293 Results

294 Lineage genetic diversity

We found significant genetic divergence between the populations of *T. hemprichii* in the Western
Tropical Indo-Pacific and Central Tropical Indo-Pacific regions. The genetic data for the two
lineages that we used for this analysis originated from disparate sources, and we found some minor
discrepancies (see Data availability) between the two datasets after carefully inspecting the
calibrated fragment lengths of the microsatellites (Hernawan *et al.* 2017; Jahnke *et al.* 2019a).
Regardless, even after deleting a few microsatellites (e.g., Thh41, TH07 and TH37), two genetic
lineages in *T. hemprichii* remained significantly diverged (i.e., CTIP and WTIP) across the

Tropical Indo-Pacific (Fig. 1b, 1c). Genetic variation among lineages accounted for 43.42% of the total genetic variation ($\Phi_{CT} = 0.43$, p < 0.0001; Supporting Information Table S1). Very limited genetic admixture was observed between the CTIP and WTIP lineages. The CTIP lineage harbored strikingly rich genetic diversity, with three times more alleles and allelic richness, and eight times fewer private alleles than the WTIP lineage (Supporting Information Table S2).

307

308 Niche differentiation between hypervolumes

The size of the realized niche of the CTIP lineage was one order of magnitude greater than that of 309 the WTIP lineage (CTIP lineage: 17295.6; WTIP lineage: 2273.2) (Fig. 2). Niche differentiation 310 311 between the two hypervolumes (0.97) was mainly due to variation in niche size (0.79), whereas 312 niche shift contributed only marginally (0.18). Difference in realized niches was easily distinguished via water depth and distance to land, with the WTIP lineage selecting a narrow 313 314 range of water depth and distance to land (Fig. 2). The two lineages also exhibited niche 315 differentiation with respect to annual mean sea surface salinity. In addition, the CTIP lineage niche was broader with respect to annual mean SST and annual range SST, whereas that of the 316 317 WTIP lineage was broader for annual mean current velocity, minimum current velocity, and annual range of sea surface salinity (Fig. 2). Niche differentiation between the two hypervolumes 318 319 was also high (0.86) when we considered only marine environmental predictors (i.e., excluding 320 water depth and distance to land) (Supporting Information Fig. S2). Regarding the niche similarity 321 analysis, after 1000 iterations the resulting *p*-values were above 0.05 for both overlap metrics 322 available in the *ecospat* package (0.10 for Schoener's D and 0.07 for Warren's I; Supporting 323 Information Fig. S3). This demonstrates that the realized environmental niche occupied by WTIP 324 is less similar to the niche occupied by CTIP when compared to random permutations over the full environmental space of WTIP. 325

327 Model performance

The tuning parameter settings with optimal complexity for the species-level and lineage-level 328 329 models ranged from relatively simple to complex. The optimal species-level model was the most complex (hinge features and 0.5 RM), while those for the lineage-level models were simpler 330 (CTIP: linear/quadratic/hinge features and 2.5 RM; WTIP: linear/quadratic features and 0.5 RM) 331 (Table 1). The average 10% omission rate was considerably lower for the WTIP lineage-level 332 333 model (3.57%) than for the other models (CTIP: 26.69%; species: 17.93%, Table 1) — as this was lower than the expectation of average 10% omission for the metric, it indicates that the optimal 334 settings results in models that may over-predict to some extent for WTIP. Although omission rate 335 336 was used primarily for model selection, the average validation AUC scores used to break ties were very high for all optimal models (Table 1); we think this is due to the fact that a majority of 337 presence data are in near-shore waters (Fig. 1a), which likely inflated the model's ability to 338 339 discriminate between these presences and background records in deeper water. In addition, all 340 three optimal models had relatively high continuous Boyce index scores (over 0.90; Table 1), indicating that final model predictions matched the presence data well. The eight predictors had 341 342 different levels of importance in the three models, but water depth and distance to land consistently played important roles (Table 2). In particular, these two predictors accounted for 343 344 more than 95% of permutation importance in the WTIP model (Table 2). For the CTIP and species 345 models, annual mean SST also had a high permutation importance ($^{29\%}$ and $^{24\%}$, respectively) 346 (Table 2). Response curves for water depth and distance to land suggest that shallow coastal waters 347 are more suitable for *T. hemprichii* (Supporting Information Fig. S4, Table S3).

348

349 Present-day habitat suitability projections

350 Under present-day conditions, species and lineage models projected similar but not identical

351 habitat suitability patterns, with a large part of the East African coast and the Pacific region as

352 suitable habitat for this species (Fig. 3). Compared with the species model, the CTIP model

353 predicted more southern distribution in Australia (Fig. 3c, 3d). In particular, the CTIP model

354 predicted suitable conditions in the Spencer Gulf, Southern Australia, where the species does not

aturally occur (Fig. 3a, 3c). The species model did not capture this pattern (Fig. 3b, 3d).

356 Moreover, the WTIP model identified more suitable habitat in the Red Sea than the species-level

357 model (Fig. 3c, 3d). Overall, species- and lineage-level models predicted comparable suitable areas

for *T. hemprichii* in the WTIP region (species model: 302,800 square km; WTIP model: 315,000

359 square km), while the species model predicted broader suitable area for the CTIP region (species

360 model: 1,873,800 square km; CTIP model: 1,757,900 square km).

361

362 Climate change impacts on habitat suitability

Species- and lineage-level models resulted in different future habitat suitability projections in the
CTIP region, with the lineage-level model resulting in predictions of more loss of suitable areas
(Table 3, Fig. 4). Both species- and lineage-level models predict considerable future loss of suitable
area in the CTIP region, especially on the Sunda Shelf (i.e., Indonesia and Malaysia) (Table 3, Fig.
Compared with the species model, the CTIP model projected more extensive range loss under
all climatic scenarios (Table 3). Interestingly, both models predicted that the species will shift
slightly southwards in Australia.

370 Species-level and lineage-level models predicted different impacts of climate change on 371 habitat suitability for *T. hemprichii* in the WTIP region (Table 3). The WTIP model predicted 372 range expansion (except under the RCP 2.6 scenario for the 2050s), whereas the species model 373 consistently indicated range contraction (Table 3). Overall, both species- and lineage-level models 374 predicted that future climate change marginally affects habitat suitability in the WTIP region and 375 that changes in range size were mostly < 15%, with the exception of a higher value ($^{2}24\%$) for the species model in the 2100s for the RCP 8.5 scenario (Table 3). The WTIP model predicted that 376 377 habitat suitability of *T. hemprichii* in the WTIP region will remain stable in the future, while the

378 species model predicted range contraction in the Red Sea and expansion in southern Madagascar379 and South Africa (Fig. 4).

Both species and CTIP models consistently showed that MESS values in the Sunda Shelf were slightly negative, which demonstrates small differences in climatic conditions between the present-day and future scenarios for this region (Supporting Information Fig. S5). For the WTIP region, the lineage and species model showed high environmental similarity except slight environmental dissimilarity in the Red Sea between present-day and future scenarios (Supporting Information Fig. S5). These results indicate a low degree of extrapolation in our model predictions.

