

Genome sequence of the marine alphaproteobacterium *Lentilitoribacter* sp. EG35 isolated from the temperate octocoral *Eunicella gazella*

Tina Keller-Costa,^{1,2} Selene Madureira,^{1,2} Ana S. Fernandes,^{1,2} Lydia Kozma,^{1,3} Jorge M.S. Gonçalves,⁴ Cristina Barroso,^{5,6} Conceição Egas,^{5,6} Rodrigo Costa^{1,2}

AUTHOR AFFILIATIONS See affiliation list on p. 3.

ABSTRACT We report the genome sequence of *Lentilitoribacter* sp. strain EG35 isolated from the octocoral *Eunicella gazella* sampled off the coast of Portugal. We reveal the coding potential for the biosynthesis of polyhydroxyalkanoates — biodegradable polyesters that may serve bioplastics production, diverse homoserine lactone-like communication signals, and four putatively novel natural products.

KEYWORDS bacterial evolution, corals, host-microbe interactions, *Rhizobiaceae*, symbiosis

The microbial communities of octocorals are unique in taxonomic composition and presumably benefit their host through nutrient provision and cycling, antioxidant production, and chemical defense (1–3). *Lentilitoribacter* (*Rhizobiaceae*, *Hyphomicrobiales*) is a marine alphaproteobacterial genus currently with only one valid species, *Lentilitoribacter donghaensis* (4) and one RefSeq genome (5, 6) available. It has so far been cultured from seawater (4), marine sponges (5), and octocorals (7), yet little is known about its role in association with marine invertebrates.

We report the genome of *Lentilitoribacter* sp. strain EG35 isolated from a healthy *Eunicella gazella* specimen, sampled by SCUBA diving at 18 m depth in the Atlantic off the coast of Portugal (Pedra da Greta: 36.979778, –7.98911) on 21 April 2021. A microbial cell suspension was retrieved from 1 g of coral soft tissue by mortar-and-pestle homogenization in 9 mL sterile Ca²⁺ and Mg²⁺-free artificial seawater as described earlier (8). Serial dilutions were plated on Marine Agar (MA) and incubated for 7 days at 24°C. Single colonies were streaked until purity on MA plates, and genomic DNA was extracted from a pure culture, freshly grown in Marine Broth using the Wizard Genomic DNA Purification kit (Promega, USA). Strain EG35 was identified by Sanger sequencing of the 16S rRNA gene amplified from genomic DNA using primers F27 (5′-AGAGTTTGATCMTGGCTCAG-3′) and R1492 (5′-TACGGY TACCTGTTACGACTT-3′) and the SILVA/SINA (v1.2.12) database for taxonomic classification. The same DNA sample was used for genome sequencing at GENOINSEQ (Biocant, Cantanhede, Portugal). The genome library was prepared with the Nextera XT DNA Library Preparation Kit, and the genome was paired-end sequenced (average read length, 257/258 bp) on an Illumina MiSeq sequencer with the MiSeq reagent Kit v3 (600 cycles). Default parameters were used for all software unless otherwise specified. Raw reads were imported to KBase (9) and quality-checked using FastQC v0.12.1. Low-quality reads were removed using Trimmomatic v0.36 (10) prior to genome assembly using SPAdes v3.15.3 (11). Contigs below 500 bp were removed. Genome completeness and contamination were assessed with CheckM v1.0.18 (12), and genome taxonomy was confirmed with GTDB-Tk v2.3.2 (13). Contigs were annotated using the NCBI Prokaryotic Genome Annotation Pipeline

Editor Elinne Becket, California State University San Marcos, San Marcos, California, USA

Address correspondence to Rodrigo Costa, rodrigocosta@tecnico.ulisboa.pt, or Tina Keller-Costa, tinakellercosta@tecnico.ulisboa.pt.

The authors declare no conflict of interest.

See the funding table on p. 4.

Received 7 August 2024

Accepted 12 October 2024

Published 24 October 2024

Copyright © 2024 Keller-Costa et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 General features of the *Lentilitoribacter* sp. EG35 genome reported in this study

Genome size (bp)	GC content (%)	Genome coverage (x)	Number of contigs	Contig N50 (bp)	Number of reads	Average Read length (bp)	Completeness (%)	Estimate (%) of:		Number of:	Counts of:			Biosample accession number						
								Contamination	Contamination		Genes ^a RNA ^a rRNA ^a tRNA ^a ncrRNA ^a	COG ^b	Pfams ^b		GenBank accession number	SRA accession number	Bioproject accession number			
4,063,774	44.5	243.4	45	307,952	2 × 1,714,668	257/258	99.92	0.00	3,935 ^a	3,977	42	3	35	4	3,131	7,130	JBFNGQ000	SRR29864	PRJNA11354	SAMN424839
									3,912 ^b								00000	805	83	28

^aAnnotation performed by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP): https://www.ncbi.nlm.nih.gov/genome/annotation_prok/
^bThe Melange pipeline (<https://github.com/sandragodinhosilva/melange>) was used to perform Protein family (Pfam) and Clusters of Orthologous Groups of Proteins (COG)-based annotations of the *Lentilitoribacter* EG35 genome. The corresponding data are available on Zenodo under <https://doi.org/10.5281/zenodo.13856673>

(PGAP v.6.7) (14) and our in-house Melange pipeline (<https://github.com/sandragodinho-silva/melange>). AntiSMASH v7.0 (15) was used to identify secondary metabolite biosynthetic gene clusters (BGCs).

