

CYP2C8 polymorphism among the Portuguese

Isa Cavaco¹, Rita Piedade¹, J. Pedro Gil² and Vera Ribeiro^{1,*}

¹ Laboratory of Molecular Toxicology, Centre for Molecular and Structural Biomedicine (CBME), University of Algarve, Faro, Portugal

² Malaria Research Laboratory, Department of Medicine, Karolinska Institute, Karolinska University Hospital, Stockholm, Sweden

Abstract

Cytochrome P450 2C8 (CYP2C8) is a polymorphic phase I drug-metabolising enzyme involved in the metabolism of a wide variety of xenobiotics, as well as a proposed player in the regulation of vascular tone. Polymorphisms in this gene may have an impact on the metabolism of therapeutic drugs such as paclitaxel and verapamil. In this report we have determined the frequencies of the main non-synonymous CYP2C8 alleles, 805A>T (CYP2C8*2), 416G>A/1196A>G (CYP2C8*3) and 792C>G (CYP2C8*4) in a sample representative of Portuguese Caucasians. The allelic frequencies determined were 1.2%, 19.8%, and 6.4% for CYP2C8*2, CYP2C8*3, and CYP2C8*4, respectively. The observed CYP2C8*3 prevalence is significantly different from the frequencies previously reported in North European populations.

Keywords: allele; CYP2C8; cytochrome P450; polymorphism; Portugal.

Introduction

The human cytochrome P450 2C8 (CYP2C8) enzyme is part of the four-member CYP2C subfamily of CYP450 enzymes. This enzyme is mainly expressed in liver, as well as in various extra-hepatic tissues, such as the vascular smooth muscles (1, 2).

The CYP2C8 gene is located in a cluster on chromosome 10 with the other CYP2C genes (3). As for all the other members of this subfamily, several single nucleotide polymorphisms (SNPs) have been described for CYP2C8 (<http://www.imm.ki.se/CYPalleles/CYP2C8.htm>). Diversity in this gene is apparently lower than that observed for the other CYP2C genes, an idea which has been recently supported by analysis of the full CYP2C8 open reading frame in a set of individuals of very diverse geographical origin (4).

The main CYP2C8 polymorphisms known code for the amino acid changes I269F, R139K, K399R and

I264M. These variants have been documented as being associated with altered enzyme activity in vitro towards the probe drug paclitaxel (5, 6). These SNPs define three main non-wild-type alleles, CYP2C8*2 (harbouring I269F), CYP2C8*3 (carrying both R139K and K399R) and CYP2C8*4 (I264M).

The CYP2C8 enzyme plays an important role in the metabolism of several therapeutic drugs, including paclitaxel (7), amodiaquine (8), troglitazone (9), amiodarone and verapamil (10, 11). Furthermore, it has been suggested that CYP2C8 is involved in the activation of procarcinogenic compounds (12). CYP2C8 also plays a role in the biosynthesis of endogenous vasoregulating factors from arachidonic acid (5). The importance of its endogenous functions might be reflected in the relatively low number of non-synonymous SNPs documented for CYP2C8.

In this work we studied the aforementioned CYP2C8 alleles among the Portuguese. In addition, due to its expected effect on the structure of the protein, we further screened our population for the presence of a recently described three-nucleotide deletion (–/TTG, 461delV) on exon 9 (<http://www.ncbi.nlm.nih.gov/SNP/>, ID: 3832694) (13).

Materials and methods

Blood samples were obtained from 164 unrelated, healthy Caucasian Portuguese from Southern Portugal (67 men and 97 women), recruited from local medical check-ups, upon informed consent. This study was approved by the Ethical Boards of the institutions involved, and followed the Declaration of Helsinki guidelines.

Genomic DNA was extracted from peripheral blood using established phenol-chloroform methods and analysed by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) according to previously described methods to test for the presence of A805T (Ile269Phe), G416A (Arg139Lys), C792G (Ile264Met) and A1196G (Lys399Arg) (5, 6, 14). To detect the deletion in exon 9, the following specific primers were designed: forward, 5'-AAC GAA TTT GTG CAG GAG A-3'; and reverse, 5'-GTA GTA TTG AGG TTC TTT AAA TCG T-3'. The 115-bp PCR product was digested with *HincII* (Fermentas, Vilnius, Lithuania), which recognises the reference sequence (without deletion), resulting in two fragments of 89 and 26 bp.