387 Discussion

This study identified two diverged genetic lineages (WTIP and CTIP) in the seagrass T. hemprichii 388 across the tropical Indo-Pacific. The observed niche differentiation between the two lineages 389 390 suggests a violation of the niche conservatism assumption for species-level SDMs, and our lineage-391 level predictions of present and future range importantly avoid this assumption. Despite 392 differences between the habitat suitability predictions of the lineage-level and species-level SDMs, 393 they consistently predict that the CTIP lineage is at greater risk of range contraction in the future. Although genetic or genomic data that can be used to construct lineage-level SDMs are not always 394 395 available, our study emphasizes how incorporating information about phylogeographical structure 396 when modelling the impacts of climate change provides more realistic predictions to better understand future range shifts (Smith et al. 2019; Zhang et al. 2021). 397

398

399 Critical marine predictor variables for seagrasses

400 Both the lineage-level and species-level SDMs showed that distance to land, water depth, and

- 401 annual mean SST represent the most essential factors in explaining the distributional patterns of *T*.
- 402 hemprichii. The importance of these three predictors has been emphasized in previous studies of

403 Thalassia species (e.g., Duarte 1991; Lapointe et al. 1994; Fourqurean & Zieman 2002; Zhang et al. 2014) and other seagrasses (e.g., Baumstark et al. 2016; Jayathilake & Costello 2018). Further, 404 405 water depth and distance to land play significant roles in other efforts to map seagrass using WorldView-2 satellite imagery (Baumstark et al. 2016). Water depth correlates with the amount 406 407 of sunlight irradiance that reaches the seagrass, and with the rate of respiration of rhizomes and 408 shoot density, thereby constraining the extension of seagrass meadows and primary productivity to shallower areas (Duarte 1991). SST affects the photosynthesis, growth, and mortality of seagrass, 409 410 as demonstrated by field measurements and experimental manipulations of *T. hemprichii* (Collier & Waycott 2014; Pedersen et al. 2016; Rasmusson et al. 2020). In general, the total nitrogen and 411 412 phosphorus concentrations of the water column decrease with increasing distance from land, 413 despite a gradient of decreasing P limitation but increasing N limitation (Lapointe et al. 1994). 414 Nutrient inputs thus interact with changing temperatures to produce impacts on seagrass shoot densities, areal production rates, and biomass (Lapointe et al. 1994; Yamakita et al. 2011). The 415 predominant roles of the two geographical predictors and the negligible roles of marine 416 environmental predictors in the WTIP lineage-level model (Table 2) may partially explain the 417 marginal impacts of climate change predicted for this region. 418 419 Due to data availability, we used marine predictors at a spatial resolution of 5 arcmin (about 9.2 km at the equator). Within each grid cell, predictors such as temperature and salinity might be 420 421 homogeneous, but water depth can vary considerably. Seagrasses mainly inhabit shallow waters (Duarte 1991) and it would be reasonable to define study extent using water depth. We found that 422 when limiting the study region using water depth, our main conclusions regarding the change in 423 range size still held, but the predictive ability of the SDMs decreased (Supporting Information 424 425 Tables S4-S5). Given the limitations of the water depth layer and the decreased model 426 performance upon its inclusion, we decided to delineate the study region using distance to land.

427 To increase the reliability of marine SDMs, future efforts should be made to develop more high-

429

428

430 Incorporating intraspecific variation into SDMs for seagrasses

resolution marine predictors.

Seagrasses provide vital ecological services in marine ecosystems and SDMs have been applied to 431 432 this taxonomic group for multiple purposes (see reviews by Robinson et al. 2011; Robinson et al. 433 2017; Melo-Merino et al. 2020). Nonetheless, all previously reported SDMs on seagrasses were 434 built at the species level and thus have not considered possible intraspecific variation. For instance, Chefaoui et al. (2018) developed species-level SDMs for two seagrasses (Posidonia 435 436 oceanica and Cymodocea nodosa) in the Mediterranean Sea and predicted that the two species are 437 likely to experience dramatic habitat loss in the future. We fully agree that species-level SDMs are by definition informative, but given the high prevalence of intraspecific variation in marine 438 439 macrophytes (e.g., King et al. 2018), and the significance of intraspecific variation in SDMs (Benito 440 Garzón et al. 2019; Smith et al. 2019; Zhang et al. 2021; Collart et al. 2021), incorporating 441 intraspecific genetic variation into forecasts of seagrass distribution should result in more realistic scenarios of the potential consequences of climate change, providing that adaptive intraspecific 442 443 variation can be distinguished from clonal variation for seagrass populations under changing 444 conditions.

The importance of taxonomic resolution in SDMs has been addressed in several terrestrial and freshwater species, but much more sparsely for marine species (see review by Smith *et al.* 2019; Collart *et al.* 2021). Species-level SDMs that disregard existing intraspecific variation can either over- or under-estimate climate change impact on distributional change. For instance, species-level models for the lodgepole pine *Pinus contorta* consistently predicted more extreme habitat loss than subspecies-level models (Oney *et al.* 2013). As another example, although a species-level model for the reef-building coral *Porites lobata* predicted over 5% habitat expansion,

when modelling this species as five genetically isolated subpopulations the prediction was ca. 50% 452 453 habitat loss (Cacciapaglia & van Woesik 2018). In the present study, the species model consistently 454 predicted low impacts of climate change in the CTIP region in comparison to the lineage model (e.g., the habitat loss vs. stability in the Sunda Shelf in Fig. 4c vs. Fig. 4d). As for the WTIP region, 455 456 we found the opposite pattern. Here, the lineage model predicted stable future habitats in the 457 southern Red Sea (Fig. 4c), whereas the species model predicted habitat loss, including to the north of Mauritius (Fig. 4d). In addition, both species and lineage models predict a southward 458 459 range expansion in the southern CTIP, but only the species model clearly predicts this in the WTIP. Southern expansion is likely correlated with future temperature increases in areas which 460 are now too cold (Supporting Information Fig. S6). We should note that MESS values in the 461 equatorial regions were slightly negative, which indicates novel future environmental conditions. 462 This is due in part to higher future SST values for this region than those used by the present-day 463 SDM (Supporting Information Fig. S7)-thus, SDM projections in this region should be associated 464 465 with more uncertainty.

466 That the difference between the future projections for the two lineages could be attributed to the large difference in sample size (479 records for CTIP and 26 records for WTIP) is unlikely, 467 468 because. i) compared with other algorithms, Maxent is less sensitive to sample size and has better performance for small sample sizes (e.g., Hernandez et al. 2006; Wisz et al. 2008); ii) models for 469 both lineages had similarly high predictive abilities when using cross-validation with spatial 470 partitioning, which results in lower performance metrics for overfit models than conventional 471 random partitioning (Roberts et al. 2017). Further, as the range of WTIP is much larger than that 472 of CTIP, it is reasonable that less occurrence data would be necessary to properly characterize the 473 occupied environments for CTIP. However, SDMs were developed in this study without 474 475 considering physiological information of the species, and traits related to dispersal and resilience 476 to climate change. Such considerations in a mechanistic SDM could result in different future

477 range projections as compared with the results obtained with our approach. Further studies

involving both field investigations and associated data updates and methodological developments
for models [e.g., developing ensembles of small models (Breiner *et al.* 2018) or changing the study
extent] would further improve our predictions for climate change impacts on *T. hemprichii* in the
Tropical Indo-Pacific.

482

483 Intraspecific variation and local adaptation in seagrass

Differences in response to thermal changes related to intraspecific variation, whether eco-484 physiological or evolutionary, are well-documented in seagrasses (King et al. 2018). This variation, 485 486 partly based on phenotypic plasticity or local adaptation, ultimately might permit seagrasses to acclimatize and adapt to changes in climate (Duarte et al. 2018). The marine predictor variables 487 that played a predominant role in our SDMs (e.g., annual mean SST and water depth) could be 488 responsible for both long- and short-term local adaptation of *T. hemprichii* to a changing climate 489 490 (King et al. 2018; Jahnke et al. 2019b). In support of this, common-garden experiments have revealed a clear local adaptation to increased temperatures in Zostera marina (Franssen et al. 2011; 491 492 2014), and to a depth gradient in Posidonia oceanica (Marín-Guirao et al. 2016; Jahnke et al. 2019b). Further, parallel adaptation of Z. marina to thermal clines along the American and 493 494 European coasts was demonstrated using a space-for-time substitution design and gene expression 495 profiling (Jueterbock et al. 2016). Such adaptive local differentiation induced by divergent environmental forces (e.g., light, depth and temperature) has led to structured populations and 496 497 lineages in seagrasses at various spatial scales (Dattolo et al. 2014; Jueterbock et al. 2016; Jahnke et 498 al. 2019b), suggesting that adaptation to local conditions is a key mechanism for seagrasses to face 499 global climate change.

