

1 A comparative evaluation of biological activities and bioactive compounds of the seagrasses *Zostera*  
2 *marina* and *Z. noltei* from Southern Portugal

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21 Abstract

22 This work assessed the antioxidant potential, acetylcholinesterase (AChE) inhibition and the *in vitro*  
23 cytotoxic activity of extracts of the seagrasses *Zostera marina* and *Z. noltei* collected from southern  
24 Portugal. The total phenolic contents (TPC), the rosmarinic acid (RA) concentration (HPLC/DAD) and  
25 the fatty acid profile (GC/MS) is also described. *Z. marina* had the highest TPC, radical scavenging  
26 activity against DPPH radicals, and copper chelating activity. *Z. noltei* had metal chelation capacity to  
27 copper and iron ions. None of the species was able to inhibit AChE. Both seagrasses had high levels of  
28 polyunsaturated fatty acids. *Z. marina* significantly and selectively reduced the viability of tumorous  
29 neuronal cells. *Z. noltei* was highly toxic for the three cell lines tested and was selective against  
30 hepatocarcinoma cells at the concentration of 100 µg/mL. RA was the main compound identified in *Z.*  
31 *marina*, but not in *Z. noltei*.

32 Keywords: Cytotoxicity; oxidative stress; PUFA; rosmarinic acid, seagrasses

## 33 **1. Introduction**

34 There are four species of seagrasses native to Europe, namely *Zostera marina*, *Z. noltei*, *Cymodocea*  
35 *nodosa* and *Posidonia oceanica*. Water and methanol extracts of *Z. marina* and *Z. noltei* collected from  
36 the French Mediterranean coast displayed algicidal effects on *Alexandrium catenella*, a dinoflagellate  
37 responsible for the occurrence of harmful algal blooms (Laabir et al., 2013). *Z. marina* leaf extracts  
38 have antioxidant and antibiotic activities (Kim et al., 2004, Kolenchenko et al., 2005; Choi et al., 2009),  
39 while organic extracts of *Z. noltei* collected in the Central Mediterranean area had strong antifungal  
40 activity against *Candida albicans* and *Aspergillus niger* (Ballesteros et al., 1992). Some bioactive  
41 compounds have been isolated from *Z. marina*, such as apigenin-7-O- $\beta$ -D-glucoside and chrysoeriol  
42 (Kim et al., 2004). Rosmarinic (RA) and zosteric acids have been detected in both *Zostera* species  
43 (Kolenchenko et al., 2005; Achamlale et al., 2009; Grignon-Dubois et al., 2012). The *Zostera* genus  
44 exhibits a high chemical plasticity and displays different phenolic profiles for different geographic  
45 areas (Grignon-Dubois et al., 2012). However, nothing has been reported so far about the biological  
46 activities and / or chemical components of *Z. marina* and *Z. noltei* from southern Portugal. Therefore,  
47 evaluated the antioxidant, anti-acetylcholinesterase (AChE) and the *in vitro* cytotoxic activities of  
48 extracts of both species, collected in Southern Portugal (Algarve). It also describes some  
49 biocompounds present in both species, including total phenolics content, rosmarinic acid (RA) levels  
50 and fatty acids (FA) profile.

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## 52 **2. Results and discussion**

53 The extraction yields of the crude methanol extracts were 18.3% for *Z. marina* and 9.3% for *Z. noltei*,  
54 while the extraction yields of the ethyl acetate fractions were similar for both species: 0.16 and 0.14%  
55 for *Z. marina* and *Z. noltei*, respectively (Table 1). *Z. marina* had a higher TPC than *Z. noltei* (Table 1).  
56 RA was found in the ethyl acetate extracts for the two species and was the main component in *Z.*  
57 *marina*, but not in *Z. noltei* (Fig S1 and Fig. S2). The different results about RA composition between  
58 the Achamlale et al. (2009) report and our study can be ascribed to the chemical plasticity and different  
59 patterns of phenolic composition depending on the collection site, and / or to the extraction procedure  
60 (Grignon-Dubois and Rezzonico, 2012; Santos-Buelga et al., 2012).