The general features of the genome are shown in Table 1. Strain EG35 shared 98.39% average nucleotide identity (ANI)—calculated with FastANI v0.1.3 (16)—with *Lentilitoribacter* sp. Alg239-R112 ([GCF_900537175.1](https://doi.org/10.1093/mra/11.12.254)), its closest relative with a sequenced genome as determined by phylogenomics inference. Strain Alg239-R112 was isolated from the marine sponge *Spongia* sp (5). at the same sampling location where strain EG35 was retrieved.

The EG35 genome encodes the poly[(R)-3-hydroxyalkanoate] polymerase subunit *phaC* (EC 2.3.1.304) crucial for the biosynthesis of polyhydroxyalkanoates (PHAs), biodegradable polyesters that can be used for bioplastics production (17), and two other genes involved in PHA metabolism, the PHA synthesis regulator *phaR*, and the poly(3-hydroxybutyrate) depolymerase *phaZ* (EC 3.1.1.75). The strain's potential to produce PHAs was confirmed *via* Nile Red staining (18). Strain EG35 harbors six BGCs coding for homoserine lactone signaling molecules, including one sharing 100% similarity to the BGC of kolossin, a peptide likely involved in interspecies communication (19). Moreover, four putatively novel BGCs coding for a terpene, arylpolyene, betalactone, and a ribosomally synthesized and post-translationally modified peptide are present.

ACKNOWLEDGMENTS

This work was financed by the Fundo Azul program of Direção - Geral de Política do Mar (DPGM; Ministry of the Sea, Portugal) through grant FA_05_2017_032, and by the “Blue Bioeconomy Pact” (Project N°. C644915664-00000026), co-funded by Next Generation EU European Fund, under the incentive line “Agendas for Business Innovation” within Funding Scheme 5-Capitalization and Business Innovation of the Portuguese Recovery and Resilience Plan (RRP). Further support was provided by the Portuguese Foundation for Science and Technology (FCT) in the scope of the projects UIDB/04565/2020 and UIDP/04565/2020 of iBB and the project LA/P/0140/2020 of i4HB. ASF is the recipient of a PhD grant conceded by the “Blue Bioeconomy Pact” project. TKC is the recipient of a Research Scientist contract conceded by the FCT (CEECIND/00788/2017). The authors are most grateful to Carlos Afonso and Adela Belackova for their help with coral sampling and Matilde Marques for her help with DNA preparation for genome sequencing.

AUTHOR AFFILIATIONS

¹Institute for Bioengineering and Biosciences and Institute for Health and Bioeconomy (i4HB), Instituto Superior Técnico, Universidade de Lisboa, Lisbon, Portugal

²Department of Bioengineering, Instituto Superior Técnico, Universidade de Lisboa, Lisbon, Portugal

³École Polytechnique Fédérale de Lausanne, Écublens, Switzerland

⁴Centre of Marine Sciences (CCMAR), University of Algarve, Campus de Gambelas, Faro, Portugal

⁵Biocant—Transfer Technology Association, BiocanPark, Cantanhede, Portugal

⁶Center for Neuroscience and Cell Biology (CNC), Rua Larga-Faculdade de Medicina, University of Coimbra, Coimbra, Portugal

AUTHOR ORCIDs

Tina Keller-Costa  <http://orcid.org/0000-0003-3702-9192>

Rodrigo Costa  <http://orcid.org/0000-0002-5932-4101>

FUNDING

Funder	Grant(s)	Author(s)
Direção Geral de Política do MAR (DGPM; Ministry of the Sea, Portugal)	FA_05_2017_032	Rodrigo Costa
Next Generation EU European Fund	C644915664-00000026	Rodrigo Costa

AUTHOR CONTRIBUTIONS

Tina Keller-Costa, Conceptualization, Data curation, Funding acquisition, Project administration, Resources, Supervision, Visualization, Writing – original draft, Writing – review and editing | Selene Madureira, Formal analysis, Investigation, Visualization | Ana S. Fernandes, Formal analysis, Investigation | Lydia Kozma, Formal analysis, Investigation | Jorge M.S. Gonçalves, Funding acquisition, Resources | Cristina Barroso, Data curation, Formal analysis | Conceição Egas, Data curation, Formal analysis, Funding acquisition, Resources | Rodrigo Costa, Conceptualization, Funding acquisition, Project administration, Resources, Supervision, Visualization, Writing – review and editing

DATA AVAILABILITY STATEMENT

The genome sequence of *Lentilitoribacter* sp. strain EG35 has been deposited at DDBJ/ENA/GenBank under the BioProject [PRJNA1135483](#) and the Whole Genome Shotgun accession number [JBFNGQ000000000](#). The antiSMASH and Melange results of BGC, COG and Pfam annotations are available on Zenodo under <https://doi.org/10.5281/zenodo.13856673>. Further details can be found in Table 1.