Although we identified high genetic differentiation between the WTIP and CTIP lineages
(Supporting Information Table S2), which may represent an extreme case of intraspecific genetic

502 divergence, we were not able to ascertain the adaptive components of divergence across a 503 common landscape within the tropical Indo-Pacific. In *T. hemprichii*, natural selection imposed 504 by environmental heterogeneity might have resulted in the evolution of locally adapted 505 populations with considerable variation in productivity, growth rate and competitive interactions 506 (Martins & Bandeira 2001; Lyimo et al. 2006; Larkum et al. 2018). Low genetic difference between 507 lineages with associated environments that are very different is likely the result of admixture 508 between geographically distant populations over evolutionary time, resulting in the species having 509 a wide range and a broad fundamental niche. In this case, lineage-level SDMs would not be appropriate and a full-species SDM should be used because the populations are distant spatially but 510 511 not genetically.

512 Future studies should focus on distinguishing neutral genetic differentiation from local 513 adaptation using reciprocal transplant trials (e.g., common gardens and provenance trials; see 514 Joyce & Rehfeldt 2013; Ralph et al. 2018). Also, it is most important to mechanistically assess the 515 sub-lethal susceptibility of *T. hemprichii* to thermal stress, including the effects of heat and 516 hypoxia on photosynthesis, respiration, and primary productivity (Pedersen et al. 2016; Rasmusson et al. 2020), before the strongest impacts of future climate change are sustained. 517 518 Intraspecific genetic diversity across populations can increase a species' adaptive capacity and 519 result in cascading effects to the entire ecosystem (Evans et al. 2017). It is thus important to identify the most temperature-tolerant genotypes from the WTIP and CTIP lineages, perhaps by 520 manipulating temperature to quantify the performance of individual genotypes of *T. hemprichii* 521 522 across thermal gradients. It is also essential to clarify whether genotype complementarity or dominance enhance the adaptive capacity in a population (Hughes & Stachowicz 2011). 523 524

525 Conservation implications 526 The challenge of designing effective actions for seagrass conservation in the Indo-Pacific exists in 527 the gap between science, policy, and practice (Fortes 2018). In this study, the separation in 528 geographic distribution and high niche differentiation between the CTIP and WTIP lineages 529 suggest that *T. hemprichii* populations may be locally adapted (Merilä & Hendry 2014). For 530 species with significant intraspecific genetic diversity, it is crucial to help maintain the species' 531 potential for adaptive responses to climate change by conserving this diversity (D'Amen et al. 532 2013). In particular, lineage differentiation can be explained by recruitment rate (Lyimo et al. 533 2006; Sherman et al. 2018), nutrient resorption (Martins & Bandeira 2001), dispersal barriers (Melroy et al. 2017), and evolutionary history from the origin center to the distributional margins 534 535 (Mukai 1993). Dramatic future habitat loss in the CTIP was predicted by both the species- and 536 lineage-level models (Fig. 4), stressing the urgency to develop monitoring programs to rescue 537 evolutionary and/or ecologically important units in *T. hemprichii*, particularly the populations and gene pools that have persisted through past long-term climate change because of local 538 539 adaptation (Bell 2017; Hernawan et al. 2017). Furthermore, the recognition of high niche differentiation between the WTIP and CTIP lineages may help to establish coherent principles 540 and regulating practices by which the different areas that *T. hemprichii* inhabits can be protected 541 542 efficiently.

543 The biomass, abundance, and productivity of seagrasses are highly correlated with both habitat suitability (Martins & Bandeira 2001; Saunders et al. 2013) and epiphytic species 544 biodiversity (Lyimo et al. 2008). Optimizing productivity of T. hemprichii in a given site or 545 546 population can help to increase associated community diversity (Eklöf et al. 2006; Lyimo et al. 2008). Thus, it is necessary to explore how community diversity and structure correlate with the 547 548 genetic composition and structure of the foundational species *T. hemprichii*. Such research can 549 help validate the results of SDMs in this study and quantify the relationship between *T*. 550 hemprichii and its relevant community components (Ikeda et al. 2017). Since populations in each

of the CTIP and WTIP lineages are locally adapted, policymakers and stakeholders are encouraged

552 to use local seed sources of *T. hemprichii* to ensure management strategies for successful

restoration and conservation purposes. To this end, mechanistic studies underlying thermal

adaptation by linking ecology to genetics should be done to better understand how *T. hemprichii*

will adapt to climate change (Duarte *et al.* 2018; Hu *et al.* 2020).

556

557 Acknowledgements

558 We thank Xiaokai Fan (Ocean University of China, China) for collecting distribution data of

559 Thalassia hemprichii and Jinxin Zhou (University of Tokyo, Japan) for figure illustration. We

560 further thank Dr Aussanee Meekaew (Shizuoka University, Japan) for kindly providing us with

561 the nice pictures of *Thalassia hemprichii* in Figure 2. This study was supported by National

562 Natural Science Foundation of China (31971395, 41761144057), Thailand Research Fund

563 (RDG6130002), the Foundation for Science and Technology (FCT) through the projects

564 UID/Multi/04326/2019 and PTDC/BIA-CBI/6515/2020 and the transitional norm -

565 DL57/2016/CP1361/CT0035 to JA. SM acknowledges support from the European commission

566 (program H2020-MSCA-IF-2019, grant award: 882221). ZZ and MY acknowledge the financial

567 support from the Japan Society for the Promotion of Science Grant-in-Aid for Scientific Research

568 (C) (18K05780). JMK acknowledges support from the Japan Society for the Promotion of Science

569Postdoctoral Fellowships for Foreign Researchers program and the Okinawa Institute of Science

570 and Technology Graduate University.

571

572 References

573 Aiello-Lammens, M. E., Boria, R. A., Radosavljevic, A., Vilela, B., & Anderson, R. P. (2015).

574 spThin: an R package for spatial thinning of species occurrence records for use in ecological

575 niche models. *Ecography*, **38**, **541–545**.