Allelic frequencies and confidence intervals were assessed using the programme Confidence Interval Analysis (15). χ^2 Testing was performed with GraphPad Software (<http://www.graphpad.com/quickcalcs/index.cfm>). The GenePop software pack was applied for the evaluation of Hardy-Weinberg equilibrium of the SNPs analysed (<http://wbiomed.curtin.edu.au/genepop/>).

Results

The most frequent non-wild-type allele observed was CYP2C8*3, with a frequency of 19.8%. The CYP2C8*2

*Corresponding author: Vera Ribeiro, PhD, Laboratory of Molecular Toxicology, CBME, University of Algarve, Campus de Gambelas, 8005-139, Faro, Portugal
Phone: +351-289800900 ext. 7683, Fax: +351-289819403, E-mail: vmarques@ualg.pt

Table 1 Frequencies observed for *CYP2C8* genotypes in the Portuguese population (n=164).

G416A (R139K)	C792G (I264M)	A805T (I269F)	A1196G (K399R)	n	Frequency (95% CI)
-/-	-/-	-/-	-/-	93	0.567 (0.491–0.643)
-/-	-/-	+/-	-/-	3	0.018 (0.004–0.053)
+/-	-/-	-/-	+/-	43	0.262 (0.195–0.330)
+/+	-/-	-/-	+/+	7	0.043 (0.017–0.086)
-/-	+/-	-/-	-/-	7	0.043 (0.017–0.086)
-/-	+/+	-/-	-/-	3	0.018 (0.004–0.053)
+/-	+/-	-/-	+/-	6	0.037 (0.014–0.078)
+/-	-/-	+/-	+/-	1	0.006 (0.0002–0.034)
+/-	+/+	-/-	+/-	1	0.006 (0.0002–0.034)

and *CYP2C8*4* alleles were found in 1.2% and 6.4% of the subjects, respectively (Table 1). Concerning the frequency of individuals homozygous for the mutated alleles, 4.3% were found for *CYP2C8*3* and 2.4% for *CYP2C8*4*, while no **2/*2* subjects were identified.

The 3-bp deletion previously detected in a large SNP survey in Japan was not found among the Portuguese.

The *CYP2C8*2* and *CYP2C8*3* alleles were found to be in Hardy-Weinberg equilibrium ($p=1$ and $p=0.8$, respectively). However, *CYP2C8*4* genotypic frequencies are not in agreement with those expected in Hardy-Weinberg equilibrium ($p=0.0016$), with an excess of homozygous *G* individuals (4 observed vs. 1.378 expected) being present in our population.

Discussion

The frequencies observed for *CYP2C8*2* and *CYP2C8*4* in the Portuguese population are in line with other observations in Europe, although few studies are available (Figure 1).

As for *CYP2C8*3*, the prevalence of nearly 20% found represents the highest value ever documented for this allele, although it is not significantly different from the data available on populations from Western

Germany (16), Northeast United Kingdom (6) and Central Spain (17). Complete concordance was observed for the frequencies of Arg139Lys and Lys399Arg substitutions, confirming the structure of the *CYP2C8*3* allele, as described by Dai and co-workers (5).

On the other hand, comparison of our results with data from Scandinavian regions – all with *CYP2C8* frequencies below 10% – shows statistically significant differences. This is true when comparing this study with a report in Sweden ($\chi^2=32.206$, $p<0.01$), a survey in a Finnish population ($\chi^2=6.252$, $p<0.05$), and a recent study performed in the relatively isolated Farøe Islands ($\chi^2=34.442$, $p<0.01$) (18–20). These data suggest the presence of particular prevalences, particularly among certain European populations from peripheral regions.

Although not statistically significant, the differences found between our study and that performed in Central Spain points to a certain singularity of the Portuguese, a trend previously noted when analysing other P450 genes, such as *CYP3A4* (21) and *CYP2D6* (22).

As for the non-detection of the 3-bp deletion previously detected in Japan, our result reinforces the awareness of the particular pharmacogenetic characteristics of the Japanese population, related to its ethnic and geographical characteristics.