REFERENCES

- Keller-Costa T, Lago-Lestón A, Saraiva JP, Toscan R, Silva SG, Gonçalves J, Cox CJ, Kyrpides N, Nunes da Rocha U, Costa R. 2021. Metagenomic insights into the taxonomy, function, and dysbiosis of prokaryotic communities in octocorals. *Microbiome* 9:72. <https://doi.org/10.1186/s40168-021-01031-y>
- Keller-Costa T, Kozma L, Silva SG, Toscan R, Gonçalves J, Lago-Lestón A, Kyrpides NC, Nunes da Rocha U, Costa R. 2022. Metagenomics-resolved genomics provides novel insights into chitin turnover, metabolic specialization, and niche partitioning in the octocoral microbiome. *Microbiome* 10:151. <https://doi.org/10.1186/s40168-022-01343-7>
- van de Water JAJM, Allemand D, Ferrier-Pagès C. 2018. Host-microbe interactions in octocoral holobionts - recent advances and perspectives. *Microbiome* 6:64. <https://doi.org/10.1186/s40168-018-0431-6>
- Park S, Lee J-S, Lee K-C, Yoon J-H. 2013. *Lentilitoribacter donghaensis* gen. nov., sp. nov., a slowly-growing alphaproteobacterium isolated from coastal seawater. *Antonie Van Leeuwenhoek* 103:457–464. <https://doi.org/10.1007/s10482-012-9825-9>
- Karimi E, Costa R. 2020. Isolation and genome sequencing of 14 *Spongia* sp. bacterial associates expands the taxonomic and functional breadth of the cultivatable marine sponge microbiome. *Microbiology*. <https://doi.org/10.1101/2020.03.20.000216>
- Almeida JF, Marques M, Oliveira V, Egas C, Mil-Homens D, Viana R, Cleary DFR, Huang YM, Fialho AM, Teixeira MC, Gomes NCM, Costa R, Keller-Costa T. 2023. Marine sponge and octocoral-associated bacteria show versatile secondary metabolite biosynthesis potential and antimicrobial activities against human pathogens. *Mar Drugs* 21:34. <https://doi.org/10.3390/md21010034>
- Kozma L. 2021. Molecular assessment of unculturable and culturable bacterial symbionts of temperate gorgonian corals. In Master's thesis in Life Sciences Engineering. EPFL - École Polytechnique Fédérale de Lausanne, Switzerland.
- Keller-Costa T, Eriksson D, Gonçalves JMS, Gomes NCM, Lago-Lestón A, Costa R. 2017. The gorgonian coral *Eunicella labiata* hosts a distinct prokaryotic consortium amenable to cultivation. *FEMS Microbiol Ecol* 93:1–19. <https://doi.org/10.1093/femsec/fix143>
- Arkin AP, Cottingham RW, Henry CS, Harris NL, Stevens RL, Maslov S, Dehal P, Ware D, Perez F, Canon S, et al. 2018. KBase: the United States department of energy systems biology knowledgebase. *Nat Biotechnol* 36:566–569. <https://doi.org/10.1038/nbt.4163>
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
- Chaumeil P-A, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a toolkit to classify genomes with the genome taxonomy database. *Bioinformatics* 36:1925–1927. <https://doi.org/10.1093/bioinformatics/bt2848>
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
- Blin K, Shaw S, Augustijn HE, Reitz ZL, Biermann F, Alanjary M, Fetter A, Terlouw BR, Metcalf WW, Helfrich EJM, van Wezel GP, Medema MH, Weber T. 2023. antiSMASH 7.0: new and improved predictions for detection, regulation, chemical structures and visualisation. *Nucleic Acids Res* 51:W46–W50. <https://doi.org/10.1093/nar/gkad344>
- Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>
- Muneer F, Rasul I, Azeem F, Siddique MH, Zubair M, Nadeem H. 2020. Microbial polyhydroxyalkanoates (PHAs): efficient replacement of synthetic polymers. *J Polym Environ* 28:2301–2323. <https://doi.org/10.1007/s10924-020-01772-1>

18. Elain A, Le Fellic M, Corre YM, Le Grand A, Le Tilly V, Audic JL, Bruzard S. 2015. Rapid and qualitative fluorescence-based method for the assessment of PHA production in marine bacteria during batch culture. *World J Microbiol Biotechnol* 31:1555–1563. <https://doi.org/10.1007/s11274-015-1904-4>
19. Bode HB, Brachmann AO, Jadhav KB, Seyfarth L, Dauth C, Fuchs SW, Kaiser M, Waterfield NR, Sack H, Heinemann SH, Arndt HD. 2015. Structure elucidation and activity of kolossin A, the D-/L-pentadecapeptide product of A giant nonribosomal peptide synthetase. *Angew Chem Int Ed Engl* 54:10352–10355. <https://doi.org/10.1002/anie.201502835>