- 576 Araújo, M. B., Thuiller, W., & Pearson, R. G. (2006). Climate warming and the decline of
- amphibians and reptiles in Europe. *Journal of Biogeography*, 33, 1712–1728.
- Assis, J., Fragkopoulou, E., Frade, D., Neiva, J., Oliveira, A., Abecasis, D., ... & Serrão, E. A. (2020).
 A fine-tuned global distribution dataset of marine forests. *Scientific Data*, 7, 119.
- 580 Assis, J., Serrão, E. A., C. Coelho, N., Tempera, F., Valero, M., & Alberto, F. (2018a). Past climate
- changes and strong oceanographic barriers structured low latitude genetic relics for the golden
 kelp *Laminaria ochroleuca. Journal of Biogeography*, 45, 2326–2336.
- 583 Assis, J., Tyberghein, L., Bosch, S., Verbruggen, H., Serrão, E. A., & De Clerck, O. (2018b). Bio-
- ORACLE v2.0: Extending marine data layers for bioclimatic modelling. *Global Ecology and Biogeography*, 27, 277–284.
- 586 Barve, N., Barve, V., Jiménez-Valverde, A., Lira-Noriega, A., Maher, S. P., Peterson, A. T., ... &
- 587 Villalobos, F. (2011). The crucial role of the accessible area in ecological niche modeling and
 588 species distribution modeling. *Ecological Modelling*, 222, 1810–1819.
- 589 Basher, Z., Bowden, D. A., & Costello, M. J. (2018). Global Marine Environment Datasets (GMED).
- 590 World Wide Web electronic publication. Version 2.0 (Rev.02.2018). Accessed at
- 591 http://gmed.auckland.ac.nz
- Baumgartner, J., & Wilson, P. (2021). *rmaxent*: Tools for working with Maxent in R. R package
 version 0.8.5.9000. https://github.com/johnbaums/rmaxent
- Baumstark, R., Duffey, R., & Pu, R. L. (2016). Mapping seagrass and colonized hard bottom in
 Springs Coast, Florida using WorldView-2 satellite imagery. *Estuarine, Coastal and Shelf Science*, 181, 83–92.
- Bell, G. (2017). Evolutionary rescue. *Annual Review of Ecology, Evolution and Systematics*, 48,
 605–627.
- 599 Benito Garzón, M., Robson, T. M., & Hampe, A. (2019). ΔTrait SDMs: species distribution models
- 600 that account for local adaptation and phenotypic plasticity. *New Phytologist*, 222, 1757–1765.
- Blondel, V. D., Guillaume, J. L., Lambiotte, R., & Lefebvre, E. (2008). Fast unfolding of
- 602 communities in large networks. *Journal of Statistical Mechanics*, P10008.
- Blonder, B., Morrow, C. B., Maitner, B., Harris, D. J., Lamanna, C., Violle, C., ... & Kerkhoff, A. J.
- 604 (2018). New approaches for delineating *n*-dimensional hypervolumes. *Methods in Ecology and*605 *Evolution*, 9, 305–319.
- 606 Blonder, B., with contributions from Harris, D. J. (2019). Hypervolume: High Dimensional
- 607 Geometry and Set Operations Using Kernel Density Estimation, Support Vector Machines, and
- 608 Convex Hulls. R package version 2.0.12. https://CRAN.R-project.org/package=hypervolume

- Bosch, S., Tyberghein, L., Deneudt, K., Hernandez, F., & De Clerck, O. (2018). In search of
- 610 relevant predictors for marine species distribution modelling using the MarineSPEED
- 611 benchmark dataset. *Diversity and Distributions*, 24, 144–157.
- Breiner, F. T., Nobis, M. P., Bergamini, A., & Guisan, A. (2018). Optimizing ensembles of small
- models for predicting the distribution of species with few occurrences. *Methods in Ecology and Evolution*, 9, 802–808.
- Brodie, G., Holland, E., N'Yeurt, A. D. R., Soapi, K., & Hills, J. (2020). Seagrasses and seagrass
- 616 habitats in Pacific small island developing states: Potential loss of benefits via human
- disturbance and climate change. *Marine Pollution Bulletin*, 160, 111573.
- Broennimann, O., Fitzpatrick, M. C., Pearman, P. B., Petitpierre, B., Pellissier, L., Yoccoz, N. G., ...
- 619 & Guisan, A. (2012). Measuring ecological niche overlap from occurrence and spatial
- 620 environmental data. *Global Ecology and Biogeography*, 21, 481–497.
- 621 Cacciapaglia, C., & van Woesik, R. (2018). Marine species distribution modelling and the effects of
- 622 genetic isolation under climate change. *Journal of Biogeography*, 45, 154–163.
- 623 Carvalho, J. C., & Cardoso, P. (2020). Decomposing the causes for niche differentiation between
 624 species using hypervolumes. *Frontiers in Ecology and Evolution*, 8, 243.
- 625 Cardoso, P., Mammola, S., Rigal, F., & Carvalho, J. C. (2020). BAT: Biodiversity Assessment Tools.
- 626 R package version 2.0.1. https://CRAN.R-project.org/package=BAT
- 627 Cardoso, P., Rigal, F., & Carvalho, J. C. (2015). BAT–Biodiversity Assessment Tools, an R package
- for the measurement and estimation of alpha and beta taxon, phylogenetic and functional
- diversity. *Methods in Ecology and Evolution*, 6, 232–236.
- 630 Chefaoui, R. M., Duarte, C. M., & Serrão, E. A. (2018). Dramatic loss of seagrass habitat under
- 631 projected climate change in the Mediterranean Sea. *Global Change Biology*, 24, 4919–4928.
- 632 Cheung, W. W. L., Watson, R., & Pauly, D. (2013). Signature of ocean warming in global fisheries
 633 catch. *Nature*, 497, 365–368.
- 634 Clauset, A., Newman, M., & Moore, C. (2004). Finding community structure in very large
- 635 networks. *Physical Review E*, **70**, **066**111.
- 636 Coles, R., Grech, A., Rasheed, M., McKenzie, L., Unsworth, R., & Short, F. (2011). Seagrass
- 637 ecology and threats in the tropical Indo-Pacific bioregion. In: Roberts SP (ed) Seagrass: Ecology,
- Uses and Threats. Nova Science Publishers, pp 225–239.
- 639 Csardi, G., & Nepusz, T. (2006). The IGRAPH software package for complex network research.
- 640 *InterJournal Complex Systems*, 1695.

- 641 Collart, F., Hedenäs, L., Broennimann, O., Guisan, A., & Vanderpoorten, A. (2021). Intraspecific
- differentiation: Implications for niche and distribution modelling. *Journal of Biogeography*, 48,
 415–426.
- 644 Collier, C. J., & Waycott, M. (2014). Temperature extremes reduce seagrass growth and induce
 645 mortality. Marine Pollution Bulletin, 83, 483–490.
- 646 Cushman, S. A., Max, T., Meneses, N., Evans, L. M., Ferrier, S., Honchak, B., ... & Allan, G. J.
- (2014). Landscape genetic connectivity in a riparian foundation tree is jointly driven by climatic
 gradients and river networks. *Ecological Applications*, 24, 1000–1014.
- 649 D'Amen, M., Zimmermann, N. E., & Pearman, P. B. (2013). Conservation of phylogeographic
- 650 lineages under climate change. *Global Ecology and Biogeography*, 22, 93–104.
- Dattolo, E., Ruocco, M., Brunet, C., Lorenti, M., Lauritano, C., D'Esposito, D., DeLuca, P., Sanges,
- 652 R., Mazzuca, S., & Procaccini, G. (2014). Response of the seagrass *Posidonia oceanica* to
- different light environments: Insights from a combined molecular and photo-physiological
- 654 study. *Marine Environmental Research*, 101, 225–236.
- Di Cola, V., Broennimann, O., Petitpierre, B., Breiner, F. T., d'Amen, M., Randin, C., ... & Guisan,
- A. (2017). ecospat: an R package to support spatial analyses and modeling of species niches and
 distributions. *Ecography*, 40, 774–787.
- Dormann, C. F., Elith, J., Bacher, S., Buchmann, C., Carl, G., Carré, G., ... Lautenbach, S. (2013).
- 659 Collinearity: a review of methods to deal with it and a simulation study evaluating their
- 660 performance. *Ecography*, 36, 27–46.
- 661 Duarte, C. M. (1991). Seagrass depth limits. Aquatic Botany, 40, 363–377.
- Duarte, B., Martins, I., Rosa, R., Matos, A. R., Roleda, M. Y., Reusch, T. B. H., ... & Jueterbock, A.
- 663 (2018). Climate change impacts on seagrass meadows and macroalgal forests: an integrative
- 664 perspective on acclimation and adaptation potential. *Frontiers in Marine Sciences*, 5, 190.
- Elith, J., Kearney, M., & Phillips, S. (2010). The art of modelling range-shifting species. *Methods in Ecology and Evolution*, 1, 330–342.
- Eklöf, J. S., Henriksson, R., & Kautsky, N. (2006). Effects of tropical open-water seaweed farming
 on seagrass ecosystem structure and function. *Marine Ecology Progress Series*, 325, 73–84.
- 669 Evans, S. M., Vergés, A., & Poore, A. G. B. (2017). Genotypic diversity and short-term response to
- 670 shading stress in a threatened seagrass: does low diversity mean low resilience? *Frontiers in*
- 671 *Plant Science*, 8, 1417.