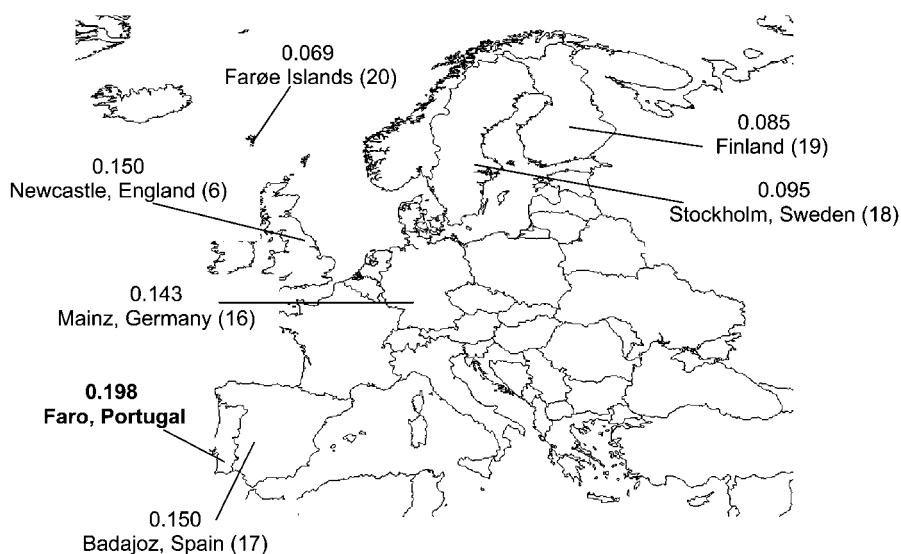


Figure 1 *CYP2C8*3* prevalence in European countries (map modified from <http://geography.about.com/library/blank/blxeurope.htm>). References are given in parentheses.

Finally, compared with the recent first data in native African populations (14), the present study shows significant differences between these and Caucasian subjects for the three alleles analysed (*2, *3 and *4), supporting conclusions from previous comparisons with Afro-American subjects (5).

Possibly related to the endogenous functions of CYP2C8 in the biosynthesis of vasoactive substances (23), the CYP2C8*3 allele has been associated with a higher risk of acute myocardial infarction (AMI) (24). In this context, the high frequency of this allele among the Portuguese is of some concern, as cardiovascular diseases – including AMI – represent the main cause of death in the country after a gradual increase in incidence in recent decades (Ministry of Health, Portugal, <http://www.dgsaude.pt/>).

Acknowledgements

This work was partially supported by Fundação para a Ciência e Tecnologia, Portugal (Praxis Grant SFRH/BD/8887/2002 to IC).