61 The fatty acids (FA) profiles of both species were mainly composed of palmitic, linoleic (LA)  
62 and  $\alpha$ -linolenic (ALA) acids, which represent more than 70% of the total FA detected (Table 2). All

63 results were consistent with those previously reported by other authors (Coelho et al., 2011). The  
64 proportion of polyunsaturated fatty acids (PUFA) in both species was higher than the proportion of  
65 saturated FA and monounsaturated FA combined (Table 2). Moreover, the PUFA found in these  
66 species were mainly from the n-3 series, and although in less amounts, the hexadecatrienoic,  
67 eicosapentaenoic (EPA) and docosahexaenoic (DHA) acids were also detected. The proportion of n-3  
68 PUFA detected in *Z. marina* and *Z. noltei* were approximately 40% of the total FA; therefore, taking  
69 into account their FA profile, both seagrasses can be considered to be a promising source of n-3 PUFA  
70 with valuable nutritional, pharmaceutical and biomedical applications.

71 *Z. marina* had the highest RSA (Table 1), which was lower than the RSA value previously reported  
72 for the ethyl acetate fraction of a crude methanol extract for the same species collected in Korea (Choi  
73 et al., 2009). *Z. marina* also had a good capacity to chelate copper, although not iron (Table 1). The  
74 antioxidant activity detected on the ethyl acetate fraction of *Z. marina* could be ascribed to RA, which  
75 is the most abundant compound in this fraction, with recognized antioxidant activity (Lu and Foo,  
76 2002). *Z. noltei* had capacity to chelate both copper and iron ions (Table 1), but none of the extracts  
77 exhibited capacity to inhibit AChE (data not shown). Nonetheless, they could help to alleviate AD  
78 symptoms by the inhibition of oxidative stress processes and deposition of  $\beta$ -amyloid plaques through  
79 the chelation of copper and iron ions (Weinreb et al., 2011).

80 The *Z. marina* extract was less toxic to HepG2 and S17 cells ( $IC_{50} > 100 \mu\text{g mL}^{-1}$ ), than to the  
81 neuroblastoma cell line ( $IC_{50} = 63.2 \mu\text{g mL}^{-1}$ ), and was able to selectively reduce the viability of this  
82 cell line at the concentrations of 50 and  $100 \mu\text{g mL}^{-1}$  (Fig. 1). Conversely, *Z. noltei* was highly toxic for  
83 the three cell lines tested ( $IC_{50} < 3 \mu\text{g mL}^{-1}$ ) although it was selective ( $SI = 2.1$ ) against HepG2 cells at  
84 the highest concentration tested ( $100 \mu\text{g mL}^{-1}$ ; Fig. 1). In *Z. marina*, the reduction of cell viability can  
85 be attributed to the presence of RA (Stanojković et al., 2013). In *Z. noltei* the cytotoxic activity is likely  
86 attributed to others compounds present in the extracts. Etoposide allowed  $IC_{50}$  values of  $1.9 \mu\text{g mL}^{-1}$   
87 (HepG2 cells),  $38.6 \mu\text{g mL}^{-1}$  (SH-SY5Y) and  $10 \mu\text{g mL}^{-1}$  (S17), and selectivity values of 5.2 for HepG2  
88 cells and 0.2 for the SH-SY5Y cell line.

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### 90 **3. Conclusions**

91 *Z. marina* and *Z. noltei* contain of antioxidant compounds, namely RA. *Z. noltei* could be a source of  
92 potential chemotherapeutic agents and / or valuable lead compounds for the semi-synthesis or total

93 synthesis of effective new drugs. The isolation and identification of the compound(s) responsible for  
94 the bioactivities of *Z. noltei* are now being pursued.