- 672 Fourqurean, J. W., & Zieman, J. C. (2002). Nutrient content of the seagrass Thalassia testudinum
- reveals regional patterns of relative availability of nitrogen and phosphorus in the Florida Keys
- 674 USA. *Biogeochemistry*, 61, 229–245.
- Fortes, M. D. (2018). Seagrass ecosystem conservation in Southeast Asia needs to link science to
 policy and practice. *Ocean and Coastal Management*, 159, 51–56.
- 677 Franssen, S. U., Gu, J., Bergmann, N., Winters, G., Klostermeier, U. C., Rosenstiel, P., Bornberg-
- Bauer, E., & Reusch, T. B. H. (2011). Transcriptomic resilience to global warming in the seagrass
- 679 *Zostera marina*, a marine foundation species. *Proceedings of the National Academy of Sciences*
- 680 USA, 108, 19276–19281.
- 681 Franssen, S. U., Gu, J., Winters, G., Huylmans, A. K., Wienpahl, I., Sparwel, M., Coyer, J. A.,
- Olsen, J. L., Reusch, T. B. H., Bornberg-Bauer, E. (2014). Genome-wide transcriptomic

responses of the seagrasses *Zostera marina* and *Nanozostera noltii* under a simulated heatwave
confirm functional types. *Marine Genomics*, 15, 65–73.

- 685 Grech, A., Chartrand-Miller, K., Erftemeijer, P., Fonseca, M., McKenzie, L, Rasheed, M., ... &
- Coles, R. (2012). A comparison of threats, vulnerabilities and management approaches in global
 seagrass bioregions. *Environmental Research Letters*, 7, 024006.
- Green, E. E. P., & Short, F. T. (2003). World atlas of seagrasses. Berkeley, CA: University of
 California Press.
- Guisan, A., Thuiller, W., & Zimmermann, N. E. (2017). Habitat Suitability and Distribution
 Models: With Applications in R. Cambridge University Press, Cambridge.
- Hao, T., Elith, J., Lahoz-Monfort, J. J., & Guillera-Arroita, G. (2020). Testing whether ensemble
- modelling is advantageous for maximising predictive performance of species distribution
 models. *Ecography*, 43, 549–558.
- 695 Hernandez, P. A., Graham, C. H., Master, L. L., & Albert, D. L. (2006). The effect of sample size
- and species characteristics on performance of different species distribution modeling methods.
 Ecography, 29, 773–785.
- 698 Hernawan, U., van Dijk K., Kendrick, G., Feng, M., Biffin, E., ... & McMahon, K. (2017).
- Historical processes and contemporary ocean currents drive genetic structure in the seagrass
 Thalassia hemprichii in the Indo-Australian Archipelago. *Molecular Ecology*, 26, 1008–1021.
- 701 Hirzel, A. H., Le Lay, G., Helfer, V., Randin, C., & Guisan, A. (2006). Evaluating the ability of
- habitat suitability models to predict species presences. *Ecological modelling*, 199, 142–152.
- 703 Hughes, A. R., & Stachowicz, J. J. (2011). Seagrass genotypic diversity increases disturbance
- response via complementarity and dominance. *Journal of Ecology*, **99**, **445–453**.

- 705 Hultine, K. R., Grady, K. C., Wood, T. E., Shuster, S. M., Stella, J. C., Whitham, T. G. (2016).
- Climate change perils for dioecious plant species. *Nature Plants*, 2, 16109.
- Hutchinson, G. (1957). Concluding remarks. *Cold Spring Harbor Symposia on Quantitative Biology*, 22, 415–427.
- 709 Hyndes, G. A., Heck, K. L., Vergés, A., Harvey, E. S., Kendrick, G. A., Lavery, P. S., ... & Wilson,
- S. (2016). Accelerating tropicalization and the transformation of temperate seagrass meadows. *BioScience*, 66, 938–948.
- 712 Ikeda, D. H., Max, T. L., Allan, G. J., Lau, M. K., Shuster, S. M., & Whitham, T. G. (2017).
- Genetically informed ecological niche models improve climate change predictions. *Global Change Biology*, 23, 164–176.
- 715 Jahnke, M., Gullström, M., Larsson, J., Asplund, M. E., Mgeleka, S., Silas, M. O., ... & Nordlund, L.
- 716 M. (2019a). Population genetic structure and connectivity of the seagrass *Thalassia hemprichii*
- in the Western Indian Ocean is influenced by predominant ocean currents. *Ecology and*
- 718 *Evolution*, 9, 8953–8964.
- 719 Jahnke, M., D'Esposito, D., Orru, L., Lamontanara, A., Dattolo, E., Badalamenti, F., Mazzuca, S.,
- Procaccini, G., & Orsini, L. (2019b). Adaptive responses along a depth and a latitudinal gradient
 in the endemic seagrass *Posidonia oceanica*. *Heredity*, 122, 233–243.
- Jayathilake, D. R., & Costello, M. J. (2018). A modelled global distribution of the seagrass biome.
 Biological Conservation, 226, 120–126.
- 724 Jiménez-Valverde, A. (2012). Insights into the area under the receiver operating characteristic
- curve (AUC) as a discrimination measure in species distribution modelling. *Global Ecology and Biogeography*, 21, 498–507.
- Johansson, M. L., Alberto, F., Reed, D. C., Raimondi, P. T., Coelho, N. C., Young, M. A., ... &
- Serrão, E. A. (2015). Seascape drivers of *Macrocystis pyrifera* population genetic structure in the
 northeast Pacific. *Molecular Ecology*, 24, 4866–4885.
- 730 Jordà, G., Marbà, N., & Duarte, C. M. (2012). Mediterranean seagrass vulnerable to regional
- 731 climate warming. *Nature Climate Change*, 2, 821–824.
- 732 Jueterbock, A., Franssen, S. U., Bergmann, N., Gu, J., Coyer, J. A., Reusch, T. B. H., Bonberg-
- 733 Bauer, E., & Olsen, J. L. (2016). Phylogeographic differentiation versus transcriptomic
- adaptation to warm temperatures in *Zostera marina*, a globally important seagrass. *Molecular*
- 735 *Ecology*, **25**, **5396–54**11.