References

- Klose TS, Blaisdell JA, Goldstein JA. Gene structure of CYP2C8 and extrahepatic distribution of the human CYP2Cs. *J Biochem Mol Toxicol* 1999;13:289–95.
- Fleming I. Cytochrome p450 and vascular homeostasis. *Circ Res* 2001;89:753–62.
- Finta C, Zaphiropoulos PG. The human CYP2C locus: a prototype for intergenic and exon repetition splicing events. *Genomics* 2000;63:433–8.
- Solus JF, Arietta BJ, Harris JR, Sexton DP, Steward JQ, McMunn C, et al. Genetic variation in eleven phase I drug metabolism genes in an ethnically diverse population. *Pharmacogenomics* 2004;5:895–931.
- Dai D, Zeldin DC, Blaisdell JA, Chanas B, Coulter SJ, Ghayayem BI, et al. Polymorphisms in human CYP2C8 decrease metabolism of the anticancer drug paclitaxel and arachidonic acid. *Pharmacogenetics* 2001;11:597–607.
- Bahadur N, Leathart JB, Mutch E, Steimel-Crespi D, Dunn SA, Gilissen R, et al. CYP2C8 polymorphisms in Caucasians and their relationship with paclitaxel 6 α -hydroxylase activity in human liver microsomes. *Biochem Pharmacol* 2002;64:1579–89.
- Rahman A, Korzekwa KR, Grogan J, Gonzalez FJ, Harris JW. Selective biotransformation of taxol to 6 α -hydroxytaxol by human cytochrome P450 2C8. *Cancer Res* 1994;54:5543–6.
- Li XQ, Bjorkman A, Andersson TB, Ridderstrom M, Masi-mirembwa CM. Amodiaquine clearance and its metabolism to N-desethylamodiaquine is mediated by CYP2C8: a new high affinity and turnover enzyme-specific probe substrate. *J Pharmacol Exp Ther* 2002;300:399–407.
- Yamazaki H, Shibata A, Suzuki M, Nakajima M, Shimada N, Guengerich FP, et al. Oxidation of troglitazone to a quinone-type metabolite catalyzed by cytochrome P-450 2C8 and P-450 3A4 in human liver microsomes. *Drug Metab Dispos* 1999;27:1260–6.
- Ohyama K, Nakajima M, Nakamura S, Shimada N, Yamazaki H, Yokoi T. A significant role of human cytochrome P450 2C8 in amiodarone N-deethylation: an approach to predict the contribution with relative activity factor. *Drug Metab Dispos* 2000;28:1303–10.
- Borlak J, Walles M, Levsen K, Thum T. Verapamil: metabolism in cultures of primary human coronary arterial endothelial cells. *Drug Metab Dispos* 2003;31:888–91.
- Yun CH, Shimada T, Guengerich FP. Roles of human liver cytochrome P4502C and 3A enzymes in the 3-hydroxylation of benzo(a)pyrene. *Cancer Res* 1992;52:1868–74.
- Haga H, Yamada R, Ohnishi Y, Nakamura Y, Tanaka T. Gene-based SNP discovery as part of the Japanese Millennium Genome Project: identification of 190,562 genetic variations in the human genome. Single-nucleotide polymorphism. *J Hum Genet* 2002;47:605–10.
- Cavaco I, Stromberg-Norklit J, Kaneko A, Msellem MI, Dahoma M, Ribeiro VL, et al. CYP2C8 polymorphism frequencies among malaria patients in Zanzibar. *Eur J Clin Pharmacol* 2005;61:15–8.
- Gardner MJ, Altman DG, editors. *Statistics with confidence – confidence intervals and statistical guidelines*. London: Br Med J 1989.
- Weise A, Grundler S, Zaumsegel D, Klotzek M, Grondahl B, Forst T, et al. Development and evaluation of a rapid and reliable method for cytochrome P450 2C8 genotyping. *Clin Lab* 2004;50:141–8.
- Garcia-Martin E, Martinez C, Tabares B, Frias J, Agundez JA. Interindividual variability in ibuprofen pharmacokinetics is related to interaction of cytochrome P450 2C8 and 2C9 amino acid polymorphisms. *Clin Pharmacol Ther* 2004;76:119–27.
- Yasar U, Lundgren S, Eliasson E, Bennet A, Wiman B, de Faire U, et al. Linkage between the CYP2C8 and CYP2C9 genetic polymorphisms. *Biochem Biophys Res Commun* 2002;299:25–8.
- Niemi M, Backman JT, Kajosaari LI, Leathart JB, Neuvonen M, Daly AK, et al. Polymorphic organic anion transporting polypeptide 1B1 is a major determinant of repaglinide pharmacokinetics. *Clin Pharmacol Ther* 2005;77:468–78.
- Halling J, Petersen MS, Damkier P, Nielsen F, Grandjean P, Weihe P, et al. Polymorphism of CYP2D6, CYP2C19, CYP2C9 and CYP2C8 in the Faroese population. *Eur J Clin Pharmacol* 2005;61:491–7.
- Cavaco I, Gil JP, Gil-Berglund E, Ribeiro V. CYP3A4 and MDR1 alleles in a Portuguese population. *Clin Chem Lab Med* 2003;41:1345–50.
- Gil JP. Polymorphisms of the CYP2D6, GSTM1, GSTP1 and NAT2 in the Portuguese population – search for susceptibility markers in cancer pathologies. Ph.D. Dissertation Thesis, Universidade Nova de Lisboa, Portugal, 2000.
- Fleming I. Cytochrome P450 epoxygenases as EDHF synthase(s). *Pharmacol Res* 2004;49:525–33.
- Yasar U, Bennet AM, Eliasson E, Lundgren S, Wiman B, de Faire U, et al. Allelic variants of cytochrome P450 2C modify the risk for acute myocardial infarction. *Pharmacogenetics* 2003;13:715–20.

Received September 23, 2005, accepted November 1, 2005