95

## 96 **Supplementary material**

97 Experimental details relating to this article are available online.

98

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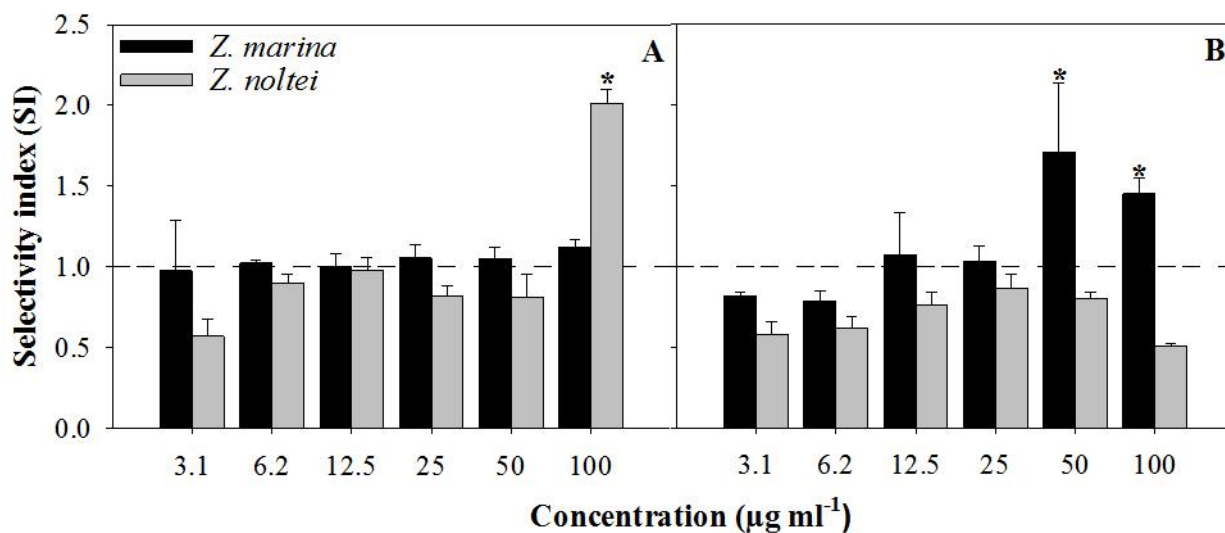
134 **Tables**

135 **Table 1.** Extraction yields (%), total phenolic contents (TPC), radical scavenging (RSA) on DPPH radicals,  
 136 and metal chelation activity on iron (MCAI) and copper (MCAC) of extracts *Z. marina* and *Z. noltei*,  
 137 expressed as IC<sub>50</sub> values (mg/mL)

Samples	Extraction yields (%)		TPC <sup>1</sup>	RA <sup>2</sup>	RSA	MCAI	MCAC
	ME	EAE					
<i>Z. marina</i>	18.3	0.16	0.14 ± 0.01	0.24	0.31 ± 0.01 <sup>b</sup>	> 10	0.61 ± 0.02 <sup>c</sup>
<i>Z. noltei</i>	9.37	0.14	0.09 ± 0.00	0.09	1.10 ± 0.15 <sup>c</sup>	4.99 ± 0.71 <sup>b</sup>	0.88 ± 0.05 <sup>b</sup>
BHT*					0.07 ± 0.00 <sup>a</sup>		
EDTA*						0.08 ± 0.00 <sup>a</sup>	0.05 ± .01 <sup>a</sup>

138 Values are means ± SD (*n* = 6). <sup>1</sup>Expressed as gallic acid equivalents [(mg GAE/g dry weight (DW)]; <sup>2</sup>RA  
 139 content quantified by HPLC analysis of the ethyl acetate fractions (mg/g<sup>-1</sup>, DW); \*Positive control; ME:  
 140 methanol extract; EAE: ethyl acetate extract.

141 **Figures**



142

143 **Figure 1.** Selectivity index (SI) of *Z. marina* and *Z. noltei* extracts against human hepatocarcinoma (A,  
144 HepG2 cells) and neuroblastoma (B, SH-SY5Y) cell lines, calculated in relation to non-tumoural cell  
145 line (S17 cells). Solid bars and errors bars represent the average and standard error values, respectively  
146 ( $n = 9$ ). \*Significantly higher than 1 (dashed line) ( $p < 0.05$ ).