- 736 Kass, J. M., Anderson, R. P., Espinosa-Lucas, A., Juárez-Jaimes, V., Martínez-Salas, E., Botello,
- F., ... & Sánchez-Cordero, V. (2020). Biotic predictors with phenological information improve
 range estimates for migrating monarch butterflies in Mexico. *Ecography*, 43, 341–352.
- 739 King, N. G., McKeown, N. J., Smale, D. A., & Moore, P. J. (2018). The importance of phenotypic
- 740 plasticity and local adaptation in driving intraspecific variability in thermal niches of marine
- 741 macrophytes. *Ecography*, 41, 1469–1484.
- 742 Kramer-Schadt, S., Niedballa, J., Pilgrim, J. D., Schröder, B., Lindenborn, J., Reinfelder, V., ... &
- Wilting, A. (2013). The importance of correcting for sampling bias in MaxEnt species
- distribution models. *Diversity and Distributions*, 19, 1366–1379.
- Lacap, C. D. A., Vermaat, J. E., Rollon, R. N., & Nacorda, H. M. (2002). Propagule dispersal of the
- SE Asian seagrasses *Enhalus acoroides* and *Thalassia hemprichii*. *Marine Ecology Progress Series*, 235, 75–80.
- 748 Lapointe, B. E., Tomasko, D. A., & Matzie, W. R. (1994). Eutrophication and trophic state
- classification of seagrass communities in the Florida Keys. *Bulletin of Marine Science*, 54, 696–
 750 717.
- Larkum, A. W. D., Pernice, M., Schliep, M., Davey, P., Szabo, M., Raven, J. A., Lichtenberg, M.,
- 752 Brodersen, K. E., & Ralph, P. J. (2018). Photosynthesis and metabolism of seagrasses. In: Larkum
- AWD, Kendrick GA and Ralph PJ (ed) Seagrasses of Australia: structure, ecology and
- conservation. Springer, pp 315–342.
- Lins, D. M., de Marco Jr, P., Andrade, A. F., & Rocha, R. M. (2018). Predicting global ascidian
 invasions. *Diversity and Distributions*, 24, 692–704.
- Lobo, J. M., Jiménez-Valverde, A., & Real, R. (2008). AUC: a misleading measure of the
 performance of predictive distribution models. *Global Ecology and Biogeography*, 17, 145–151.
- Lowe, W. H., & Allendorf, F. W. (2010). What can genetics tell us about population connectivity? *Molecular Ecology*, 19, 3038–3051.
- 761 Lowen, J. B., Hart, D. R., Stanley, R. R., Lehnert, S. J., Bradbury, I. R., & DiBacco, C. (2019).
- Assessing effects of genetic, environmental, and biotic gradients in species distribution
- modelling. *ICES Journal of Marine Science*, 76, 1762–1775.
- 764 Lyimo, T. J., Mvungi, E. F., Lugomela, C., & Björk, M. (2006). Seagrass biomass and productivity in
- seaweed and non-seaweed farming areas in the east coast of Zanzibar, Tanzania. *Western*
- 766 *Indian Ocean Journal of Marine Science*, 5, 141–152.

- 767 Lyimo, T. J., Mvungi, E. F., & Mgaya, Y. D. (2008). Abundance and diversity of seagrass and
- 768 macrofauna in the intertidal areas with and without seaweed farming activities on the east coast
- 769 of Zanzibar. *Tanzania Journal Science*, 34, 42–52.
- 770 Mammola, S., & Cardoso, P. (2020). Functional diversity metrics using kernel density *n*-
- dimensional hypervolumes. *Methods in Ecology and Evolution*, 11, 986–995.
- 772 Marín-Guirao, L., Ruiz, J. M., Dattolo, E., Garcia-Munoz, R., & Procaccini, G. (2016).
- 773 Physiological and molecular evidence of differential short-term heat tolerance in
- 774 Mediterranean seagrasses. *Scientific Reports*, 6, 28615.
- 775 Martins, A. R. O., & Bandeira, S. O. (2001). Biomass distribution and leaf nutrient concentrations
- and resorption of *Thalassia hemprichii* at Inhaca Island, Mozambique. *South African Journal of Botany*, 67, 439–442.
- Mukai, H. (1993). Biogeography of the tropical seagrasses in the western Pacific. *Australian Journal of Freshwater Research*, 44, 1–17.
- 780 Muscarella, R., Galante, P. J., Soley-Guardia, M., Boria, R. A., Kass, J. M., Uriarte, M., & Anderson,
- 781 R. P. (2014). ENMeval: An R package for conducting spatially independent evaluations and
- r82 estimating optimal model complexity for Maxent ecological niche models. *Methods in Ecology*r83 *and Evolution*, 5, 1198–1205.
- 784 Melo-Merino, S. M., Reyes-Bonilla, H., & Lira-Noriega, A. (2020). Ecological niche models and
- species distribution models in marine environments: A literature review and spatial analysis of
 evidence. *Ecological Modelling*, 415, 108837.
- 787 Melroy, L. M., Smith, R. J., & Cohen, C. S. (2017). Phylogeography of direct-developing sea stars
- in the genus *Leptasterias* in relation to San Francisco Bay outflow in central California. *Marine Biology*, 164, 152.
- Merilä, J., & Hendry, A. P. (2014). Climate change, adaptation, and phenotypic plasticity: the
 problem and the evidence. *Evolutionary Applications*, 7, 1–14.
- 792 Miyoshi, K., Hattori, R. S., Strüssmann, C. A., Yokota, M., & Yamamoto, Y. (2020).
- 793 Phenotypic/genotypic sex mismatches and temperature-dependent sex determination in a wild
- population of an Old World atherinid, the cobaltcap silverside *Hypoatherina tsurugae*.
- 795 *Molecular Ecology*, 29, 2349–2358.
- 796 Olsen, Y. S., Collier, C., Ow, Y. X., & Kendrick, G. A. (2018). Global warming and ocean
- acidification: effects on Australian seagrass ecosystems. In: Larkum AWD, Kendrick GA and
- Ralph PJ (ed) Seagrasses of Australia: structure, ecology and conservation. Springer, pp. 705–
- 799 **742**.

- 800 Orth, R. J., Carruthers, T. J. B., Dennison, W. C., Duarte, C. M., Fourqurean, J. W., Heck, Jr. K.
- L., ... & Williams, S. L. (2006). A global contemporary crisis for seagrass ecosystems. *Bioscience*,
 56, 987–996.
- Oney, B., Reineking, B., O'Neill, G., & Kreyling, J. (2013). Intraspecific variation buffers projected
 climate change impacts on *Pinus contorta. Ecology and Evolution*, 3, 437–449.
- 805 Pazzaglia, J., Reusch, T. B. H., Terlizzi, A., Marín-Guirao, L., & Procaccini, G. (2021). Phenotypic
- 806 plasticity under rapid global changes: the intrinsic force for future seagrasses survival.
- 807 *Evolutionary Applications*, 14, doi:10.1111/eva.13212.
- 808 Pearman, W. S., Wells, S. J., Silander, O. K., Freed, N. E., & Dale, J. (2020). Concordant geographic
- and genetic structure revealed by genotyping-by-sequencing in a New Zealand marine isopod.
- 810 *Ecology and Evolution*, 10, 13624–13639.
- 811 Pedersen, O., Colmer, T. D., Borum, J., Zavala-Perez, A., & Kendrick, G. A. (2016). Heat stress of
- 812 two tropical seagrass species during low tides impact on underwater net photosynthesis, dark
- respiration and diel *in situ* internal aeration. New Phytologist, 210, 1207–1218.
- 814 Peterson, M. L., Doak, D. F., & Morris, W. F. (2019). Incorporating local adaptation into forecasts
- of species' distribution and abundance under climate change. *Global Change Biology*, 25, 775–
 793.
- 817 Phillips, S. J. (2017). A Brief Tutorial on Maxent. Available from URL:
- 818 http://biodiversityinformatics.amnh.org/open_source/maxent/. Accessed on 2021-02-10.
- 819 Phillips, S. J., Anderson, R. P., Dudík, M., Schapire, R. E., & Blair, M. E. (2017). Opening the black
- box: An open-source release of Maxent. *Ecography*, 40, 887–893.
- Pinsky, M. L., Selden, R. L., & Kitchel, Z. J. (2020). Climate-driven shifts in marine species ranges:
 scaling from organisms to communities. *Annual Review of Marine Science*, 12, 153–179.
- 823 Radosavljevic, A., & Anderson, R. P. (2014). Making better Maxent models of species distributions:
- complexity, overfitting and evaluation. *Journal of Biogeography*, 41, 629–643.
- 825 Ralph, P. J., CRosswell, J. R., Cannard, T., & Steven, A. D. L. (2018). Estimating seagrass blue
- carbon and policy implications: the Australian perspective. In: Larkum AWD, Kendrick GA and
- Ralph PJ (ed) Seagrasses of Australia: structure, ecology and conservation. Springer, pp 743–758.
- 828 Rasmusson, L. M., Buapet, P., George, R., Gullström, M., Gunnarsson, P. C. B., & Björk, M. (2020).
- 829 Effects of temperature and hypoxia on respiration, photorespiration, and photosynthesis of
- seagrass leaves from contrasting temperature regimes. *ICES Journal of Marine Science*, 77,
- 831 **2056–2065**.

- 832 Razgour, O., Forester, B., Taggart, J. B., Bekaert, M., Juste, J., Ibáñez, C., ... & Manel, S. (2019).
- 833 Considering adaptive genetic variation in climate change vulnerability assessment reduces
- species range loss projections. *Proceedings of the National Academy of Sciences USA*, 116,
 10418–10423.
- 836 Repolho, T., Duarte, B., Dionísio, G., Paula, J. R., Lopes, A. R., Rosa, I. C., ... & Rosa, R. (2017).
- 837 Seagrass ecophysiological performance under ocean warming and acidification. *Scientific*838 *Reports*, 7, 41443.
- 839 Roberts, D. R., Bahn, V., Ciuti, S., Boyce, M. S., Elith, J., Guillera-Arroita, G., ... & Dormann, C. F.
- 840 (2017). Cross-validation strategies for data with temporal, spatial, hierarchical, or phylogenetic
 841 structure. *Ecography*, 40, 913–929.
- 842 Robinson, L. M., Elith, J., Hobday, A. J., Pearson, R. G., Kendall, B. E., Possingham, H. P., &
- Richardson, A. J. (2011). Pushing the limits in marine species distribution modelling: lessons
- from the land present challenges and opportunities. *Global Ecology and Biogeography*, 20, 789–
 802.
- 846 Robinson, N. M., Nelson, W. A., Costello, M. J., Sutherland, J. E., & Lundquist, C. J. (2017). A
- systematic review of marine-based species distribution models (SDMs) with recommendations
 for best practice. *Frontiers in Marine Science*, 4, 421.
- Sanciangco, J. C., Carpenter, K. E., Etnoyer, P. J., & Moretzsohn, F. (2013). Habitat availability and
 heterogeneity and the Indo-Pacific warm pool as predictors of marine species richness in the
 Tropical Indo-Pacific. *PLoS ONE*, 8, e56245.
- 852 Saunders, M. I., Leon, J., Phinn, S. R., Callaghan, D. P., O'Brien, K. R., Roelfsema, C. M., Lovelock,
- C. E., Lyons, M. B., & Mumby, P. J. (2013). Coastal retreat and improved water quality mitigate
 losses of seagrass from sea level rise. *Global Change Biology*, 19:2569–2583
- 855 Sherman, C. D. H., Smith, T. M., York, P. H., Jarvis, J. C., Ruiz-Montoya, L. R., & Kendrick, G. A.
- 856 (2018). Reproductive, dispersal and recruitment strategies in Australian seagrasses. In: Larkum
- AWD, Kendrick GA and Ralph PJ (ed) Seagrasses of Australia: structure, ecology and
- 858 conservation. Springer, pp 213–256.
- Short, F.T., Carruthers, T., Dennison, W., & Waycott M. (2007). Global seagrass distribution and
 diversity: A bioregional model. *Journal of Experimental Marine Biology and Ecology*, 350, 3–20.
- 861 Smale, D. A., Wernberg, T., Oliver, E. C. J., Thomen, M., Harvey, B. P., Straub, S. C., ... & Moore,
- P. J. (2019). Marine heatwaves threaten global biodiversity and the provision of ecosystem
- services. *Nature Climate Change*, 9, 306–312.

- 864 Smith, A. B., Godsoe, W., Rodríguez-Sánchez, F., Wang, H. H., & Warren, D. (2019). Niche
- 865 estimation above and below the species level. *Trends in Ecology & Evolution*, 34, 260–273.
- 866 Spalding, M. D., Fox, H. E., Allen, G. R., Davidson, N., Ferdana, Z. A., ... & Robertson, J. (2007).
- Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. *BioScience*, 57,
 573–583.
- 869 Stephenson, F., Goetz, K., Sharp, B. R., Mouton, T. L., Beets, F. L., Roberts, J., ... & Lundquist, C. J.
- 870 (2020). Modelling the spatial distribution of cetaceans in New Zealand waters. Diversity and
- 871 Distributions, 26, 495–516.
- 872 Thomson, J. A., Burkholder, D. A., Heithaus, M. R., Fourqurean, J. W., Fraser, M. W., ... &
- 873 Kendrick, G. A. (2015). Extreme temperatures, foundation species, and abrupt ecosystem
- change: an example from an iconic seagrass ecosystem. *Global Change Biology*, 21, 1463–1474.
- 875 Unsworth, R. K. F., McKenzie, L. J., Nordlund, L. M., & Cullen-Unsworth, L. C. (2018). A
- changing climate for seagrass conservation? *Current Biology*, 28, R1221–R1232.
- 877 Valavi, R., Elith, J., Lahoz-Monfort, J. J., & Guillera-Arroita, G. (2019). blockCV: An r package for
- generating spatially or environmentally separated folds for k-fold cross-validation of species
 distribution models. *Methods in Ecology and Evolution*, 10, 225–232.
- 880 Vale, C. G., Tarroso, P., & Brito, J. C. (2014). Predicting species distribution at range margins:
- testing the effects of study area extent, resolution and threshold selection in the Sahara–Sahel
 transition zone. *Diversity and Distributions*, 20, 20–33.
- 883 Waycott, M., Duarte, C.M., Carruthers, T. J. B., Orth, R. J., Dennison, W. C., Olyarnik, S., ... &
- Williams, S. L. (2009). Accelerating loss of seagrasses across the globe threatens coastal
- ecosystems. *Proceedings of the National Academy of Sciences USA*, 106, 12377–12381.
- 886 Wisz, M. S., Hijmans, R. J., Li, J., Peterson, A. T., Graham, C. H., & Guisan, A. & NCEAS
- 887 Predicting Species Distributions Working Group (2008). Effects of sample size on the
- performance of species distribution models. *Diversity and Distributions*, 14, 763–773.
- 889 Wu, K. Y., Chen, C. N. N., & Soong, K. (2016). Long distance dispersal potential of two seagrasses
- 890 *Thalassia hemprichii* and *Halophila ovalis. PLoS ONE*, 11, e0156585.
- Yamakita, T., Watanabe, K., & Nakaoka, M. (2011). Asynchronous local dynamics contributes to
 stability of a seagrass bed in Tokyo Bay. *Ecography*, 34, 519–528.
- 893 Zhang, J. P., Huang, X. P., & Jiang, Z. J. (2014) Physiological responses of the seagrass Thalassia
- 894 *hemprichii* (Ehrenb.) Aschers as indicators of nutrient loading. *Marine Pollution Bulletin*, 83,
- 895 **508–515**.

- 896 Zhang, Z., Capinha, C., Karger, D. N., Turon, X., MacIsaac, H. J., & Zhan, A. (2020a). Impacts of
- 897 climate change on geographical distributions of invasive ascidians. *Marine Environmental*898 *Research*, 104993.
- Zhang, Z., Mammola, S., McLay, C. L., Capinha, C., & Yokota, M. (2020b). To invade or not to
 invade? Exploring the niche-based processes underlying the failure of a biological invasion
- using the invasive Chinese mitten crab. *Science of The Total Environment*, 138815.
- 902 Zhang, Z., Mammola, S., Liang, Z., Capinha, C., Wei, Q., Wu, Y., ... & Wang, C. (2020c). Future
- 903 climate change will severely reduce habitat suitability of the Critically Endangered Chinese
- giant salamander. *Freshwater Biology*, 65, 971–980.
- 905 Zhang, Z., Kass, J. M., Mammola, S., Koizumi, I., Li, X., Tanaka, K., ... & Usio, N. (2021). Lineage-
- 906 level distribution models lead to more realistic climate change predictions for a threatened
- 907 crayfish. *Diversity and Distributions*, 27, 684–695.
- 908

909 Data availability

- 910 Environmental layers can be downloaded from the Global Marine Environment Datasets
- 911 (http://gmed.auckland.ac.nz) and the Bio-ORACLE database v2.1 (https://www.bio-oracle.org).
- 912 Microsatellite datasets and presence records of *Thalassia hemprichii* have be deposited in Dryad
- 913 entry https://doi.org/10.5061/dryad.vhhmgqnsh.
- 914

915 Author contributions

- 916 Z.-M.H. and Z.Z. conceived the project. Z.-M.H. and J.Z. collected molecular data, J.Z. and P.F.
- 917 performed genetic analyses, Z.Z. collected species distribution data and conducted ecological
- niche analyses with critical suggestions from J.M.K. and S.M. Z.-M.H., Z.Z., S.M., J.M.K., Z.-Q.Z.,
- 919 S.D., J.A., and A.J. interpreted and discussed the results, wrote and/or revised the manuscript. All
- 920 authors approved the final version of the manuscript.

922 **Tables**

923 Table 1. Maxent parameters, performance metrics on spatially withheld data (validation AUC and

924 omission rate) and the full dataset (continuous Boyce index), and 10th percentile presence

925 thresholds of the two lineage-level models and the species-level model. Validation AUC and 10%

926 omission rate results are expressed as means ± standard deviation across spatial partitions.

Model	$\mathbf{R}\mathbf{M}^1$	Feature class²	Average validation AUC	Average 10% omission rate (%)	10% omission threshold	Continuous Boyce index
WTIP ³	0.5	LQ	0.99(± 0.01)	3.57(± 7.14)	0.12	0.92
CTIP ⁴	2.5	LQH	$0.96(\pm 0.03)$	26.69(± 37.42)	0.37	0.99
Species	0.5	Н	$0.96(\pm 0.02)$	17.93(± 17.62)	0.30	0.99

927 ¹RM: regularization multiplier.

928 ²Feature Class: L (linear), Q (quadratic), and H (hinge) were considered.

929 ³WTIP: the Western Tropical Indo-Pacific lineage-level model

930 ⁴CTIP: the Central Tropical Indo-Pacific lineage-level model

932 **Table 2**. Permutation importance (%) of marine predictors in each Maxent model. Values in bold

933 showed important predictors.

Predictors	WTIP ¹	CTIP ²	Species
Water depth [m]	14.10	51.05	46.91
Distance to land [km]	81.87	11.58	9.30
Minimum current velocity [m/s]	0.46	0.00	0.03
Annual mean current velocity [m/s]	0.15	0.04	0.27
Annual mean sea surface salinity [PSS]	0.80	2.49	1.92
Annual range of sea surface salinity [PSS]	0.93	4.96	13.38
Annual mean sea surface temperature [°C]	0.83	28.68	23.47
Annual range of sea surface temperature [°C]	0.87	1.20	4.73

934 ¹WTIP: Western Tropical Indo-Pacific lineage-level model

935 ²CTIP: Central Tropical Indo-Pacific lineage-level model

936

938 **Table 3.** Range size change (%) of *Thalassia hemprichii* under future climate scenarios. Values in

RCP ¹		WTIP ²	(CTIP ³		
KGI	2050s	2100s	2050s	2100s		
RCP 2.6	-0.2 (-2.5)	4.4 (-3.8)	-40.6 (-29.3)	-34.5 (-27.8)		
RCP 4.5	1.3 (-6.3)	4.2 (-6.3)	-49.7 (-26.1)	-55.7 (-26.4)		
RCP 6.0	3.4 (-0.4)	7.5 (-15.0)	-43.8 (-30.3)	-63.7 (-23.2)		
RCP 8.5	4.0 (-10.9)	13.2 (-23.7)	-53.7 (-27.4)	-72.1 (-25.8)		

939 parentheses indicate range size change predicted by the species-level model.

940 ¹RCP: representative concentration pathway.

941 ²WTIP: Western Tropical Indo-Pacific lineage-level model

942 ³CTIP: Central Tropical Indo-Pacific lineage-level model

943

945 Figure Legends

946 Figure 1. (a) Map of study regions and presence records used in this study. Blue and red points represent sample collection locations for molecular analyses in the Western Tropical Indo-Pacific 947 948 (WTIP) and the Central Tropical Indo-Pacific (CTIP). Gray crosses show presence records used for 949 the species distribution models. (b) Network of microsatellite genetic differentiation (Cavalli-950 Sforza and Edwards chord distances). The topology results from pruning the network for pairwise 951 genetic distances <0.534. The smallest chord distance (0.499) between the WTIP and CTIP 952 lineages is shown. (c) Genetic lineage division over space based on STRUCTURE clustering (k = 2). 953 Population abbreviations are the same as in Hernawan et al. (2017) and Jahnke et al. (2019a), and 954 their classification to network analysis is in agreement with that of STRUCTURE (Fig. 1c).

Figure 2. The realized niches for the two lineages of *Thalassia hemprichii* quantified via eightdimensional hypervolumes. The axes represent unitless values as the predictors were standardized

957 before analyses. Circles with white rims indicate hypervolume centroids. Boundaries and shapes of

958 hypervolumes were delineated by 10,000 points randomly sampled within each hypervolume. The

959 photograph depicts *T. hemprichii*.

Figure 3. Present-day continuous (a, b) and binary (c, d) habitat suitability predictions for *Thalassia hemprichii* by lineage-level (a, c) and species-level (b, d) Maxent models. Dashed lines
represent the equator. To improve the legibility of the binary predictions, we increased the pixel
size by downscaling the spatial resolution to 30 arcmin. High-resolution predictions can be
downloaded from Dryad (https://doi.org/10.5061/dryad.vhhmgqnsh).

Figure 4. Changes in continuous (a, b) and binary (c, d) habitat suitability for *Thalassia hemprichii* projected by lineage-level (a, c) and species-level (b, d) Maxent models under the RCP 8.5 scenario in the 2050s. Dashed lines indicate the equator. The category "stable" represents areas predicted to be suitable under both present-day and future climatic conditions, "loss" indicates areas predicted to be suitable under present-day conditions but unsuitable in the future, and "gain" indicates areas predicted to be unsuitable under present-day conditions but suitable in the future. To improve the legibility of the binary predictions, we increased the pixel size by downscaling the spatial

- 972 resolution to 30 arcmin. High-resolution predictions can be downloaded from Dryad
- 973 (https://doi.org/10.5061/dryad.vhhmgqnsh).
- 974

975 Supporting Information

- 976 Additional supporting information can be found online in the Supporting Information section at
- 977 the end of the article.